

WORLD SEPSIS DAY 2019

stop
sepsis
save
lives



Foreword by:

Prof. Jean-Louis Vincent

Consultant,
Department of Intensive Care, Erasme
University Hospital, Brussels, Belgium;
Professor of Intensive Care Medicine,
Université Libre de Bruxelles, Brussels,
Belgium;
Past-President of the World Federation
of Societies of Intensive and Critical Care
Medicine

Access the contents and video on the online portal:



<http://collections.medengine.com/critical-care/world-sepsis-day-2019-booklet-2/>

In MDR Gram-negative Bacilli Infections

R_x

POLYFIC

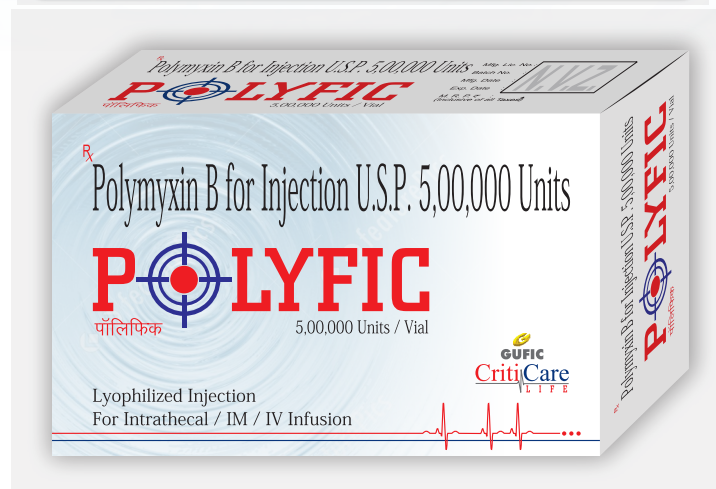
Inj. Polymyxin B Sulphate 5,00,000 Units

Reconsider to treat Gram-negative infections

**Indian manufacturer of
Polymyxin-B**

**API imported from Xellia
pharmaceutical's-Denmark**

Micronized particle size




GUFIC
CritiCare
L I F E

Website: www.gufic.com • Email: info@guficbio.com



WORLD SEPSIS DAY 2019

All rights reserved. No part of this publication may be reproduced, transmitted or stored in any form or by any means either mechanical or electronic, including photocopying, recording or through an information storage and retrieval system, without the written permission of the copyright holder.

Although great care has been taken in compiling the content of this publication, the publisher and its servants are not responsible or in any way liable for the accuracy of the information, for any errors, omissions or inaccuracies, or for any consequences arising therefrom. Inclusion or exclusion of any product does not imply its use is either advocated or rejected. Use of trade names is for product identification only and does not imply endorsement. Opinions expressed do not necessarily reflect the views of the Publisher, Editor/s, Editorial Board or Authors.

Please consult the latest prescribing information from the manufacturer before issuing prescriptions for any products mentioned in this publication. The product advertisements published in this reprint have been provided by the respective pharmaceutical company and the publisher and its servants are not responsible for the accuracy of the information.

© Springer Healthcare 2019

July 2019

 **Springer** Healthcare

This edition is created in India for free distribution in India.

This edition is published by Springer Nature India Private Limited.
Registered Office: 7th Floor, Vijaya Building, 17, Barakhamba Road, New Delhi 110 001, India.
T: +91 (0) 11 4575 5888
www.springerhealthcare.com

Part of the Springer Nature group

Contents

Foreword	vii
Jean-Louis Vincent		

Critical Appraisal

1. Polymyxin B-immobilized hemoperfusion and mortality in critically ill adult patients with sepsis/septic shock: a systematic review with meta-analysis and trial sequential analysis	1
Tomoko Fujii, Riki Ganeko, Yuki Kataoka, <i>et al.</i>		
2. Rationalizing antimicrobial therapy in the ICU: a narrative review	13
Jean-François Timsit, Matteo Bassetti, Olaf Cremer, <i>et al.</i>		

Research Update

3. Cardiovascular clusters in septic shock combining clinical and echocardiographic parameters: a post hoc analysis	31
Guillaume Geri, Philippe Vignon, Alix Aubry, <i>et al.</i>		
4. Fever control in critically ill adults. An individual patient data meta-analysis of randomised controlled trials	42
Paul J. Young, Rinaldo Bellomo, Gordon R. Bernard, <i>et al.</i>		
5. Early PREdiction of sepsis using leukocyte surface biomarkers: the ExPRES-sepsis cohort study	51
Manu Shankar-Hari, Deepankar Datta, Julie Wilson, <i>et al.</i>		

Comprehending Sepsis

6. Non-antiarrhythmic interventions in new onset and paroxysmal sepsis-related atrial fibrillation	64
Antoine Vieillard-Baron, John Boyd		

Expert Comments

7. Using multiple 'omics strategies for novel therapies in sepsis	68
James A. Russell, Peter Spronk, Keith R. Walley		

Step by step procedure for online viewing:

1. Go to <http://collections.medengine.com/critical-care/world-sepsis-day-2019-booklet-2/> or scan QR code.
2. Web page of the issue will open on the screen.
3. View and read the PDF version and watch the videos online.
4. The PDF file can be downloaded too for offline reading.



Foreword

Sepsis is a life-threatening condition associated with high mortality rates and considerable long-term morbidity with sepsis survivors often facing persistent physical or psychological sequelae many years after discharge. Mortality rates in patients who survive sepsis also remain elevated for several years after the initial insult. Early diagnosis, enabling rapid appropriate, and effective management to be started is essential if the global burden of sepsis is to be reduced.

Caused by a dysregulated host response to infection, sepsis can be difficult to diagnose accurately and rapidly. Multiple biomarkers of sepsis have been proposed, but none is specific for sepsis and few are widely available. Once sepsis is diagnosed, the patient should be managed by a sepsis team wherever possible, with treatment focusing on the three key arms: infection control, hemodynamic stabilization, and modulation of the sepsis response. Diagnosis of infection using microbiological cultures is time-consuming, and can delay the start of appropriate antibiotics. Initial antibiotics should therefore, be chosen to cover all possible bacteria according to likely site and source of infection and local microbiological patterns. Once culture results are available, the antibiotic prescription can be altered to reduce risks of unwanted adverse effects from unnecessary antibiotics and limit the development of antibiotic resistance. Newer molecular methods that can identify infection without the need for culture processing are beginning to become available and will help speed the use of appropriate antibiotics. Any infectious source, e.g., infected line or intraperitoneal abscess, must be removed as a matter of urgency.

Hemodynamic status should be stabilized using fluids and vasopressor agents according to the four SOSP stages: Salvage, Optimization, Stabilization, and De-escalation, respectively. Targets for fluid and vasopressor administration should be adapted to individual patient requirements as much as is possible as, for example, a cardiac index or mean arterial pressure that is adequate for one patient may not be suitable for another. Improved techniques to monitor the microcirculation should help identify ongoing need for fluids and/or vasoactive agents but remain experimental at present. Other organ support, e.g., mechanical ventilation and renal replacement therapy should be started as required, when available. International guidelines on sepsis management and hemodynamic support are available, but must be adapted to local conditions and available resources.

No drugs are currently available that modulate the sepsis response, with the possible exception of corticosteroids in patients with severe septic shock. However, many potential agents are being studied. New techniques, including those based on omics technologies, are being developed to help characterize specific clinical and biological phenotypes of individual patients with sepsis, which will help in the identification of effective treatments and in the appropriate targeting of treatment choices in the future.

In this booklet, published in conjunction with the World Sepsis Day 2019 and aimed at advanced critical care intensivists in India, some key published articles about various aspects of the management of sepsis have been reproduced. Written by International experts in the field, these provide an up-date on some of the latest developments in the field of sepsis, and I am sure will be of interest to all those involved in the treatment of patients with sepsis.


Happy reading!

Prof. Jean-Louis Vincent

SYSTEMATIC REVIEW



Polymyxin B-immobilized hemoperfusion and mortality in critically ill adult patients with sepsis/septic shock: a systematic review with meta-analysis and trial sequential analysis

Tomoko Fujii^{1,2} , Riki Ganeko³, Yuki Kataoka⁴, Toshi A. Furukawa⁵, Robin Featherstone⁶, Kent Doi⁷, Jean-Louis Vincent⁸, Daniela Pasero⁹, René Robert¹⁰, Claudio Ronco¹¹ and Sean M. Bagshaw^{12*}

© 2017 Springer-Verlag GmbH Germany, part of Springer Nature and ESICM

Abstract

Purpose: Polymyxin B-immobilized hemoperfusion (PMX-HP) is an adjuvant therapy for sepsis or septic shock that clears circulating endotoxin. Prior trials have shown that PMX-HP improves surrogate endpoints. We aimed to conduct an evidence synthesis to evaluate the efficacy and safety of PMX-HP in critically ill adult patients with sepsis or septic shock.

Methods: We searched for randomized controlled trials (RCTs) in MEDLINE, EMBASE, the Cochrane Library, the Health Technology Assessment Database, CINAHL, “Igaku Chuo Zasshi”, the National Institute of Health Clinical Trials Register, the World Health Organization International Clinical Trials Registry Platform, the University Hospital Medical Information Network Clinical Trials Registry, the reference lists of retrieved articles, and publications by manufacturers of PMX-HP. The primary outcomes were 28-day all-cause mortality, the number of patients with at least one serious adverse event, and organ dysfunction scores. The GRADE methodology for the certainty of evidence was used.

Results: Six trials (857 participants; weighted mean age 62.5 years) proved eligible. Patient-oriented primary outcomes were assessed. The pooled risk ratio (RR) for 28-day mortality associated with PMX-HP was 1.03 [95% confidence interval (CI) 0.78–1.36; $I^2 = 25\%$; $n = 797$]. The pooled RR for adverse events was 2.17 (95% CI 0.68–6.94; $I^2 = 0\%$; $n = 717$). Organ dysfunction scores over 24–72 h after PMX-HP treatment did not change significantly (standardized mean difference -0.26 ; 95% CI -0.64 to 0.12 ; $I^2 = 78\%$; $n = 797$). The certainty of the body of evidence was judged as low for both benefit and harm using the GRADE methodology.

Conclusions: There is currently insufficient evidence to support the routine use of PMX-HP to treat patients with sepsis or septic shock.

Registration: PROSPERO International Prospective Register of Systematic Reviews (CRD42016038356).

Keywords: Sepsis, Septic shock, Polymyxin B-immobilized hemoperfusion, Systematic review, Meta-analysis

*Correspondence: bagshaw@ualberta.ca

¹² Department of Critical Care Medicine, Faculty of Medicine and Dentistry, University of Alberta, 2-124 Clinical Sciences Building, 8440-112 ST NW, Edmonton, Canada T6G 2B7

Full author information is available at the end of the article

Introduction

Sepsis remains desperately fatal and septic shock has a hospital mortality rate as high as 20–50% worldwide [1–5]. Many interventions have been evaluated to improve the prognosis of sepsis, but large multi-centered trials of various therapies have failed to demonstrate consistent benefit [6]. As fundamental elements of sepsis treatment, including timely and appropriate antimicrobial therapies, adequate fluids, and vasopressors, have not changed for decades [7, 8], there currently is dire need for new and effective therapies.

Endotoxin, a principal component of the outer membrane of Gram-negative bacteria, is recognized as a potent mediator of the host response to infection and development of sepsis [9]. Studies measuring endotoxin levels in patients with septic shock have found that high levels of endotoxin activity correlated with worse clinical outcomes [10, 11]. Polymyxin B (PMX) is a cyclic cationic polypeptide antibiotic with high affinity for endotoxin. A novel strategy whereby PMX is bound and immobilized to polystyrene fibers in a hemoperfusion device was developed in Japan [12, 13]. The suggested mechanism of PMX hemoperfusion (PMX-HP) is to remove circulating endotoxin by adsorption, which modulates and limits the maladaptive host response to infection and the progression of the organ injury cascade of sepsis.

Selected clinical trials have suggested PMX-HP can improve the physiological profile of patients with sepsis [14–16]; however, it remains uncertain whether PMX-HP can reproducibly improve patient outcomes, as the trials have largely focused on surrogate endpoints or have been underpowered to detect effects on clinically important outcomes [16]. Additional studies have recently been completed evaluating PMX-HP, including two large multi-center randomized controlled trials (RCTs) [17, 18].

We therefore conducted an up-to-date systematic review and evidence synthesis evaluating the impact of PMX-HP as an adjuvant therapy for critically ill adult patients with sepsis or septic shock on clinical outcomes and health services utilization. We hypothesized that use of PMX-HP would improve survival among adult critically ill patients with sepsis or septic shock.

Methods

Protocol and registration

This systematic review was conducted using guidelines in the Cochrane Collaboration and Centre for Reviews and Dissemination and reported according to the Preferred Reporting Items for Systematic Reviews and Meta-Analyses guideline [19]. The protocol has been registered with the PROSPERO International

Prospective Register of Systematic Reviews; registration number CRD42016038356, and published in full elsewhere [20].

Eligibility criteria

All relevant RCTs that investigated the effect of PMX-HP for patients with sepsis or septic shock were included. The primary research question was “what is the efficacy, effectiveness and potential harm of PMX-HP compared with standard therapy?” We obtained all relevant studies irrespective of language or publication status. Adults aged 18 years or older with sepsis, severe sepsis or septic shock were included. The diagnosis of sepsis was based on clinically suspected or documented systemic infection with any signs of systemic inflammatory response syndrome. Septic shock was classically defined as hypotension resistant to fluid administration and requiring norepinephrine or other vasopressors [21]. The intervention was use of the PMX-HP for the adjuvant treatment of sepsis or septic shock. The comparison was standard treatment only or sham hemoperfusion. Primary outcomes were 28-day all-cause mortality, the number of patients with at least one serious adverse event, and organ dysfunction scores [22] over 24–72 h after the treatment. Secondary outcomes included 90-day all-cause mortality, mean arterial blood pressure over 24–72 h after the treatment, endotoxin levels over 24–72 h after the treatment, duration of vasopressor therapy or vasopressor-free days, the receipt of renal replacement therapy (RRT), costs related to health services, and total mortality defined as mortality at 28 days or any follow-up duration when available.

Information sources

The search strategy was developed in collaboration with an experienced health research librarian. We searched MEDLINE (from the inception to Oct 2017), EMBASE, the Cochrane Library, the Health Technology Assessment Database, Cumulative Index to Nursing and Allied Health Literature, Pubmed, and “Igaku Chuo Zasshi” of the Japan Medical Abstract Society (from the inception to June 2016). The search strategies for MEDLINE were developed and were modified for searching all the other databases (eMethod 1). The search strategies were further peer-reviewed by a second research librarian [23]. For ongoing trials, we searched the National Institute of Health Clinical Trials Register, the World Health Organization International Clinical Trials Registry Platform, and the University Hospital Medical Information Network Clinical Trials Registry. We also searched citations from all included studies. We contacted experts in the field of critical care nephrology and selected commercial entities

that develop or license PMX-HP to identify additional unpublished and/or on-going trials.

Study selection

Two authors independently screened titles and abstracts of all trial reports we identified by the search to code them as 'retrieve' (eligible or potentially eligible/unclear) or 'do not retrieve'. The full texts of reports classified as 'retrieve' were reviewed independently according to predetermined eligibility criteria. Discrepancies were resolved through discussion with a third reviewer, as required. We identified and excluded duplicates of the same study.

Data collection process

Two reviewers independently extracted data using standardized and piloted data extraction sheets. We abstracted the following information: study characteristics, patient characteristics, sample size, interventions, comparators, potential biases in the conduct of the trial, outcomes, methods of statistical analysis, and funding support. Agreement between the two reviewers concerning the primary outcome and the risk of bias for the primary outcome was reported as percentage agreement with an intra-class correlation coefficient, and percentage agreement with a weighted kappa, respectively.

Assessment of risk of bias in individual studies

Two reviewers independently assessed the risk of bias of the included studies using the tool described in the Cochrane Handbook for Systematic Reviews of Interventions [24], which consists of eight domains (eTable 1). The risk of bias assessment was done at the outcome level for the primary outcomes. When the original reports provided insufficient details, we made direct inquiry of the study authors. When the assessors disagreed, the final rating was decided through discussion or with the involvement of another member of the review group, if necessary. The key domain of risk of bias for 28-day mortality was allocation concealment. The overall risk of bias was also summarized in further subgroup analyses. More details of assessment of risk of bias is provided in the protocol [20].

Summary measures

As the measure of treatment effect for dichotomous outcomes, we used the risk ratio (RR) and its 95% confidence interval (CI). Continuous outcomes were pooled by calculating the mean difference (MD) with a 95% CI except for organ dysfunction scores. As the data for the organ dysfunction scores were available in the sepsis-related organ failure assessment (SOFA) score [25] or multiple organ dysfunction score [26] (MODS), we pooled

standardized mean differences (SMDs) with a 95% CI [27].

Synthesis of results

We analyzed data from the included studies using Review Manager [28]. The proportion of treatment failure was calculated according to the intention-to-treat (ITT) principle. All randomized patients for whom outcome data were not available were assumed as no events. The effect of imputation was explored by a sensitivity analysis. Given the clinical heterogeneity including variability in the etiologies of sepsis in the population of interest, we used a random-effects model in all analyses [29]. We assessed overall heterogeneity by visual inspection of the forest plots, and statistical heterogeneity using the I^2 statistic and Chi-squared test. I^2 values above 50% were considered to represent substantial statistical heterogeneity. To assess reporting bias, we constructed funnel plots, and visual inspection was performed to investigate the asymmetry. Certainty of the body of evidence was assessed using the grading of recommendations assessment, development and evaluation (GRADE) framework [30]. The GRADE framework characterizes the certainty of a body of evidence on the basis of study limitations, imprecision, inconsistency, indirectness, and other considerations. The starting point for certainty in each estimate is high, but is downgraded according to the assessments of these five domains if there are serious concerns. When the effect estimates were affected substantially by the risk of bias of included studies, then we downgraded the certainty of the evidence in a domain of risk of bias.

Additional analyses

To test the robustness of the effect estimates of PMX-HP, and to explain heterogeneity, we used sensitivity analyses and subgroup analyses. We planned the following sensitivity analyses for 28-day mortality: (1) risk of bias; we included only trials with low risk of bias in allocation concealment; (2) imputed missing data; we imputed missing data on 28-day mortality in two ways: assuming the missing outcomes as events (death) in the PMX-HP group, and as no event in the control group (worst-case scenario); and assuming the missing outcomes as no event in the PMX-HP group, and as event in the control group (best-case scenario); (3) per protocol; and (4) statistical method; we used a fixed-effect model. We performed a priori subgroup analyses for the participant group and the intervention if sufficient detail was present in the eligible studies with the following hypotheses: (1) participants with abdominal sepsis, culture-confirmed sepsis, gram-negative infections, surgery, acute kidney injury (AKI), or septic shock will show greater treatment effect than patients without those conditions; and (2)

greater dose of intervention (i.e., longer duration; more than one treatment) will show greater treatment effect. eMethod 2 explains changes from the protocol [20].

Post hoc analyses

Trial sequential analysis (TSA) was done with a diversity-adjusted information size calculated using a two-sided α of 0.05, a power of 80%, an anticipated relative risk reduction of 20.0%, and a control event rate of 35.0%. TSA viewer version 0.9.5.10 Beta (Copenhagen Trial Unit, Centre for Clinical Intervention Research, Copenhagen, DE. 2016) was used. An additional sensitivity analysis including zero total event studies using continuity correction was done using R version 3.4.1 (R Foundation for Statistical Computing, Vienna, Austria. 2017). We added post hoc subgroup analysis for the overall risk of bias with a different criterion of assessment in the included studies, and the maximum time window from the onset of sepsis/septic shock or surgery to the first therapy.

Results

Of the 1700 citations identified from electronic and hand searches, 12 reports were identified for the review, and after exclusion of ongoing trials or inadequate reports (eTable 2), we included 6 unique trials [14, 15, 17, 18, 31, 32] in the meta-analysis (Fig. 1). The agreement of eligibility between the two reviewers was 90% [Cohen's weighted kappa: 0.79 (95% CI 0.62–0.97)]. Table 1 shows the characteristics of the trials included in the meta-analysis. All the trials used a PMX B-immobilized hemoperfusion device (Toraymyxin 20R). The number of participants across trials ranged between 16 and 450. The weighted mean age of study participants was 62.5 years (range 56.0–69.7). Sixty-one percent were male. Agreement for the primary outcome and the risk of bias items between the two reviewers was 100% (intra-class correlation coefficient: 1).

Primary outcomes

For 401 patients involved in the 5 studies [14, 15, 17, 18, 31] and contributing to 28-day mortality data (representing 83% of the included participants), the pooled RR was 1.03 (95% CI 0.78–1.36; $I^2 = 25\%$; $n = 797$; Table 2, Fig. 2a). All five trials were adjudicated as low risk of bias for the outcome (eTable 3a). The number of patients with at least one serious adverse event was reported in three studies [14, 17, 18]. The pooled RR was 2.17 (95% CI 0.68–6.94; $I^2 = 0\%$; $n = 717$; Fig. 2b). Cruz et al. [15] reported only device-related adverse events in the PMX-HP group (eTable 4). Five studies [14, 15, 17, 18, 31] reported either organ dysfunction scores at 24–72 h after the treatment or their changes over 24–72 h after

the treatment. Only the EUPHRATES trial [18] reported MODS, and the others [14, 15, 17, 31] reported SOFA score. For 797 patients in the 5 studies [14, 15, 17, 18, 31], the SMD for the organ dysfunction scores was -0.26 (95% CI -0.64 to 0.12 ; Fig. 2c). The heterogeneity between the 5 trials was high ($I^2 = 78\%$).

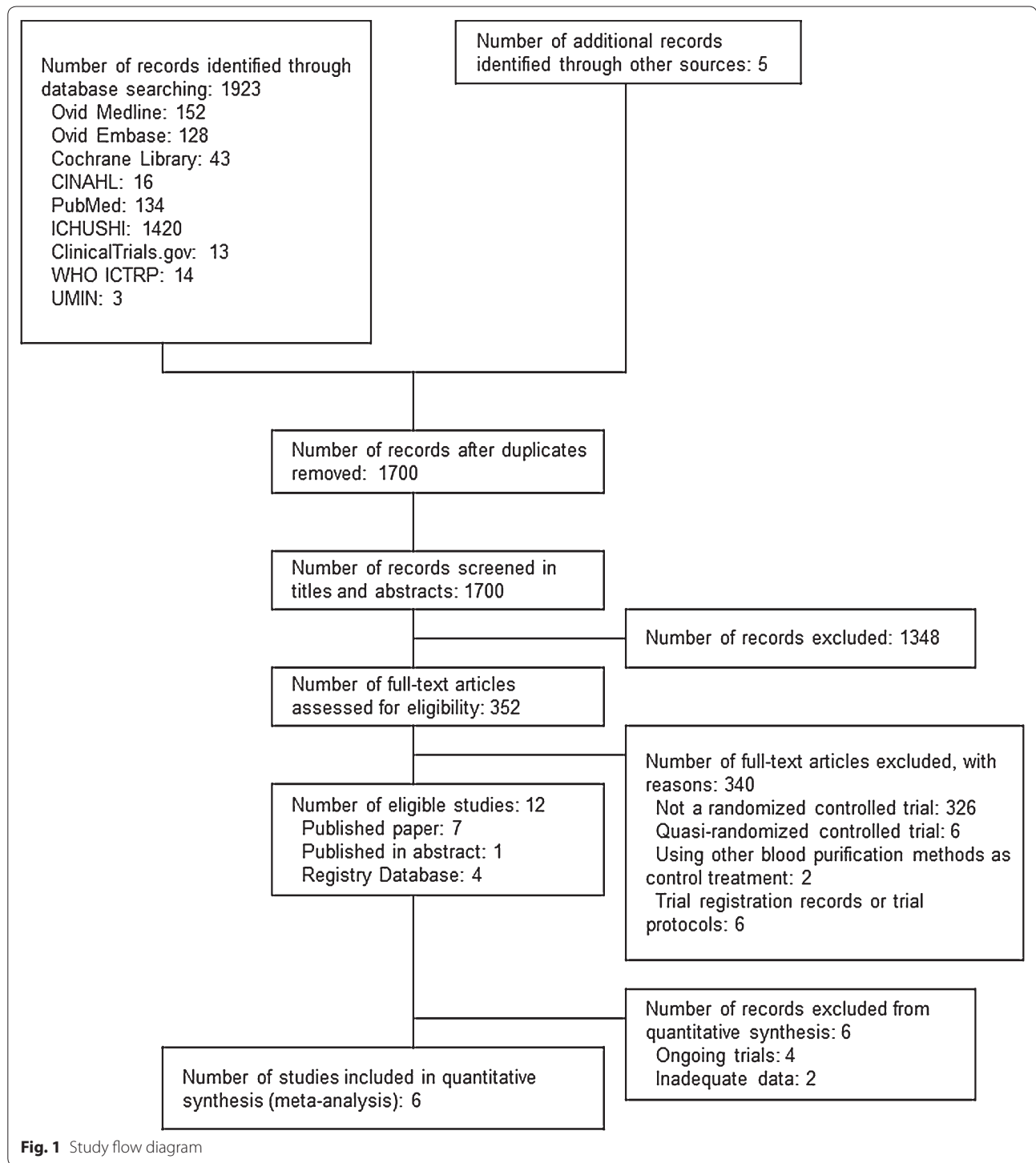
Secondary outcomes

Among 232 patients in the ABDO-MIX trial [17] that reported 90-day mortality, PMX-HP did not reduce 90-day mortality (RR, 1.41; 95% CI 0.93–2.13; Table 2). Four trials [14, 15, 18, 31] reported mean arterial pressure. Pooled results found a statistically significant relationship between PMX-HP and increase in mean arterial pressure (MD, 5.23; 95% CI 2.75–7.72; $I^2 = 0\%$; $n = 565$; Table 2, eFigure 1). Three trials [14, 31, 32] involving 109 patients reported endotoxin levels (pg/mL) over 24–72 h after treatment measured by limulus amoebocyte lysate assay (MD, -40.77 ; 95% CI -118.53 to 36.99 ; $I^2 = 96\%$; Table 2, eFigure 2). There were no differences in vasopressor-free days at 28 days ($n = 283$, 3 trials, eFigure 3), ICU length of stay ($n = 347$, 4 trials, eFigure 4), or the receipt of RRT ($n = 565$, 4 trials, eFigure 5; Table 2). Two studies [15, 18] provided data for duration of RRT (5.2 days for PMX-HP vs. 5.6 days for standard; $p = 0.03$) [15] and RRT-free days to day 28 (14.7 vs. 15.0 days; $p = 0.81$) [18].

One study [33] performed an economic analysis using data collected in the EUPHAS trial involving 64 patients [15] and suggested PMX-HP was cost-effective. They adopted the Italian healthcare provider's perspective and showed a mean incremental cost-effective ratio of EUR 2558 per incremental undiscounted life-year gained and EUR 3864 per incremental discounted life-year gained [33]. For 856 patients involved in the 6 studies [14, 15, 17, 18, 31, 32] contributing mortality data at 28 days or any follow-up duration, the pooled RR for death among patients treated with PMX-HP was 0.85 (95% CI 0.58–1.26; $I^2 = 64\%$; Table 2, eFigure 6).

Sensitivity and subgroup analyses

None of the 5 studies contributing to 28-day mortality [14, 15, 17, 18, 31] was rated at high risk of bias in allocation concealment. Sensitivity analyses with imputation of missing data with the worst-case scenario (pooled RR, 1.05; 95% CI 0.74–1.50; $I^2 = 47\%$, eFigure 7a) and with the best-case scenario (pooled RR, 1.02; 95% CI 0.84–1.24; $I^2 = 0\%$, eFigure 7b), and sensitivity analysis using a fixed-effect model (pooled RR, 1.07; 95% CI 0.88–1.31; $I^2 = 25\%$, eFigure 8), and per-protocol mortality (pooled RR, 0.89; 95% CI 0.62–1.29; $I^2 = 46\%$, eFigure 9) attested to the robustness of the primary analysis.



Subgroup analyses for 5 studies reporting 28-day mortality [14, 15, 17, 18, 31] by trial participants with different sepsis etiologies (abdominal only vs. various etiologies including abdominal), trial participants with sepsis confirmed by culture (culture-confirmed vs. mixed

or not confirmed), trial participants with gram-negative infections (culture-confirmed vs. others), trial participants with surgery (surgical vs. mixed or medical), or severity of trial participants (septic shock only vs. sepsis or septic shock) did not show any subgroup interaction

Table 1 Characteristics of the trials included in the meta-analysis

Source	Country	Study cites	Funding	Total no. of patients	Exclusion from ITT analysis	Age, mean (SD), years	Sex, male, female, <i>n</i>	Patient status	Duration, no. of sessions	Primary outcome
Nakamura 2003 [32]	Japan	NA	Investigator-initiated	60	0	56 (NA)	40, 20	Culture-positive sepsis	2 h, twice	Unclear ^b
Vincent 2005 [14]	Six countries in Europe	Multicenter	Industry-sponsored	35	0	57.7 (15.6) ^a	22, 13	Abdominal sepsis, surgical	2 h, once	The SOFA score
Cantaluppi 2008 [31]	Italy	Two centers	Investigator-initiated	16	0	60 (11.3)	12, 4	Confirmed gram-negative sepsis	2 h, twice	Viability of renal cell cultures
Cruz 2009 [15], Berto 2011 [33]	Italy	Multicenter	Industry-sponsored	64	0	63.8 (14.2) ^a	42, 22	Abdominal sepsis, surgical	2 h, twice	MAP and vasopressor requirement
Payen 2015 [17]	France	Multicenter	Industry-sponsored	243	11	69.7 (11.6)	134, 98	Abdominal sepsis, surgical, septic shock	2 h, twice	28-day mortality
EUPHRATES 2017 [18], Klein 2014 [34]	USA, Canada	Multicenter	Industry-sponsored	450	0	59.8 (14.9) ^a	273, 177 ^a	Septic shock, high endotoxin activity assay	2 h, twice	28-day mortality

ITT Intention to treat, SD standard deviation, NA not available, SOFA sequential organ failure assessment, MAP mean arterial pressure

^a Data provided by the study authors^b Reported endotoxin levels after treatment, survival at unknown follow-up period, adverse events

Table 2 Outcome measures

	Studies	Study reference no.	PMX-HP	Standard	Effect estimate (95% CI)	<i>I</i> ² (%)
Primary outcomes						
28-day mortality	5 ^a	14, 15, 17, 18, 31	135/402	124/395	Pooled RR, 1.03 (0.78, 1.36)	25
Number of patients with at least one serious adverse event	3 ^a	14, 17, 18	8/360	3/357	Pooled RR, 2.17 (0.68, 6.94)	0
Change in organ dysfunction scores over 24–72 h after treatment	5 ^a	14, 15, 17, 18, 31			SMD, − 0.26 (− 0.64, 0.12)	78
Secondary outcomes						
90-day all-cause mortality	1	17	40/119	27/113	RR, 1.41 (0.93, 2.13)	NA
Change in mean arterial blood pressure over 24–72 h after the treatment	4 ^a	14, 15, 18, 31			MD, 5.23 (2.75, 7.72)	0
Endotoxin levels measured by LAL assay over 24–72 h after the treatment	3 ^a	14, 31, 32			MD, − 40.77 (− 118.53, 36.99)	96
28-day vasopressor-free days	3 ^a	14, 17, 31			MD, − 1.10 (− 4.05, 1.85)	10
ICU length of stay	4 ^a	14, 15, 17, 31			MD, − 1.95 (− 7.91, 4.00)	70
The need for RRT	4 ^a	14, 15, 18, 31			Pooled RR, 0.76 (0.33, 1.71)	61
Mortality at 28 days or any follow-up duration	6 ^a	14, 15, 17, 18, 31, 32	144/436	140/420	Pooled RR, 0.85 (0.58, 1.26)	64

CI Confidence interval, RR risk ratio, NA not available, SMD standardized mean difference, MD mean difference, ICU intensive care unit

^a Includes data provided from the study authors

(Table 3, Fig. 2a, eFigure 10–13). The subgroup of patients with AKI was not reported for their 28-day mortality in any of the included studies.

Certainty of evidence

The visual inspection of the funnel plot for the 28-day mortality suggested no apparent publication bias, but there were few studies to assess for asymmetry (eFigure 14). The certainty of evidence for the three primary outcomes was downgraded by one level each for risk of bias and imprecision, and were all considered low (Table 4).

Post hoc analyses

TSA showed the adjusted CI for 28-day mortality was 0.58–1.82 ($I^2 = 25\%$; $n = 797$). The required information size to show a relative riskreduction (RRR) of 20% was 2744 (eFigure 15), and to show 2-point reduction of organ dysfunction scores was 895. An additional analysis including zero total event studies using continuity correction showed the pooled RR of the number of patients with at least one serious adverse event as 2.03 (95% CI 0.67–6.17; $I^2 = 0\%$; $n = 733$). Post hoc subgroup analyses did not show subgroup interaction for the efficacy and safety outcomes. Assessment of overall risk of bias with different criteria did not affect our results (eFigures 16–18).

Discussion

Summary of key findings

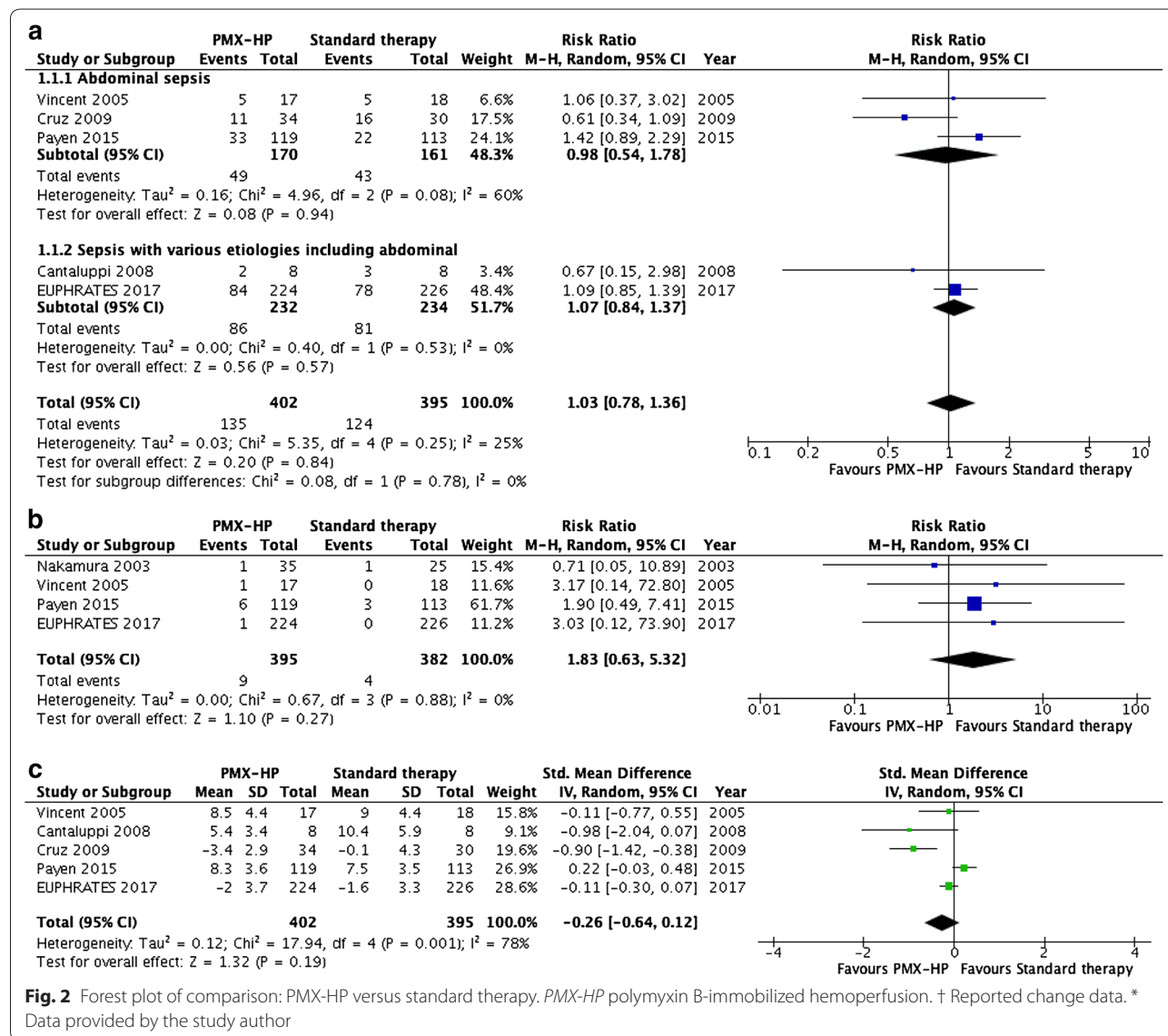
The current systematic review showed that PMX-HP did not reduce 28-day mortality or organ dysfunction scores

of adult sepsis or septic shock patients, and did not appear to significantly increase the risk of adverse events. PMX-HP did not reduce 90-day mortality, or significantly reduce the utilization of health resources.

Context with prior literature

Early experimental or clinical studies evaluating blood purification in sepsis have largely focused on methods of hemofiltration [35–37]. Meanwhile, large multi-centered clinical trials have found intensity of RRT beyond conventionally recommended doses does not improve survival of patients with AKI and sepsis [37–41]. Moreover, the early application of continuous venovenous hemofiltration (CVVH) was implied to worsen the severity of organ dysfunction in severe sepsis [41]. These observations suggest that alternative strategies to better target blood purification and improved survival in sepsis are necessary.

Previous systematic reviews implied that the use of PMX-HP was associated with a survival benefit, improvements in hemodynamics, and reduction in circulating endotoxin levels [16, 42, 43]. In our up-to-date systematic review, we excluded six studies where randomization was not rigorous (e.g., in five studies, allocation was alternating; and in one study, allocation was performed through discussion with patients) following direct inquiry with the study authors [44–49]. Five out of the six excluded studies [45–49] that showed large beneficial effect with relatively small sample sizes were included in previous systematic reviews [16, 42, 43]. We included two newly completed high-profile randomized trials [17, 18], and found no apparent benefit on survival.



There might be several possible explanations for our findings. First, there may be no beneficial effect of using PMX-HP in patients with sepsis or septic shock. Vincent et al. [14] and a post hoc analysis of ABDO-MIX study [50] measured cytokines after completion of PMX-HP treatment, including TNF- α , interleukins, and IFN- γ , and no significant differences were found [50]. These results imply PMX-HP treatment may not significantly remove endotoxin and/or suppress inflammatory cytokines sufficiently to modify the course of organ dysfunction and risk of death.

Second, the pooled analysis with 797 patients may still lack sufficient statistical power to detect small but clinically meaningful effects of PMX-HP treatment, as shown in the TSA. If we assume an absolute risk reduction of 15% (i.e., 43% RRR) with an estimated baseline mortality

of 35%, as adopted in the EUPHRATES trial [34], we have already accumulated sufficient information to conclude a null effect. Kaukonen et al. showed mortality in patients with severe sepsis has declined considerably over the last decade [1], likely in part implying substantial temporal progress in the overarching care provided to critically ill patients. The effect of PMX-HP, if any, could be heterogeneous and much smaller than expected, possibly due largely to PMX-HP being only an adjuvant therapy in the context of multiple interventions used to manage adult critically ill patients with sepsis.

Third, patient selection and case mix may have influenced the expected outcomes. Biologically targeted therapy is sensible to enrich the trial population with patients most likely to derive benefit from the intervention; however, the EUPHRATES trial, the only study that adopted

Table 3 Subgroup analyses of 28-day mortality related to polymyxin B-immobilized hemoperfusion

Subgroup	Studies	Patients	Pooled risk ratio	I^2 (%)	p value
Participants: culture	5	797		0	0.57
Culture-positive sepsis	1 ^a	16	0.67 [0.15, 2.98]		
Not confirmed	4 ^a	781	1.04 [0.76, 1.42]		
Participants: gram-negative infection	5	797		0	0.57
Confirmed gram-negative infection	1 ^a	16	0.67 [0.15, 2.98]		
Not confirmed gram-negative	4 ^a	781	1.04 [0.76, 1.42]		
Participants: surgical	5	797		0	0.78
Surgical	3	331	0.98 [0.54, 1.78]		
Mixed or medical	2 ^a	466	1.07 [0.84, 1.37]		
Participants: severity of sepsis	5	797		72	0.06
Septic shock	2 ^a	682	1.15 [0.92, 1.43]		
Sepsis or septic shock	3 ^a	115	0.69 [0.42, 1.12]		
Intervention: no. of sessions	5	797		0	0.93
Single session	1	35	1.06 [0.37, 3.02]		
Two sessions	4 ^a	762	1.01 [0.72, 1.42]		
More than two sessions	0	0	NA		

NA Not available

^a Includes data provided from the study authors

endotoxin activity as an eligibility criterion, may still have been too small to detect a small but clinically important difference. There are three ongoing trials measuring endotoxin activity at the inclusion of the trials (eTable 2); however, the sample sizes of these trials are also likely to be too small to likely detect a clinically important effect of PMX-HP or to change the overall conclusions of sequential meta-analyses.

Strengths and limitations

We have conducted a rigorous peer-reviewed literature search to identify relevant randomized trials, including a database in Japan where the PMX-HP filter was developed. Furthermore, we have directly contacted all the study authors to assess the eligibility and the quality of each trial to minimize bias in our effect estimation. The inclusion of two new and larger studies [17, 18] has empowered the pooled analysis and enabled the up to date evidence synthesis. We have also performed several predefined sensitivity analyses to confirm the robustness of the findings. However, there are several limitations for this review. Limited numbers of studies did not allow detailed analysis and interpretation to address the issue of heterogeneity in the case mix and in treatment effect in response to PMX-HP. Second, as we conducted meta-analysis with aggregated data, we could not classify participants involved in the studies into complementary subgroups at each patient level. Third, we observed considerable heterogeneity in several analyses. Practice variation across the included studies may have contributed to

heterogeneity. Similarly, there may be a biological basis for responsiveness to PMX-HP among a heterogeneous population of patients with sepsis that is incompletely understood. Fourth, as we have performed multiple analyses in this systematic review, we recommend caution when interpreting significant findings, such as the modest increase of mean arterial pressure. Finally, we have made some changes from our original protocol, due largely to the availability of the data.

Implications for clinicians, policy, and future research

Our review would suggest there is no definitive evidence to support the routine use of PMX-HP for adult critically ill patients with sepsis or septic shock. While there was no significant difference in risk shown in our review, the potential risk of serious adverse events with use of PMX-HP should be considered. The available evidence did not prove its efficacy for improved survival, and as such, performing an economic evaluation may not be justified.

The imprecision of the results does not preclude further trials to assess the efficacy of PMX-HP. In the EUPHRA-TES trial, post hoc exploratory per-protocol analyses showed a beneficial effect among adult patients with a MODS greater than 9 [18], a finding warranting further verification. Future clinical trials should aim to explore specific patient populations with adequate sample size, for example, those with elevated blood endotoxin level, or high organ dysfunction scores, if any clinical effect of PMX-HP is to be detected.

Table 4 Evidence table of the systematic review

Quality assessment			No. of patients					Effect		Certainty	
No. of studies	Study design	Risk of bias	Inconsistency	Indirectness	Imprecision	Other considerations	PMX-HP	Standard therapy	Relative (95% CI)		Absolute (95% CI)
28-day mortality											
5	Randomized trials	Serious ^a	Not serious	Not serious	Serious ^b	None	135/402 (33.6%)	124/395 (31.4%)	RR 1.03 (0.78–1.36)	9 more per 1000 (from 69 fewer to 113 more)	⊕⊕○○ LOW
The number of patients with at least one serious adverse event											
3	Randomized trials	Serious ^a	Not serious	Not serious	Serious ^b	None	8/360 (2.2%)	3/357 (0.8%)	RR 2.17 (0.68–6.94)	10 more per 1000 (from 3 fewer to 50 more)	⊕⊕○○ LOW
Organ dysfunction score over 24–72 h after treatment											
5	Randomized trials	Serious ^a	Not serious	Not serious	Serious ^b	None	402	395	–	SMD 0.26 lower (0.64 lower to 0.12 higher)	⊕⊕○○ LOW

CI Confidence interval, RR risk ratio, SMD standardized mean difference

^a One trial was at high risk of bias in the blinding and sponsorship domains^b Because of the wide confidence interval

Conclusions

Among adult patients with sepsis or septic shock, use of PMX-HP compared with standard therapy alone was not proven to reduce 28-day mortality or to reduce organ dysfunction scores, or significantly increase the risk of serious adverse events. Considering the certainty of the body of evidence was low for both benefit and harm, to date, there is no strong evidence to support the routine use of PMX-HP as an adjuvant therapy in critically ill adult patients with sepsis or septic shock.

Electronic supplementary material

The online version of this article (<https://doi.org/10.1007/s00134-017-5004-9>) contains supplementary material, which is available to authorized users.

Author details

¹ Department of Epidemiology and Preventive Medicine, Kyoto University Graduate School of Medicine, Yoshida Hon-machi, Sakyo-ku, Kyoto, Japan. ² Japan Society for the Promotion of Science, 5-3-1 Kojimachi, Chiyoda-ku, Tokyo, Japan. ³ Department of Surgery, Kyoto University Hospital, 54 Kawaharacho, Syogoin, Sakyo-ku, Kyoto, Japan. ⁴ Department of Respiratory Medicine, Hyogo Prefectural Amagasaki General Medical Center, Higashi-Naniwa-Cho 2-17-77, Amagasaki, Hyogo, Japan. ⁵ Department of Health Promotion and Human Behavior, Kyoto University Graduate School of Medicine, Yoshida Konoe-cho, Sakyo-Ku, Kyoto, Japan. ⁶ Department of Pediatrics, Alberta Research Center for Health Evidence (ARCHE), University of Alberta, 11405-87 Avenue Edmonton, Alberta, Canada. ⁷ Department of Emergency and Critical Care Medicine, The University of Tokyo Hospital, 7-3-1 Hongo, Bunkyo-ku, Tokyo, Japan. ⁸ Department of Intensive Care, Erasme University Hospital, Université Libre de Bruxelles, Route de Lennik 808, 1070 Brussels, Belgium. ⁹ Department of Anesthesia and Intensive Care Medicine, San Giovanni Battista Hospital, University of Turin, Via Giuseppe Verdi, 8, 10124 Turin, Italy. ¹⁰ Service de Réanimation Médicale, CHU-Poitiers, University of Poitiers, 86000 Poitiers, France. ¹¹ Department of Nephrology Dialysis and Transplantation, International Renal Research Institute of Vicenza (IRRV), San Bortolo Hospital, Viale Rodolfi 37, 36100 Vicenza, Italy. ¹² Department of Critical Care Medicine, Faculty of Medicine and Dentistry, University of Alberta, 2-124 Clinical Sciences Building, 8440-112 ST NW, Edmonton, Canada T6G 2B7.

Acknowledgements

We thank Mr. Toshiyuki Suwa at Osaka University for his assistance with the search strategy design in Japanese databases, Ms. Tara Landry at Montreal General Hospital, McGill University Health Centre, for her peer review of the MEDLINE search strategy, Prof. Vladimir Saenko at Nagasaki University for helping to review trial reports in Russian, Ms. Ayumi Horie at Hyogo Prefectural Amagasaki General Medical Center for obtaining the Japanese full-text articles, Toray Industries, Inc. for providing the list of published/unpublished trials and conference proceedings, assisting to get contact with an author of the included study, and all the corresponding persons of ongoing trials for providing information of the current status of the trials. No specific compensation was provided to these individuals.

Author contributions

Drs. TF and SMB had full access to all the data in the study and take responsibility for the integrity of the data and the accuracy of the data analysis. Study concept and design: TF, SMB, RG, YK, RF, and TAF. Acquisition of data: TF, SMW, KD, RG, YK, RF, JV, DP, RR, and CR. Analysis and interpretation of data: TF, SMB, YK, TAF, JV, DP, RR, and CR. Drafting of the manuscript: TF, TAF, SMB, and KD. Critical revision of the manuscript for important intellectual content: RF, RG, YK, JV, DP, RR, CR. All authors gave final approval of the version to be published and agreed to be accountable for all aspects of this work.

Compliance with ethical standards

Funding

TF is supported by the Japan Society for the Promotion of Science (JSPS) and has received a grant from JSPS. This study was funded in part by Grant-in-Aid

for JSPS Research Fellow JP16J10320 to TF. SMB is supported by a Canada Research Chair in Critical Care Nephrology and is a steering committee member of the EUPHRATES trial. He has consulted and received speaking fees from Baxter Healthcare Corp. TAF has received lecture fees from Eli Lilly, Janssen, Meiji, Mitsubishi-Tanabe, MSD, and Pfizer. He has received royalties from Igaku-Shoin and Nihon Bunka Kagaku-sha publishers. He has received research support from Mitsubishi-Tanabe and Mochida. KD received lecture fees from Asahi Kasei Medical Corp, Baxter Healthcare Corp., and Toray Medical Corp. JLV, DP, RB, and CR were engaged in the clinical trials included in this review, and provided some of the data not reported in their published papers. None of these funding organizations have contributed to the study design; collection, management, analysis, and interpretation of data; writing of the report, or the decision to submit the report for publication. Toray Industries provided some of the data included in this review, as noted in the body of the article.

Conflicts of interest

The authors declare that they have no conflicts of interest.

Received: 30 September 2017 Accepted: 22 November 2017

Published online: 4 December 2017

References

- Kaukonen KM, Bailey M, Suzuki S, Pilcher D, Bellomo R (2014) Mortality related to severe sepsis and septic shock among critically ill patients in Australia and New Zealand, 2000–2012. *JAMA* 311:1308–1316
- Rhee C, Dantes R, Epstein L et al (2017) CDC prevention epicenter program: benchmarking the incidence and mortality of severe sepsis in the United States. *JAMA*. <https://doi.org/10.1001/jama.2017.13836> (Epub ahead of print)
- Quenot JP, Binquet C, Kara F et al (2013) The epidemiology of septic shock in French intensive care units: the prospective multicenter cohort EPISS study. *Crit Care* 17:R65
- Leligdowicz A, Dodek PM, Norena M, Co-operative Antimicrobial Therapy of Septic Shock Database Research Group et al (2014) Association between source of infection and hospital mortality in patients who have septic shock. *Am J Respir Crit Care Med* 189:1204–1213
- Singer M, Deutschman CS, Seymour CW et al (2016) The third international consensus definitions for sepsis and septic shock (Sepsis-3). *JAMA* 315:801–810
- Suffredini AF, Munford RS (2011) Novel therapies for septic shock over the past 4 decades. *JAMA* 306:194–199
- Dellinger RP, Carlet JM, Masur H, Surviving Sepsis Campaign Management Guidelines Committee et al (2004) Surviving Sepsis Campaign guidelines for management of severe sepsis and septic shock. *Crit Care Med* 32:858–873
- Dellinger RP, Levy MM, Rhodes A, Surviving Sepsis Campaign Guidelines Committee including the Pediatric Subgroup et al (2013) Surviving sepsis campaign: international guidelines for management of severe sepsis and septic shock: 2012. *Crit Care Med* 41:580–637
- Danner RL, Elin RJ, Hosseini JM, Wesley RA, Reilly JM, Parillo JE (1991) Endotoxemia in human septic shock. *Chest* 99:169–175
- Casey LC, Balk RA, Bone RC (1993) Plasma cytokine and endotoxin levels correlate with survival in patients with the sepsis syndrome. *Ann Intern Med* 119:771–778
- Marshall JC, Foster D, Vincent JL, MEDIC study et al (2004) Diagnostic and prognostic implications of endotoxemia in critical illness: results of the MEDIC study. *J Infect Dis* 190:527–534
- Shoji H, Tani T, Hanasawa K, Kodama M (1998) Extracorporeal endotoxin removal by polymyxin B immobilized fiber cartridge: designing and antiendotoxin efficacy in the clinical application. *Ther Apher* 2:3–12
- Shoji H (2003) Extracorporeal endotoxin removal for the treatment of sepsis: endotoxin adsorption cartridge (Toraymyxin). *Ther Apher Dial* 7:108–114
- Vincent JL, Laterre PF, Cohen J et al (2005) A pilot-controlled study of a polymyxin B-immobilized hemoperfusion cartridge in patients with severe sepsis secondary to intra-abdominal infection. *Shock* 23:400–405

15. Cruz DN, Antonelli M, Fumagalli R et al (2009) Early use of polymyxin B hemoperfusion in abdominal septic shock: the EUPHAS randomized controlled trial. *JAMA* 301:2445–2452
16. Cruz DN, Perazella MA, Bellomo R et al (2007) Effectiveness of polymyxin B-immobilized fiber column in sepsis: a systematic review. *Crit Care* 11:R47
17. Payen DM, Guilhot J, Launey Y, ABDOMIX Group et al (2015) Early use of polymyxin B hemoperfusion in patients with septic shock due to peritonitis: a multicenter randomized control trial. *Intensive Care Med* 41:975–984
18. Dellinger P (2016) EUPHRATES: evaluating the use of polymyxin B hemoperfusion in a randomized controlled trial of adults treated for endotoxemia and septic shock. *Canadian Critical Care Forum*, Toronto
19. Moher D, Liberati A, Tetzlaff J, Altman DG, PRISMA Group (2009) Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. *PLoS Med* 6:e1000097
20. Fujii T, Ganeko R, Kataoka Y, Featherstone R, Bagshaw S, Furukawa TA (2016) Polymyxin B-immobilised haemoperfusion and mortality in critically ill patients with sepsis/septic shock: a protocol for a systematic review and meta-analysis. *BMJ Open* 6:e012908
21. Levy MM, Fink MP, Marshall JC, International Sepsis Definitions Conference et al (2003) 2001 SCCM/ESICM/ACCP/ATS/SIS international sepsis definitions conference. *Intensive Care Med* 29:530–538
22. Vincent JL, de Mendonça A, Cantraine F et al (1998) Use of the SOFA score to assess the incidence of organ dysfunction/failure in intensive care units: results of a multicenter, prospective study. Working group on "sepsis-related problems" of the European Society of Intensive Care Medicine. *Crit Care Med* 26:1793–1800
23. Sampson M, McGowan J, Cogob E, Grimshaw J, Moher D, Lefebvre C (2009) An evidence-based practice guideline for the peer review of electronic search strategies. *J Clin Epidemiol* 62:944–952
24. Higgins JPT, Green S (2008) *Cochrane handbook for Systematic Reviews of Interventions*. John Wiley & Sons, Chichester (UK)
25. Vincent JL, Moreno R, Takala J et al (1996) The SOFA (Sepsis-related Organ Failure Assessment) score to describe organ dysfunction/failure. On behalf of the working group on Sepsis-Related Problems of the European Society of Intensive Care Medicine. *Intensive Care Med* 22:707–710
26. Cook R, Cook D, Tilley J, Lee K, Marshall J, Canadian Critical Care Trials Group (2001) Multiple organ dysfunction: baseline and serial component scores. *Crit Care Med* 29:2046–2050
27. da Costa BR, Nüesch E, Rutjes AW et al (2013) Combining follow-up and change data is valid in meta-analyses of continuous outcomes: a meta-epidemiological study. *J Clin Epidemiol* 66:847–855
28. Review Manager (RevMan) [Computer program] (2014) Version 5.3. The Nordic Cochrane Centre, The Cochrane Collaboration, Copenhagen
29. DerSimonian R, Laird N (1986) Meta-analysis in clinical trials. *Control Clin Trials* 7:177–188
30. Guyatt G, Oxman AD, Akl EA et al (2011) GRADE guidelines: 1. Introduction-GRADE evidence profiles and summary of findings tables. *J Clin Epidemiol* 64:383–394
31. Cantaluppi V, Assenzio B, Pasero D et al (2008) Polymyxin-B hemoperfusion inactivates circulating proapoptotic factors. *Intensive Care Med* 34:1638–1645
32. Nakamura T, Ushiyama C, Suzuki Y et al (2003) Combination therapy with polymyxin B-immobilized fibre haemoperfusion and teicoplanin for sepsis due to methicillin-resistant *Staphylococcus aureus*. *J Hosp Infect* 53:58–63
33. Berto P, Ronco C, Cruz D, Melotti RM, Antonelli M (2011) Cost-effectiveness analysis of polymyxin-B immobilized fiber column and conventional medical therapy in the management of abdominal septic shock in Italy. *Blood Purif* 32:331–340
34. Klein DJ, Foster D, Schorr CA, Kazempour K, Walker PM, Dellinger RP (2014) The EUPHRATES trial (evaluating the use of polymyxin B hemoperfusion in a randomized controlled trial of adults treated for endotoxemia and septic shock): study protocol for a randomized controlled trial. *Trials* 15:218
35. Honore PM, Jamez J, Wauthier M et al (2000) Prospective evaluation of short-term, high-volume isovolemic hemofiltration on the hemodynamic course and outcome in patients with intractable circulatory failure resulting from septic shock. *Crit Care Med* 28:3581–3587
36. Bellomo R, Kellum JA, Gandhi CR et al (2000) The effect of intensive plasma water exchange by hemofiltration on hemodynamics and soluble mediators in canine endotoxemia. *Am J Respir Crit Care Med* 161:1429–1436
37. Cornejo R, Downey P, Castro R et al (2006) High-volume hemofiltration as salvage therapy in severe hyperdynamic septic shock. *Intensive Care Med* 32:713–722
38. VA/NIH Acute Renal Failure Trial Network (2008) Intensity of renal support in critically ill patients with acute kidney injury. *N Engl J Med* 359:7–20
39. RENAL Replacement Therapy Study Investigators (2009) Intensity of continuous renal-replacement therapy in critically ill patients. *N Engl J Med* 361:1627–1638
40. Van Wert R, Friedrich JO, Scales DC et al (2010) High-dose renal replacement therapy for acute kidney injury: systematic review and meta-analysis. *Crit Care Med* 38:1360–1369
41. Payen D, Mateo J, Cavaillon JM et al (2009) Impact of continuous veno-venous hemofiltration on organ failure during the early phase of severe sepsis: a randomized controlled trial. *Crit Care Med* 37:803–810
42. Zhou F, Peng Z, Murugan R, Kellum JA (2013) Blood purification and mortality in sepsis: a meta-analysis of randomized trials. *Crit Care Med* 41:2209–2220
43. Chang T, Tu YK, Lee CT et al (2017) Effects of polymyxin b hemoperfusion on mortality in patients with severe sepsis and septic shock: a systemic review, meta-analysis update, and disease severity subgroup meta-analysis. *Crit Care Med* 45:e858–e864
44. Nemoto H, Nakamoto H, Okada H et al (2001) Newly developed immobilized polymyxin B fibers improve the survival of patients with sepsis. *Blood Purif* 19:361–368
45. Nakamura T, Kawagoe Y, Matsuda T, Koide H (2004) Effect of polymyxin B-immobilized fiber on bone resorption in patients with sepsis. *Intensive Care Med* 30:1838–1841
46. Nakamura T, Ushiyama C, Suzuki S et al (2000) Effect of polymyxin B-immobilized fiber hemoperfusion on sepsis-induced rhabdomyolysis with acute renal failure. *Nephron* 86:210
47. Nakamura T, Ushiyama C, Shoji H, Koide H (2002) Effects of hemoperfusion on serum cardiac troponin T concentrations using polymyxin B-immobilized fibers in septic patients undergoing hemodialysis. *ASAIO J* 48:41–44
48. Nakamura T, Ushiyama C, Suzuki Y, Shoji H, Shimada N, Koide H (2002) Hemoperfusion with polymyxin B immobilized fibers for urinary albumin excretion in septic patients with trauma. *ASAIO J* 48:244–248
49. Nakamura T, Ushiyama C, Suzuki Y et al (2003) Hemoperfusion with polymyxin B-immobilized fiber in septic patients with methicillin-resistant *Staphylococcus aureus*-associated glomerulonephritis. *Nephron* 94:c33–c39
50. Coudroy R, Payen D, Launey Y, ABDOMIX Group et al (2017) Modulation by polymyxin-B hemoperfusion of inflammatory response related to severe peritonitis. *Shock*. 47:93–99

REVIEW

Rationalizing antimicrobial therapy in the ICU: a narrative review

Jean-François Timsit^{1,2*} , Matteo Bassetti³, Olaf Cremer⁴, George Daikos⁵, Jan de Waele⁶, Andre Kallil⁷, Eric Kipnis⁸, Marin Kollef⁹, Kevin Laupland¹⁰, Jose-Artur Paiva¹¹, Jesús Rodríguez-Baño¹², Étienne Ruppé^{2,13}, Jorge Salluh¹⁴, Fabio Silvio Taccone¹⁵, Emmanuel Weiss^{16,17} and François Barbier¹⁸

© 2019 Springer-Verlag GmbH Germany, part of Springer Nature

Abstract

The massive consumption of antibiotics in the ICU is responsible for substantial ecological side effects that promote the dissemination of multidrug-resistant bacteria (MDRB) in this environment. Strikingly, up to half of ICU patients receiving empirical antibiotic therapy have no definitively confirmed infection, while de-escalation and shortened treatment duration are insufficiently considered in those with documented sepsis, highlighting the potential benefit of implementing antibiotic stewardship programs (ASP) and other quality improvement initiatives. The objective of this narrative review is to summarize the available evidence, emerging options, and unsolved controversies for the optimization of antibiotic therapy in the ICU. Published data notably support the need for better identification of patients at risk of MDRB infection, more accurate diagnostic tools enabling a rule-in/rule-out approach for bacterial sepsis, an individualized reasoning for the selection of single-drug or combination empirical regimen, the use of adequate dosing and administration schemes to ensure the attainment of pharmacokinetics/pharmacodynamics targets, concomitant source control when appropriate, and a systematic reappraisal of initial therapy in an attempt to minimize collateral damage on commensal ecosystems through de-escalation and treatment-shortening whenever conceivable. This narrative review also aims at compiling arguments for the elaboration of actionable ASP in the ICU, including improved patient outcomes and a reduction in antibiotic-related selection pressure that may help to control the dissemination of MDRB in this healthcare setting.

Keywords: Antibiotic stewardship, Antimicrobial resistance, Empirical therapy, Critical illness, Carbapenem, Outcome, Sepsis

Introduction

Antibiotics are massively used in ICUs around the world [1]. While the adequacy and the early implementation of empirical coverage are pivotal to cure patients with community- and hospital-acquired sepsis, antimicrobial therapy is not always targeted and, in more than one out of two cases, may be prescribed in patients without

confirmed infections. Moreover, antibiotic de-escalation is insufficiently considered. The resulting selection pressure together with the incomplete control of cross-colonization with multidrug-resistant bacteria (MDRB) makes the ICU an important determinant of the spread of these pathogens in the hospital. As instrumental contributors of antimicrobial stewardship programs (ASP), intensivists should be on the leading edge of conception, optimization, and promotion of therapeutic schemes for severe infections and sepsis, including the limitation of antimicrobial overuse.

In this narrative review based on a literature search (MEDLINE database) completed in September 2018,

*Correspondence: jean-francois.timsit@aphp.fr

¹ Medical and Infectious Diseases ICU, APHP, Bichat-Claude Bernard Hospital, 46 Rue Henri-Huchard, 75877 Paris Cedex 18, France
Full author information is available at the end of the article

we sought to summarize recent advances and emerging perspectives for the optimization of antibiotic therapy in the ICU, notably better identification of patients at risk of MDRB infection, more accurate diagnostic tools enabling a rule-in/rule-out approach for bacterial sepsis, an individualized reasoning for the selection of single-drug or combination empirical regimen, the use of adequate dosing and administration schemes to ensure the attainment of pharmacokinetics/pharmacodynamics targets, concomitant source control when appropriate, and a systematic reappraisal of initial therapy in an attempt to minimize collateral damage on commensal ecosystems through de-escalation and treatment-shortening whenever conceivable. We also aimed to compile arguments for the elaboration of actionable ASP in the ICU, including improved patient outcomes and a reduction in antibiotic-related selection pressure that may help to control the dissemination of MDRB in this healthcare setting.

How antimicrobial therapy influences bacterial resistance

The burden of infections due to extended-spectrum beta-lactamase-producing *Enterobacteriaceae* (ESBL-E) and MDR *Pseudomonas aeruginosa* is rising steadily, carbapenem-resistant *Acinetobacter baumannii* and carbapenemase-producing *Enterobacteriaceae* (CRE) are spreading globally, while methicillin-resistant *Staphylococcus aureus* (MRSA) and vancomycin-resistant enterococci generate major issues in several geographical areas [2–14]. These trends now apply for both ICU-acquired infections and imported bacterial sepsis as a result of the successful dissemination of MDRB in hospital wards and other healthcare environments (Fig. 1).

Up to 70% of ICU patients receive empirical or definite antimicrobial therapy on a given day [1]. The average volume of antibiotic consumption in this population has been recently estimated as 1563 defined daily doses (DDD) per 1000 patient-days (95% confidence interval 1472–1653)—that is, almost three times higher than in ward patients, with marked disparities for broad-spectrum agents such as third-generation cephalosporins [15]. Whilst most of the underlying mechanisms ensue from a succession of sporadic genetic events that are not directly induced by antibiotics, the selection pressure exerted by these drugs stands as a potent driver of bacterial resistance (Fig. 2) [16, 17].

At the patient level, antimicrobial exposure allows the overgrowth of pathogens with intrinsic or acquired resistance to the administered drug within commensal ecosystems or, to a lesser extent, at the site of infection. Of note, some mechanisms may confer resistance

Take-home message

This narrative review summarizes the available evidence, emerging options, and unsolved controversies for the optimization of antibiotic therapy in the ICU. The potential benefit of antibiotic stewardship programs to improve patient outcomes and reduce the ecological side effects of these drugs is also discussed.

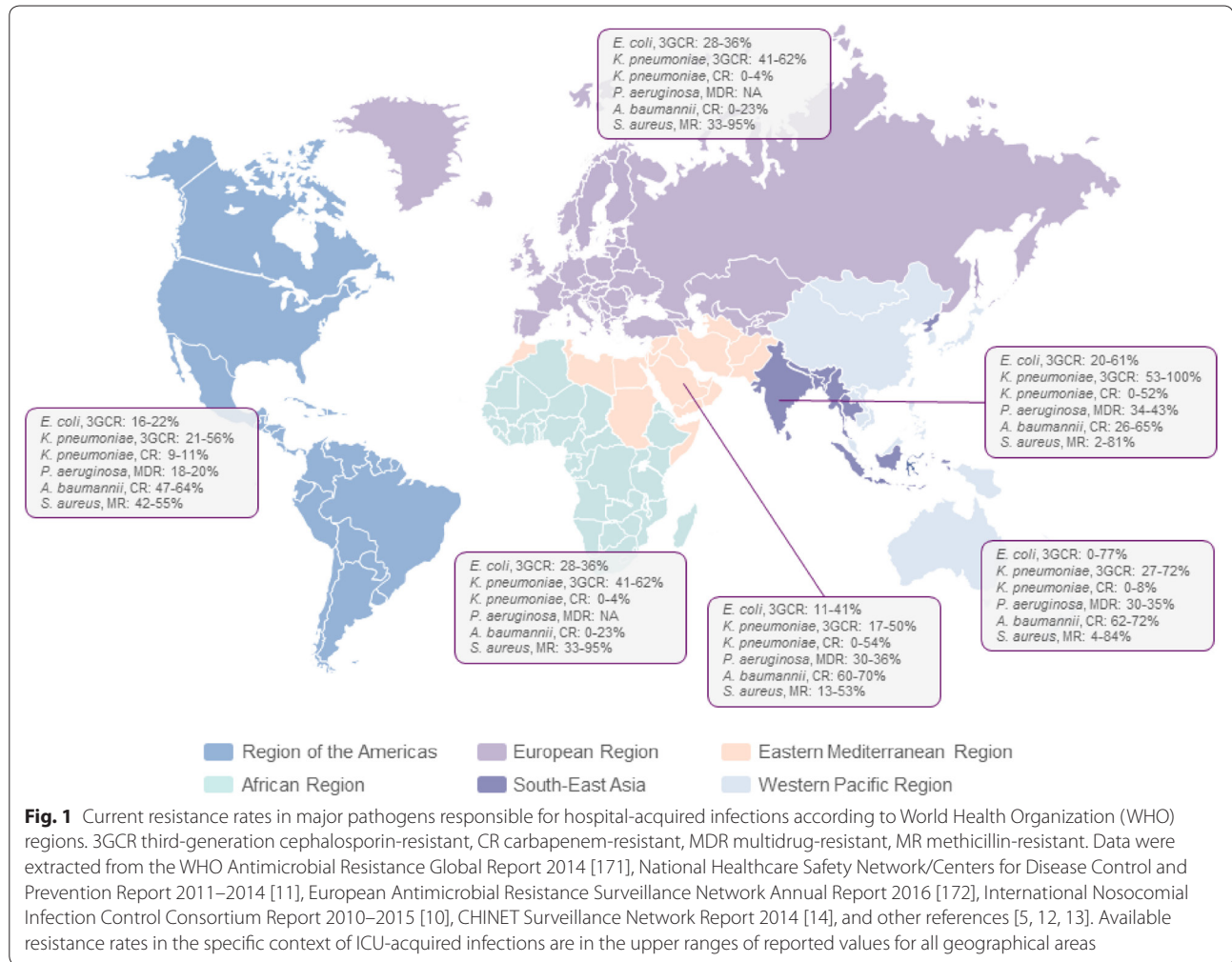
to various classes, notably the overexpression of efflux pumps in non-fermenting Gram-negative bacteria, thereby resulting in the selection of MDR mutants following only a single-drug exposure [18]. At the ICU scale, consumption volumes of a given class correlate with resistance rates in clinical isolates, including for carbapenems or polymyxins [19–26], although this may fluctuate depending on bacterial species and settings [27, 28].

Yet, in addition to its clinical spectrum, anti-anaerobic properties should be considered when appraising the ecological impact of each antibiotic [29]. Indeed, acquisition of MDR Gram-negative bacteria through in situ selection, cross-transmission, or environmental reservoirs may be eased by antimicrobial-related alterations of the normal gut microbiota—primarily resident anaerobes—and the colonization resistance that it confers [30]. A prior course of anti-anaerobic drugs may notably predispose to colonization with ESBL-E [31], AmpC-hyperproducing *Enterobacteriaceae* [32], or CRE [33, 34]. The degree of biliary excretion of the drug appears as another key factor to appraise its potential impact on intestinal commensals [35–37].

Risk factors for multidrug-resistant pathogens

The clinical value of identifying risk factors for MDRB infection is to guide empirical therapy before the availability of culture results—that is, pathogen identification and antimicrobial susceptibility testing (AST). However, no single algorithm may be used to predict a MDRB infection given the complex interplay between the host, the environment, and the pathogen, thus requiring an individualized probabilistic approach for the selection of empirical drugs (Table 1).

Colonization markedly amplifies the risk of subsequent infection with a given MDRB. However, the positive predictive value of this risk factor never exceeds 50% whatever the colonizer is [2, 38–40]. For instance, ESBL-E infections occur during the ICU stay in only 10–25% of ESBL-E carriers [41]. Whether an MDRB carrier becomes infected is related to a further series of factors that may or not be related to those associated with the risk of acquired colonization [2, 38, 39]. Overall, the presence or absence of documented carriage should not



be considered as the unique requisite for the choice of empirical therapy.

Patients with advanced co-morbid illnesses, prolonged hospital stays, use of invasive procedures, and prior antibiotic exposure are at increased risk of MDRB infections [42, 43].

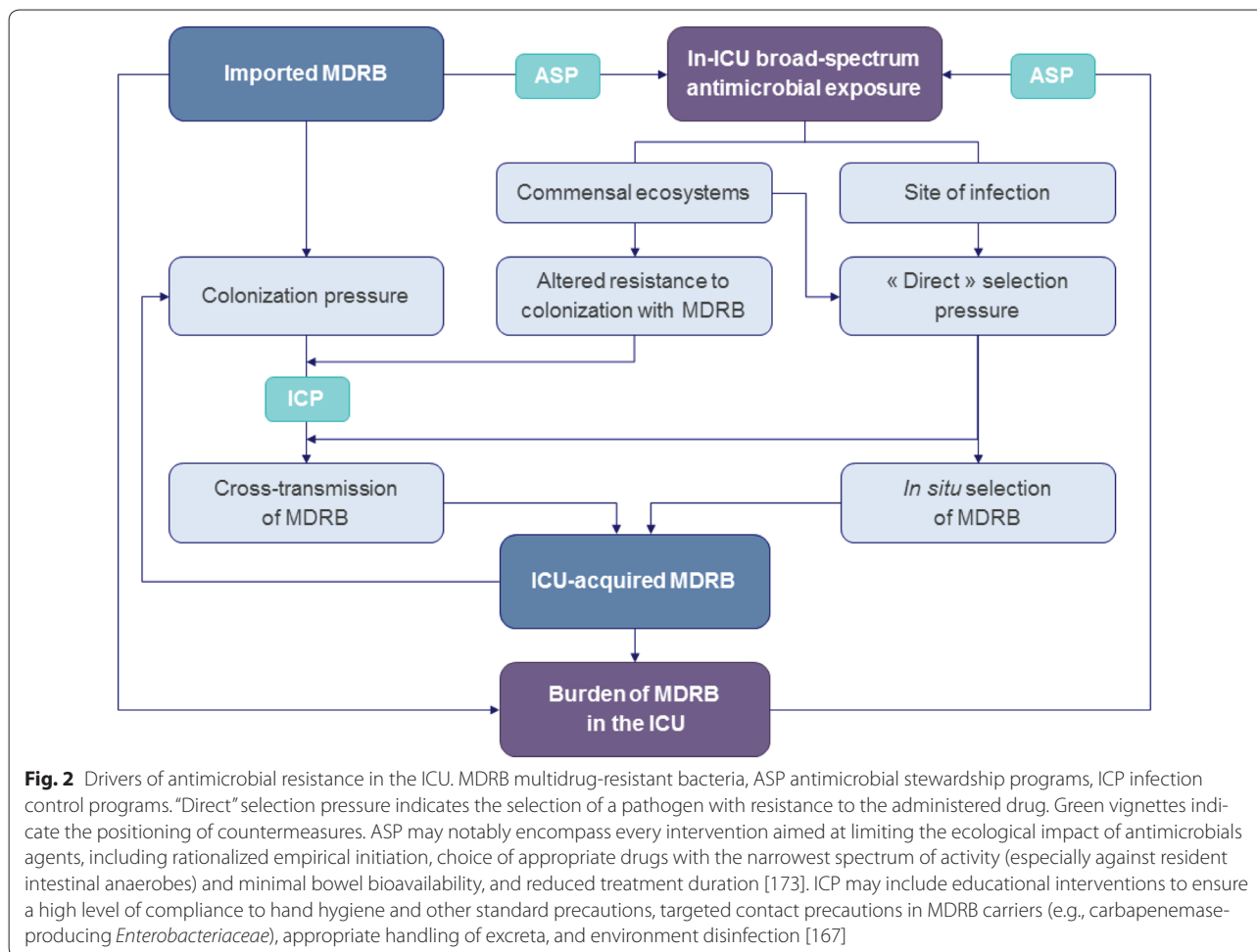
The patient location is another determinant of risk as there are vast differences in the epidemiology of MDRB globally, regionally, and even within hospitals in the same city [2, 44]. Reasons for these discrepancies may include socioeconomic factors as well as variations in case-mix, antimicrobial consumption, and hygiene practices.

When not to start antimicrobials in the ICU

Although mixed [45], the available evidence supports a beneficial effect of prompt antibiotic administration on survival rates in sepsis and septic shock, irrespective of the number of organ dysfunctions [46–49]. However, the clinical diagnosis of sepsis is challenging in critically ill patients having multiple concurrent disease processes,

with up to 50% of febrile episodes being of non-infectious origin [50]. Furthermore, collection of microbiological evidence for infection is typically slow, and previous antibiotic exposure may render results unreliable. Indeed, cultures remain negative in 30–80% of patients clinically considered infected [51, 52]. Uncertainty regarding antibiotic initiation in patients with suspected lower respiratory tract infection is further complicated by the fact that as many as one-third of pneumonia cases requiring ICU admission are actually viral [53, 54].

In 2016, the Sepsis-3 task force introduced the quick sepsis-related organ failure assessment score (qSOFA), a bedside clinical tool for early sepsis detection [55]. Although the predictive value of qSOFA for in-hospital mortality has been the focus of several external validation studies [56], it remains to be investigated whether this new score may help to rationalize antibiotic use in patients with suspected infection. Yet, published data suggest that qSOFA may lack sensitivity for early

**Table 1** Determinants of increased risk of MDRB infection at ICU admission and during the ICU stay

Predictors of MDRB infection	At ICU admission	During the ICU stay
Patient features	Co-morbid illness/immunosuppression/recent hospital and/or ICU stay	Higher severity of acute illness/Invasive interventions
Type of infection	Hospital-acquired > healthcare-associated > community-acquired	ICU-acquired > others
Antimicrobial selection pressure	Prior antibiotics*/antifungals	Antibiotics*/antifungals in the ICU
Colonization status	Previously documented colonization with MDRB	In-ICU acquisition of MDRB
Local epidemiology	Epidemiology of MDRB in community/hospital/areas recently traveled to	Local epidemiology of MDRB in the ICU
Infection prevention measures	Poor hygiene practices in hospital	Poor hygiene practices in the ICU

MDRB multidrug-resistant bacteria, ICU intensive care unit

*Especially if agents with broad-spectrum and/or potent activity against intestinal anaerobes

identification of patients meeting the Sepsis-3 criteria for sepsis [57].

Hence, antibiotics are mostly used empirically in ICU patients [58]. A provocative before–after study, however, suggested that aggressive empirical antibiotic use might be harmful in this population [59]. In fact, a conservative approach—with antimicrobials started only

after confirmed infection—was associated with a more than 50% reduction in adjusted mortality as well as higher rates of appropriate initial therapy and shorter treatment durations.

Biomarkers may help to identify or—perhaps more importantly—rule out bacterial infections in this setting, thus limiting unnecessary antibiotic use and

encouraging clinicians to search for alternative diagnoses. Many cytokines, cell surface markers, soluble receptors, complement factors, coagulation factors, and acute phase reactants have been evaluated for sepsis diagnosis [60], yet most offer only poor discrimination [61]. Procalcitonin (PCT) levels are high in bacterial sepsis but remain fairly low in viral infections and most cases of non-infectious systemic inflammatory response syndrome (SIRS). However, a PCT-based algorithm for initiation (or escalation) of antibiotic therapy in ICU patients neither decreases overall antimicrobial consumption nor shortens time to adequate therapy or improves patient outcomes [62]. Thus, PCT is currently not recommended as part of the decision-making process for antibiotic initiation in ICU patients [49].

Considering the complexity of the host response and biomarker kinetics, a combined approach which integrates the clinical pretest probability of infection could facilitate the discrimination between bacterial sepsis and non-infectious SIRS in emergency departments and probably also in critically ill patients [63]. Given their high sensitivity, such multi-marker panels may be primarily used to rule out sepsis, albeit only in a subset of patients as a result of their suboptimal specificity (Table E1). In contrast, novel molecular assays for rapid pathogen detection in clinical samples show good specificity, yet poor sensitivity, thus providing a primarily rule-in method for infection (see below). For the foreseeable future, however, physicians will remain confronted with considerable diagnostic uncertainty and, in many cases, still have to rely on their clinical judgment for decisions to withhold or postpone antimicrobial therapy.

Impact of immune status

The host immune status is a key factor for the initial choice of antimicrobial therapy in the ICU [64]. Solid organ transplant recipients receiving immunosuppressive medications to prevent allograft rejection can present with sepsis or septic shock and very few or even no typical warning signs such as fever or leukocytosis. The level of required immunosuppression and the site of infections vary according to the allograft type; the timing of infection from original transplant surgery delineates the occurrence of nosocomial sepsis and opportunistic infections (Table 2) [65]. In hematological or solid cancer patients receiving cytotoxic chemotherapy, the duration and level of neutropenia will be essential factors for the choice of empirical therapy [66, 67]. HIV-infected patients are not only susceptible to community-acquired infections but also to a vast panel of opportunist infections depending on CD4 cell count [68]. Other host

immune profiles encompass immunoglobulin deficiencies and iatrogenic immunosuppression (Table 2) [69]. Because immunocompromised patients may have multiple concomitant dysfunctional immune pathways, co-infections (bacterial and/or viral and/or fungal) are possible and, when suspected, required several antimicrobials as part of empirical therapy. Of note, ageing has been associated with impairments in both innate and adaptive immunity that may predispose to severe bacterial infections; yet, the impact of immunosenescence on the management of ICU patients warrants further investigation [70, 71].

Early microbiological diagnosis: from empirical to immediate adequate therapy

The concepts of empirical therapy and de-escalation originate from the timeframe of routine bacteriological diagnosis. With culture-based methods, the turnaround time from sampling to AST results necessitates 48 h or more, leaving much uncertainty about the adequacy of empirical coverage at the acute phase of sepsis. Molecular diagnostic solutions have therefore been developed to accelerate the process without losing performance in terms of sensitivity and specificity.

A wide array of automated PCR-based systems targeting selected pathogens and certain resistance markers have recently been introduced (Table 3). Several panels are now widely available in clinical laboratories for specific clinical contexts (e.g., suspected bloodstream infections, pneumonia, or meningoencephalitis), offering a “syndromic approach” to microbiological diagnosis [72, 73]. Syndromic tests can be run with minimal hands-on time and identify pathogens faster than conventional methods (i.e., 1.5–6 h), especially when implemented as point-of-care systems. However, these tests remain expensive (>100 USD per test) and must be performed alongside conventional cultures, which they cannot entirely replace. They also provide partial information about antibiotic susceptibility since only a limited number of acquired resistance genes are screened (e.g., those encoding ESBL or carbapenemase). Overall, further investigations are warranted to fully appraise their potential impact on patient outcomes [72].

A next step will be the daily use of clinical metagenomics—that is, the sequencing of nucleic acids extracted directly from a given clinical sample for the identification of all bacterial pathogens and their resistance determinants [74]. Fast sequencers such as the Nanopore (Oxford Nanopore Technologies, Oxford, UK) allow turnaround times of 6–8 h at similar costs to that of syndromic tests [75, 76]. This approach can also assess the host response

Table 2 Spectrum of empirical antimicrobial therapy in immunocompromised patients

Type of immune deficiency	Infection risk to guide antimicrobial rationale	Antimicrobial empirical coverage
Solid organ transplant	Timing from transplant surgery 0–2 months: high risk of HAI 2–6 months: high risk of both HAI and CAI 6–12 months: low risk of HAI, moderate risk of HAI and OI > 12 months: low risk of HAI, moderate risk of CAI and OI	<i>Pseudomonas</i> spp., <i>S. aureus</i> , <i>Candida</i> spp., <i>Aspergillus</i> spp., <i>Cryptococcus</i> spp. <i>Nocardia</i> spp., endemic mycoses, CMV PCP, tuberculosis, <i>S. pneumoniae</i>
Neutropenia	Absolute neutrophil count, duration, and comorbidities > 500 cells/ μ L, anticipated to last < 7 days < 100 cells/ μ L, anticipated to last > 7 days Shock, mucositis, diarrhea, central line	Low risk <i>Pseudomonas</i> spp., <i>S. aureus</i> , <i>S. viridans</i> , molds <i>Pseudomonas</i> spp., <i>S. aureus</i> , <i>S. viridans</i> , <i>Candida</i> spp.
HIV	CD4 cell count 200–500 cells/ μ L: low risk of OI 50–200 cells/ μ L: high risk of OI < 50 cells/ μ L: very high risk of OI HIV-induced humoral immunodeficiency at any CD4 level HIV and intravenous drug abuse	Tuberculosis Tuberculosis, PCP Cryptococcosis, toxoplasmosis, CMV <i>S. pneumoniae</i> <i>S. aureus</i>
Immunoglobulin deficiency	Common variable immunodeficiency Chronic lymphocytic leukemia Multiple myeloma Chronic granulomatous disease	Encapsulated bacteria ^a Encapsulated bacteria ^a , <i>S. aureus</i> Encapsulated bacteria ^a <i>S. aureus</i> , <i>Burkholderia cepacia</i> , <i>Aspergillus</i> spp.
Iatrogenic immunosuppression	Steroids (prednisone > 20 mg/day) Inhibitors of TNF, IL-1, IL-6, IL-17, IL-12/23 Anti-CD20 monoclonal antibodies Anti-CD52 monoclonal antibodies	<i>Candida</i> spp., PCP, <i>Nocardia</i> spp. Tuberculosis, <i>S. aureus</i> , <i>Listeria</i> spp., <i>Legionella</i> Low risk <i>Aspergillus</i> spp., <i>Mucor</i> , <i>Listeria</i> spp.

HAI hospital-acquired infection, CAI community-acquired infection, OI opportunistic infection, CMV cytomegalovirus, PCP *Pneumocystis jirovecii* pneumonia

^a Encapsulated bacteria: *S. pneumoniae*, *H. influenza*, *N. meningitidis*, *Salmonella* spp., *Klebsiella* spp.

Table 3 New diagnostic tools for bacterial infection in critically ill patients

Method	Based on	Available	Pros	Cons
Direct AST	Culture	Yes	Cheap Decreases TAT by 24 h	Lacks standardization Does not work for polymicrobial infection
Accelerate Pheno™	Culture	Yes	Faster than conventional methods Automatized 1 h for identification, 6–8 h for AST	Expensive Low throughput For positive blood cultures only
Lab automation	Culture	Yes	Real-time culturing decreasing TAT	Integration with stewardship Cost Exploitation of results outside working hours
Syndromic tests	PCR	Yes	Fast (TAT 1–8 h) Minimal hands-on time	Expensive Not exhaustive Minimal information on antibiotic resistance
Clinical metagenomics	NGS	In development	Exhaustive Potentially fast Host response	Experimental Interpretation of results Expensive

AST antimicrobial susceptibility testing, TAT turnaround time, NGS next-generation sequencing

at the infection site by sequencing the retro-transcribed RNA, possibly adding to its diagnostic yield [77]. Nonetheless, significant improvements in nucleic extraction rates, antibiotic susceptibility inference, and the exploitation of results into actionable data must be made before clinical metagenomics can be part of routine diagnostic algorithms.

Besides new-generation tools, rapidly applicable information can still be obtained from culture-based methods such as direct AST on lower respiratory tract samples

(time from sample collection ca. 24 h) [78] or lab automation with real-time imaging of growing colonies—for instance, the Accelerate Pheno™ system (Accelerate Diagnostics, Tucson, AZ) provides AST results in 6–8 h from a positive blood culture [79].

To be effective, all these tests must be integrated into the clinical workflow, thereby raising other organizational challenges and requiring the implementation of ASP [80].

The right molecule(s) but avoid the wrong dose

Key features to appraise the optimal dosing of a given antibiotic include the minimum inhibitory concentration (MIC) of the pathogen and the site of infection. Still, for most cases, clear guidance on how to adapt the dose on the basis of such characteristics is lacking, leaving much uncertainty on this issue. Defining the right dose in patients with culture-negative sepsis is a further challenge, although targeting potential pathogens with the highest MICs may appear to be a reasonable approach.

Underdosing of antibiotics is frequent in critically ill patients. Indeed, up to one out of six patients receiving beta-lactams does not reach the minimal concentration target (i.e., free antibiotic concentrations above the MIC of the pathogen during more than 50% of the dosing interval), and many more do not reach the target associated with maximal bacterial killing (i.e., concentrations above $4 \times \text{MIC}$ during 100% of the dosing interval) [81]. This is particularly worrisome in the first hours of therapy when a maximal effect is highly desirable. Unfortunately, no standard remedy for this problem is available, and the solution depends on the physicochemical properties of the drug (e.g., hydrophilic versus lipophilic), patient characteristics, administration scheme, and the use of organ support (e.g., renal replacement therapy or extracorporeal membrane oxygenation) [82].

The volume of distribution—an important determinant of adequate antibiotic concentrations—is not measurable in critically ill patients. Yet, those with evidence for increased volume of distribution (e.g., positive fluid balance) require a higher loading dose to rapidly ensure adequate tissue concentrations, particularly for hydrophilic antibiotics, and for both intermittent and continuous infusion schemes [83]. This first dose must not be adapted to the renal function for antibiotics with predominant or exclusive renal clearance.

Many antibiotics used in the ICU are cleared by the kidneys; so, dosing adaptation for subsequent infusions must be considered in case of acute kidney injury (AKI) or augmented renal clearance (i.e., a measured creatinine clearance of $130 \text{ mL/min/1.73 m}^2$ or higher). This latter situation is associated with lower antibiotic exposure [84] and implies higher maintenance doses to keep concentrations at the targeted level, yet therapeutic drug monitoring (TDM) appears necessary to avoid overdosing.

These features can be integrated into pharmacokinetics (PK)/pharmacodynamics (PD) optimized dosing which can be considered a three-step process (Fig. 3). PK models can be used when selecting the dose for each of these steps [85] even if these predictions are estimations only with still important intra- and inter-individual variations. These are nowadays available in several stand-alone software packages, and integration in prescription drug

monitoring systems (PDMS) will be the next step. TDM can be used to further refine therapy for many antibiotics [86].

Is there a role for routine therapeutic drug monitoring?

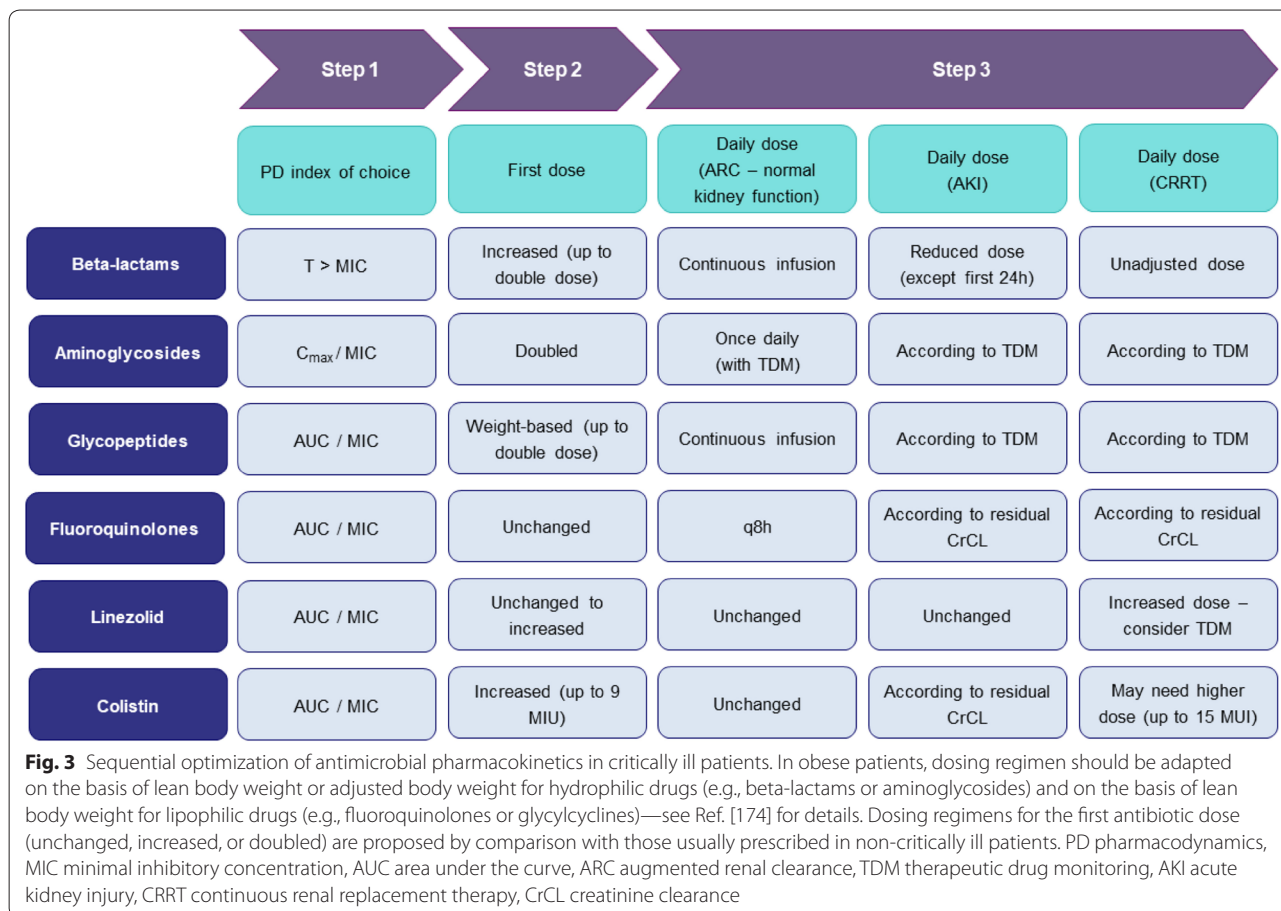
TDM may be employed to minimize the risk of antimicrobial toxicity and maximize drug efficacy through optimized PK, especially for aminoglycosides and glycopeptides. Indeed, high peak levels of aminoglycosides over the pathogen MIC appear beneficial in patients with ventilator-associated pneumonia or other life-threatening MDRB infections [87–89], while adequate trough vancomycin concentrations improve the clinical response in those with bloodstream infection due to MRSA [90].

However, the role of routine TDM in optimizing beta-lactam dosing remains controversial. The main issues that nowadays prevent the implementation of such a strategy in clinical practice are (1) the lack of a standardized method to reliably measure beta-lactam concentrations with a high intercenter reproducibility, (2) the delayed results of TDM for clinicians (i.e., the lack of a “point-of-care” for beta-lactam TDM in most of hospitals), (3) the optimal timing and number of samples to adequately describe the time course of drug concentrations, (4) the fact that the association between insufficient beta-lactam concentrations and the increased risk of therapeutic failure or impaired outcome is based only on retrospective studies, (5) the absence of clinical data showing a potential role of adequate beta-lactam levels in the emergence of resistant strains, (6) the poor characterization of the optimal duration of beta-lactam levels exceeding the MIC of the infective pathogen, when available, or of the optimal PK target in case of empirical therapy, and (7) the time needed to obtain the MIC of the infective pathogen, which precludes an adequate targeted therapy using PK principles [81, 91]. It is therefore possible that epidemiological cutoff (ECOFF) values are an acceptable option [92], but further studies are needed before the routine TDM of beta-lactams becomes available in most ICUs. Interestingly, high beta-lactam concentrations may result in drug-related neurotoxicity, which represents another potential role for TDM in critically ill patients [93, 94].

Key questions about antimicrobials

New and long-established antimicrobials

Polymyxins are considered the cornerstone of therapy for infections due to extremely drug-resistant (XDR) Gram-negative bacteria, including carbapenem-resistant *A. baumannii*, *P. aeruginosa*, and *K. pneumoniae*. Of note, recent studies indicate that colistin and polymyxin B are associated with less renal and neurological



toxicity than previously reported. Several questions remain incompletely addressed, including the need and type of combination therapies, optimal dosing regimen, ways to prevent the emergence of resistance, and role of aerosolized therapy. Fosfomycin may also have a role in these infections.

Drugs newly approved or in late development phase mainly include ceftolozane–tazobactam, ceftazidime–avibactam, ceftaroline–avibactam, aztreonam–avibactam, carbapenems combined with new beta-lactamase inhibitors (e.g., vaborbactam, relebactam), cefiderocol, plazomicin, and eravacycline (Table 4). These drugs have mainly been tested in complicated urinary tract infection, complicated intra-abdominal infections (cIAI), or skin and soft tissue infection (SSTI). Limited data are currently available in ICU patients [95], notably for dosing optimization in severe MDRB infections. Piperacillin–tazobactam appears less effective than carbapenems in bloodstream infections caused by ESBL-E [96, 97]; however, ceftolozane–tazobactam and ceftazidime–avibactam might be considered as carbapenem-sparing options for treatment of such infections in areas with

high prevalence of CRE. The actual question is should we still save carbapenems instead of saving new antibiotics?

In addition to glycopeptides, long-established antibiotics with activity against MRSA mainly include daptomycin (e.g., for bloodstream infections) and linezolid (e.g., for hospital-acquired pneumonia, HAP) [98, 99]. These alternatives may be preferred in patients with risk factors for AKI. Daptomycin appears safe even at high doses and in prolonged regimens, with rhabdomyolysis representing a rare, reversible side effect. Conversely, linezolid has been linked with several adverse events most often associated with specific risk factors (e.g., renal impairment, underlying hematological disease, or extended therapy duration), suggesting a role for TDM in patients at high risk of toxicity. Next, “new-generation” cephalosporins such as ceftaroline and ceftobiprole have been approved for the treatment of MRSA infections and seem promising in overcoming the limitations associated with the older compounds. Other new agents with activity against MRSA include lipoglycopeptides (dalbavancin, oritavancin, and telavancin), fluoroquinolones (delafloxacin, nemonoxacin, and zabofloxacin), an oxazolidinone (tedizolid), a dihydrofolate reductase inhibitor (iclaprim), and

Table 4 Indications and doses of new and long-established antibiotics for treating MDR bacteria

Drug	Usual dosing regimen for serious infections	Indication	Status
Recent release or late development phases			
Ceftaroline	600 mg q12 h, IV	BSI, CAP, cSSTI	Approved
Ceftobiprole	500 mg q8 h IV	BSI, HAP	Approved
Ceftazidime/avibactam	2.5 g q8 h IV	BSI, HAP, VAP, cIAI, UTI	Approved
Ceftolozane/tazobactam	1.5 g q8 h/3 g q8 h (VAP) IV	BSI, UTI, cIAI, HAP, VAP	Approved for cIAI and UTI Phase 3 for HAP and VAP
Aztreonam/avibactam	6500 mg ATM/2167 mg AVI q24 h on day 1 followed by 6000 mg ATM/2000 mg AVI q24 h, IV	HAP, VAP, BSI, UTI	Phase 3
Meropenem/vaborbactam	2 g/2 g q8 h IV	BSI, UTI, cIAI, HAP/VAP	Approved (FDA)
Cefiderocol	2 g q8 h IV	BSI, HAP, VAP, cIAI, UTI	Phase 3
Imipenem/relebactam	500 mg/250–125 mg q6 h IV	BSI, HAP, VAP, cIAI, UTI	Phase 3
Eravacycline	1 mg/kg q12 h IV	cIAI	Under evaluation (EMA and FDA)
Plazomicin	15 mg/kg q24 h IV	In combination for BSI, UTI, HAP, VAP	Approved
Tedizolid	200 mg q24 h IV, oral	cSSTI, HAP/VAP	Approved for cSSTI, phase 3 for HAP and VAP
Long-established antibiotics			
Piperacillin/tazobactam	4.5 g every 6 h CI	BSI, HAP, VAP, UTI, cIAI	Approved
Ceftazidime	6 g every 24 h CI	BSI, HAP, VAP, UTI	Approved
Cefepime	2 g every 8 h or CI	BSI, HAP, VAP, UTI	Approved
Aztreonam	1 g (2 g) every 8 h	BSI, HAP, VAP, UTI, SSTI	Approved
Imipenem/cilastatin	500 mg (1 g) every 6 h	BSI, HAP, VAP, UTI, cIAI	Approved
Meropenem	1 g (2 g) every 8 h or CI	BSI, HAP, VAP, UTI, cIAI	Approved
Tigecycline	100–200 mg loading those, then 50–100 mg every 12 h	cIAI	Approved
"Old" antibiotics			
Gentamicin	7 mg/kg/day every 24 h	In combination for BSI, UTI, c HAP, cIAI, VAP	Approved
Amikacin	25–30 mg/kg/day every 24 h	In combination for BSI, UTI, VA HAP, VAP	Approved
Colistin	9 MU loading dose, 4.5 MU every 8–12 h	In combination for BSI, UTI, HAP, VAP	Approved
Fosfomycin	4–6 g every 6 h CI	In combination for BSI, UTI, HAP, VAP	Approved
Vancomycin	15–30 mg/kg loading dose, 30–60 mg/kg every 12 h, 6 h or CI	BSI, HAP, VAP	Approved
Linezolid	600 mg every 12 h	BSI, HAP, VAP, SSTI	Approved

BSI bloodstream infection, HAP hospital-acquired pneumonia, VAP ventilator-associated pneumonia, cIAI complicated intra-abdominal infection, UTI urinary tract infection, CI continuous infusion, FDA US Food and Drug Administration, EMA European Medicines Agency

a tetracycline (omadacycline); yet, the yield of these new options remains to be investigated in critically ill patients with severe MRSA infection [100].

Single-drug or combination regimen

The question of whether antibiotic combinations provide a beneficial effect beyond the empirical treatment period remains unsettled. Meta-analyses of randomized controlled trials (RCTs) comparing beta-lactams vs. beta-lactams combined with another agent demonstrate no difference in clinical outcomes in a variety of infections caused by Gram-negative pathogens; however, patients

with sepsis or septic shock were underrepresented [101, 102]. In contrast, a meta-analysis of randomized and observational studies focused on sepsis or septic shock showed that combination therapy is beneficial in high-risk patients (i.e., projected mortality rate greater than 25%) [103]. This positive impact may be especially pronounced in neutropenic patients and when a pathogen with reduced antimicrobial susceptibility is involved (e.g., *P. aeruginosa*) [104].

To date, there is no RCT to examine whether combination therapy is superior to monotherapy for CRE infections. Observational studies suggest that the benefit of

combination therapy is mainly observed in patients with serious underlying diseases or high pretreatment probability of death (e.g., septic shock) [105–109]. The most effective regimen is challenging to define, as only one of the aforementioned studies reported survival benefit with a specific drug combination (colistin plus tigecycline plus meropenem) after adjustment for potential confounders [109].

Although there have been five RCTs and several meta-analyses for the treatment of carbapenem-resistant *A. baumannii* infections, the optimal treatment regimen has not yet been determined [110–115]. Notably, none of the RCTs demonstrated a survival benefit with combination therapy, although one study showed a better clinical response with colistin plus high-dose ampicillin/sulbactam and three studies reported faster microbiological clearance when combining colistin with rifampin or fosfomycin. A recent meta-analysis, however, demonstrated survival benefit in bacteremic patients who were receiving high doses of colistin (more than 6 MIU per day) in combination with another agent [116].

Continuous prolonged or intermittent administration of beta-lactams and other time-dependent antimicrobials

The proportion of the interdose interval with drug concentration above the pathogen MIC is predictive of efficacy for time-dependent antibiotics, including beta-lactams. This parameter may be increased by reducing the interdose interval and/or by using extended infusions (EI) over 3–4 h or continuous infusion (CI). Stochastic models show that prolonged beta-lactam infusions increase the probability of target attainment against isolates with borderline MIC, especially in patients with ARC or increased volume of distribution [117].

Most RCTs comparing intermittent versus prolonged beta-lactam infusions could not find significant differences in outcomes. However, in a recent meta-analysis of RCTs comparing prolonged (EI or CI) and intermittent infusions of antipseudomonal beta-lactams in patients with sepsis, prolonged infusion was associated with improved survival, including when carbapenems or beta-lactam/beta-lactam inhibitor combinations were analyzed separately [118]. Prolonged infusions might only be needed in some patients—e.g., those with beta-lactam underdosing using intermittent administration schemes, or infections caused by isolates with elevated MICs. Because these features cannot be anticipated, it seems reasonable to consider the use of prolonged infusions of sufficiently stable antipseudomonal beta-lactams in all patients with sepsis.

For some other drugs such as vancomycin, the ratio area under the curve/MIC is considered the PK/PD parameter predictive of efficacy (Fig. 3). A recent

meta-analysis suggested that continuous vancomycin infusion is associated with lower nephrotoxicity but not better cure or lower mortality than intermittent infusions [119]; nevertheless, included studies had many limitations and further investigations are needed to address this issue.

De-escalation: impact in practice

Conceptually, de-escalation is a strategy whereby the provision of effective antibiotic treatment is achieved, while minimizing unnecessary exposure to broad-spectrum agents that would promote the development of resistance. Practically, it consists in the reappraisal of antimicrobial therapy as soon as AST results are available. However, no clear consensus on de-escalation components exists and various definitions have been used (e.g., changing the “pivotal” agent for a drug with a narrower spectrum and/or lower ecological effects on microbiota, or discontinuing an antimicrobial combination), resulting in equivocal interpretation of the available evidence [120, 121].

De-escalation is applied in only 40–50% of inpatients with bacterial infection [121]. This reflects physician reluctance to narrow the covered spectrum when caring for severely ill patients with culture-negative sepsis and/or MDRB carriage [120]. Importantly, the available evidence does not suggest a detrimental impact of de-escalation on outcomes [120, 122], including in high-risk patients such as those with bloodstream infections, severe sepsis, VAP, and neutropenia [123, 124]. However, further well-designed RCTs are needed to definitely solve this issue.

Increasing physician confidence and compliance with de-escalation has become a cornerstone of ASP. Paradoxically, there is a lack of clinical data regarding the impact of de-escalation on antimicrobial consumption and emergence of resistance [120]. While this strategy has been associated with reduced use of certain antimicrobial classes [125, 126], no study demonstrated that it may allow a decrease in overall antimicrobial consumption, and an increase in antibiotic exposure has even been observed [123, 125, 127]. Similarly, the few studies that addressed this point reported no impact—or only a marginal effect—of de-escalation on the individual hazard of MDRB acquisition or local prevalence of MDRB [125–127].

In light of these uncertainties, efforts should focus on microbiological documentation to increase ADE rates in patients with sepsis. New diagnostic tools should be exploited to hasten pathogen identification and AST availability. Lastly, human data on the specific impact of each antimicrobial on commensal ecosystems and the risk of MDRB acquisition are needed to optimize

antibiotic streamlining and further support de-escalation strategies [37, 128].

Duration of antibiotic therapy and antibiotic resistance

Prolonged durations of antibiotic therapy have been associated with the emergence of antimicrobial resistance [129]. Yet, short-course antibiotic therapy has been shown to be effective and safe in a number of infections, including community-acquired pneumonia, VAP, urinary tract infections, cIAI, and even some types of bacteremia [130–136]. The shortening of antibiotic durations on the basis of PCT kinetics has also been shown to be safe, including in patients with sepsis [51, 52]. However, the recent ProACT trial failed to confirm the ability of PCT to reduce the duration of antibiotic exposure compared to usual care in suspected lower respiratory tract infections [137]. Given the importance of overruling in available RCTs and the relatively long duration of therapy in control groups, the question remains unresolved. In particular, the efficacy and costs of PCT if an active ASP is in place remain to be evaluated.

Many national and international guidelines encourage physicians to shorten the overall durations of antibiotic therapy for a number of infections. Shorter courses are now recommended for pneumonia, urinary tract infections, and cIAI with source control [49, 138–142]. However, despite the presence of these recommendations, recent studies suggest that excessive durations of antibiotics are still being administered, thereby offering further opportunities for ASP [143, 144]. However, clinicians should also be aware that, under some circumstances, short-course therapy may be detrimental to patient outcomes, especially in case of prolonged neutropenia, lack of adequate source control, infection due to XDR Gram-negative bacteria, and endovascular or foreign body infections [130, 145].

Source control

Source control to eliminate infectious foci follows principles of drainage, debridement, device removal, compartment decompression, and often deferred definitive restoration of anatomy and function [146]. If required, source control is a major determinant of outcome, more so than early adequate antimicrobial therapy [147–149], and should never be considered as “covered” by broad-spectrum agents. Therefore, surgical and radiological options for intervention must be systematically discussed, especially in patients with cIAI or SSTI. The efficacy of source control is time-dependent [150–153] and adequate procedures should therefore be performed as rapidly as possible in patients with septic shock [49],

while longer delays may be acceptable in closely monitored stable patients. Failure of source control should be considered in cases of persistent or new organ failure despite resuscitation and appropriate antimicrobial therapy, and requires (re)imaging and repeated or alternative intervention. Importantly, source control procedures should include microbiological sampling whenever possible to facilitate ADE initiatives.

Antibiotic stewardship programs in the ICU

Implementing ASP in the ICU improves antimicrobial utilization and reduces broad-spectrum antimicrobial use, incidence of infections and colonization with MDRB, antimicrobial-related adverse events, and healthcare-associated costs, all without increase in mortality [26, 154, 155]. According to the ESCMID Study Group for Antimicrobial Stewardship, ASP should be approached as “a coherent set of actions which promote using antimicrobials in ways that ensure sustainable access to effective therapy for all who need them” [156]. Therefore, ASP should be viewed as a quality improvement initiative, requiring (1) an evidence-based, ideally bundled, change package, (2) a clear definition of goals, indicators, and targets, (3) a dynamic measurement and data collection system with feedback to prescribers, (4) a strategy for building capacity, and (5) a plan to identify and approach areas for improvement and solve quality gaps. This necessarily implies the appointment of a member of the ICU staff as a leader with expertise in the field of antimicrobial therapy and prespecified functions for the implementation of the local ASP.

Three main kinds of interventions may be used in ASP [157–159]:

- Restrictive, in which one tries to reduce the number of opportunities for bad behavior, such as formulary restrictions, pre-approval by senior ASP doctor (either an external infectious disease specialist or a specified expert in the ICU team), and automatic stop orders
- Collaborative or enhancement, in which one tries to increase the number of opportunities and decrease barriers for good behavior, such as education of prescribers, implementation of treatment guidelines, promotion of ADE, use of PK/PD concepts, and prospective audit and feedback to providers
- Structural, which may include the use of computerized antibiotic decision support systems, faster diagnostic methods for antimicrobial resistance, antibiotic consumption surveillance systems, ICU leadership commitment, staff involvement, and daily collaboration between ICU staff, pharmacists, infection control units, and microbiologists

The implementation of ASP should take into account the need for a quick answer from the system in case of severe infections (e.g., regarding as unacceptable the delay in the first antimicrobial delivery due to too restrictive pharmacy-driven prescription policies).

An ASP should consensually rest on multifaceted interventions to achieve its fundamental goals (Table 5), namely improving outcomes and decreasing antimicrobial-related collateral damage in infected patients. Yet, the weight of each component must be customized according to the context and culture of every single ICU in terms of habits for antibiotic prescription, MDRB prevalence, local organizational aspects, and available resources. For this purpose, concepts of implementation science should be applied—that is, identifying barriers and facilitators that impact the staff's compliance to guidelines in order to design and execute a structured plan for improvement [160].

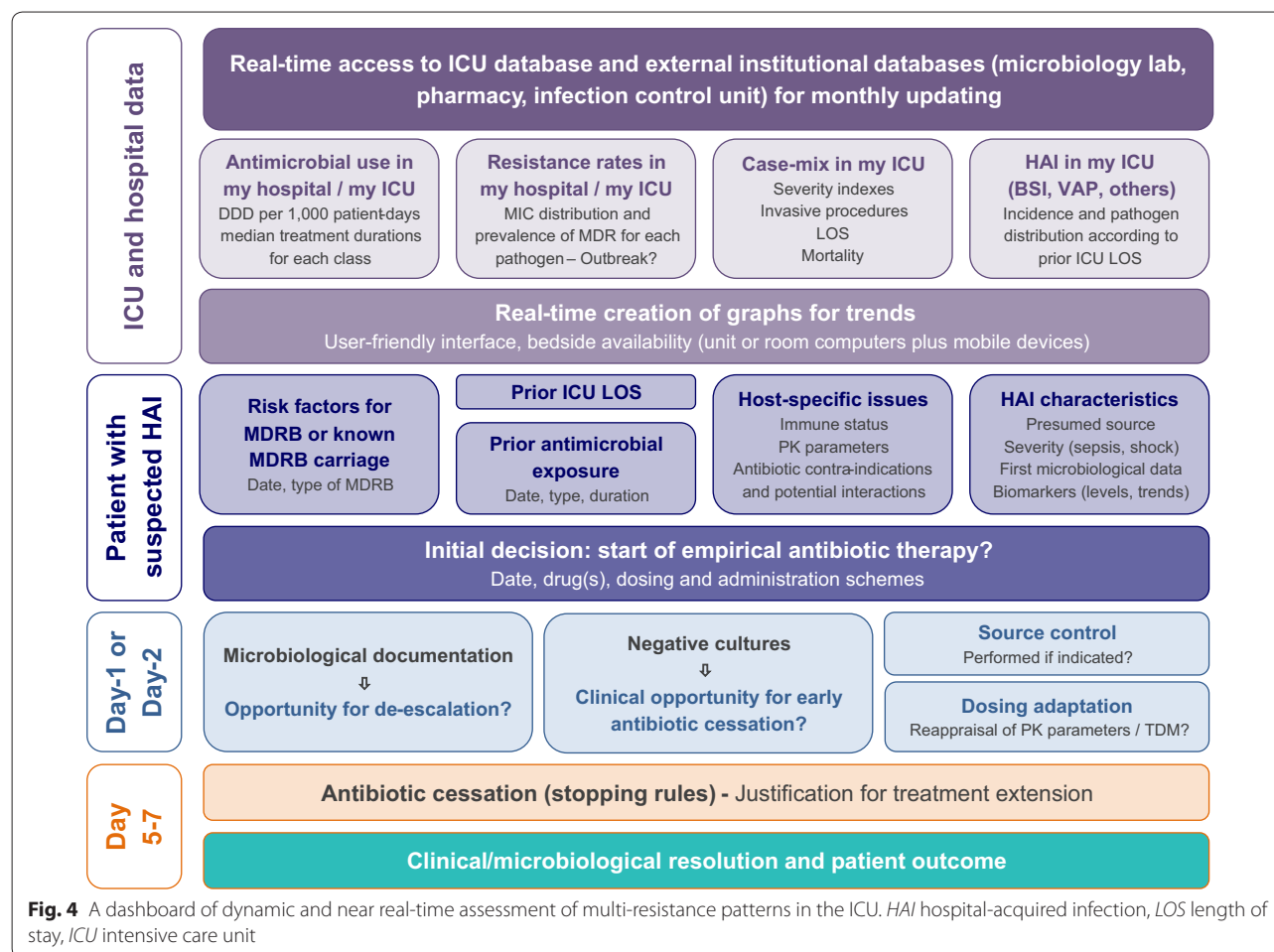
The appropriate dashboard in the ICU

The availability of constantly updated information is pivotal to improve decision-making processes in the ICU

[161, 162]. As the epidemiology of MDRB is continuously evolving, close monitoring of local resistance patterns may help to rationalize the empirical use of broad-spectrum antibiotics in this setting. With the expanding utilization of electronic medical records and applications specifically developed for the ICU, streaming analytics can provide dashboards containing real-time and easily accessible data for intensivists [162, 163]. Such dashboards should capture data from medical records and microbiology systems, display an intuitive and user-friendly interface, and be available on both ICU computers and mobile devices to allow easy access to actionable data at the bedside. Finally, a complete dashboard should include information not only on dynamics of resistance patterns but also on local antimicrobial consumption, adherence to protocols of care and antibiotic guidelines, healthcare-associated infections (e.g., source, type, severity), and general patient characteristics (e.g., comorbidities, severity of illness, main diagnosis, and length of the ICU stay) (Fig. 4). Although studies demonstrating the efficacy of such dashboards in reducing resistance have not been published so far, these tools could

Table 5 Implementation and objectives of antibiotic stewardship programs in the ICU

Implementation of ASP	
Pre-requisites	Evidence-based, ideally bundled change package Dynamic data collection systems with feedback to prescribers Strategy for building capacity, including the appointment of an ICU staff member as ASP leader
Pre-implementation phase	Identification of determinants for antibiotic prescription and opportunities for improvement at the ICU level
Implementation phase	Building of a customized plan to solve quality gaps, based on educational and behavioral interventions Continuous collaboration between ICU staff members, microbiologists, pharmacists, and infection control units Clear definition of goals and indicators
Pragmatic objectives of ASP	
Stewardship of empirical antibiotic therapy	Distinction between bacterial infections, non-bacterial infections, and non-infectious inflammatory syndromes Early identification of sepsis (<i>antibiotic initiation might be delayed pending microbiological data in certain patients without new or worsening organ failure</i>) Consideration of local resistance patterns and patient's individual risk factors for MDRB for the choice of empirical drugs Efforts to obtain early microbiological documentation (including rapid diagnostic tools, conventional cultures, and source control when appropriate) Optimization of PK/PD Promotion of single-drug regimen whenever possible for patients without septic shock and/or risk factors for MDRB Restricted use of broad-spectrum, costly, and/or potentially toxic antibiotics
Stewardship of definite antibiotic therapy	Reappraisal of the diagnosis of bacterial infection at day 2–3 (microbiological and radiological data, clinical evolution) Early antibiotic cessation in patients without confirmed infection In patients with likely or confirmed infection: dosing adaptation when appropriate (e.g., if changes in Vd and/or renal clearance), routine discussion for de-escalation (e.g., spectrum narrowing and switching from combination to single-drug regimen) and shortening of treatment duration (e.g., PCT-based algorithms, adequate source control, favorable clinical evolution)
Overall objectives	Improvement in patient outcomes Reduction in ecological and non-ecological (e.g., toxicity or allergy) side effects of antibiotics Reduction of antibiotic- and resistance-related costs



allow a structured audit-feedback approach that is one of the cornerstones of ASP implementation in the ICU [164–166].

Concluding remarks

Both the poor outcomes associated with bacterial sepsis and the current epidemiology of MDRB urge the need for improving the management of antibiotic therapy in ICU patients. Well-designed studies remain warranted to definitely address several aspects of this issue, notably the clinical input of rapid diagnostic tools and TDM, the potential benefit of combination versus single-drug therapies, the optimal dosing regimens before the availability of AST results or for patients with culture-negative sepsis, and the prognostic yield of ASP. Although beyond the scope of this review, the exploitation of other research axes may further help to control the spread of MDRB in the ICU setting, including optimization of infection control policies [167], a comparative appraisal of the impact of broad-spectrum antibiotics on the gut microbiota through novel metagenomics approaches [168], and the

evaluation of emerging options such as orally administered antimicrobial-adsorbing charcoals, probiotics, or fecal microbiota transplantation to protect or restore the commensal ecosystems of ICU patients [29, 169, 170].

Electronic supplementary material

The online version of this article (<https://doi.org/10.1007/s00134-019-05520-5>) contains supplementary material, which is available to authorized users.

Author details

¹ Medical and Infectious Diseases ICU, APHP, Bichat-Claude Bernard Hospital, 46 Rue Henri-Huchard, 75877 Paris Cedex 18, France. ² INSERM, IAME, UMR 1137, Paris-Diderot Sorbonne-Paris Cité University, Paris, France. ³ Infectious Diseases Division, Department of Medicine, University of Udine and Azienda Sanitaria Universitaria Integrata di Udine, Udine, Italy. ⁴ Department of Intensive Care Medicine, University Medical Center Utrecht, Utrecht, The Netherlands. ⁵ School of Medicine, National and Kapodistrian University of Athens, Athens, Greece. ⁶ Department of Critical Care Medicine, Ghent University Hospital, Ghent, Belgium. ⁷ Department of Internal Medicine, Division of Infectious Diseases, University of Nebraska Medical Center, Omaha, NE, USA. ⁸ Surgical Critical Care Unit, Department of Anesthesiology, Critical Care and Perioperative Medicine, CHU Lille, Lille, France. ⁹ Critical Care Research, Washington University School of Medicine and Respiratory Care Services, Barnes-Jewish Hospital, St. Louis, MO, USA. ¹⁰ Department of Medicine, Royal Inland Hospital, Kamloops, Canada. ¹¹ Intensive Care Medicine Department, Centro Hospitalar

São João and Faculty of Medicine, University of Porto, Porto, Portugal.

¹² Clinical Unit of Infectious Diseases, Microbiology and Preventive Medicine, Hospital Universitario Virgen Macarena, Department of Medicine, University of Sevilla, Biomedicine Institute of Sevilla (IBIS), Sevilla, Spain. ¹³ Bacteriology Laboratory, Bichat-Claude Bernard Hospital, APHP, Paris, France. ¹⁴ Department of Critical Care and Graduate Program in Translational Medicine, D'Or Institute for Research and Education, IDOR, Rio De Janeiro, Brazil. ¹⁵ Department of Intensive Care, Erasme Hospital, Brussels, Belgium. ¹⁶ Department of Anesthesiology and Critical Care, Beaujon Hospital, AP-HP, Clichy, France. ¹⁷ INSERM, CRI, UMR 1149, Paris-Diderot Sorbonne-Paris Cité University, Paris, France. ¹⁸ Medical ICU, La Source Hospital, CHR Orléans, Orléans, France.

Compliance with ethical standards

Conflicts of interests

Dr. Kollef's effort was supported by the Barnes-Jewish Hospital Foundation. Dr. Barbier received consulting fees, speaker fees, and conference invitation from MSD, and conference invitation from Pfizer. Dr. De Waele has been consultant for Accelerate Diagnostics, Bayer Healthcare, Grifols, MSD, and Pfizer (all honoraria paid to institution). Dr. Timsit received fees for scientific advisory boards from Pfizer, Bayer pharma, Merck, Nabriva and received fees for lectures from Biomerieux, Pfizer, Merck, Gilead. Dr. Rodríguez-Baño received honoraria for accredited educational activities from Merck and for coordination of a non-product-related research project by Astra-Zeneca. Dr. Ruppé received speaker fees from MSD, Sanofi, and Mobidiag, fees for scientific advisory board from Pathoquest and MaaT Pharma, and consulting fees from DaVolterra. Dr. Daikos reports grants from Pfizer, Gilead, and honoraria from Pfizer, Achaogen, MSD, and Rempex, outside of this work. Dr. Kipnis received fees for scientific advisory boards, lectures, and conference invitation from MSD, for lectures and conference invitations from Pfizer and Fresenius, and conference invitations from LFB and Gilead. Dr. Weiss reports speaker fees from Baxter, MSD, and Gilead, congress and travel reimbursements from MSD and Eumedica, and having participated in an advisory board from Biomerieux.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Received: 14 November 2018 Accepted: 4 January 2019

Published online: 18 January 2019

References

- Versporten A, Zarb P, Caniaux I, Gros MF, Drapier N, Miller M et al (2018) Antimicrobial consumption and resistance in adult hospital inpatients in 53 countries: results of an internet-based global point prevalence survey. *Lancet Glob Health* 6(6):e619–e629
- Detsis M, Karanika S, Mylonakis E (2017) ICU acquisition rate, risk factors, and clinical significance of digestive tract colonization with extended-spectrum beta-lactamase-producing Enterobacteriaceae: a systematic review and meta-analysis. *Crit Care Med* 45(4):705–714
- Kollef MH, Chastre J, Fagon JY, Francois B, Niederman MS, Rello J et al (2014) Global prospective epidemiologic and surveillance study of ventilator-associated pneumonia due to *Pseudomonas aeruginosa*. *Crit Care Med* 42(10):2178–2187
- Bulens SN, Yi SH, Walters MS, Jacob JT, Bower C, Reno J et al (2018) Carbapenem-nonsusceptible *Acinetobacter baumannii*, 8 US metropolitan areas, 2012–2015. *Emerg Infect Dis* 24(4):727–734
- Hsu LY, Apisarnthanarak A, Khan E, Suwantarant N, Ghafur A, Tambyah PA (2017) Carbapenem-resistant *Acinetobacter baumannii* and Enterobacteriaceae in South and Southeast Asia. *Clin Microbiol Rev* 30(1):1–22
- Rodríguez CH, Balderrama Yaruhui N, Nastro M, Nunez Quezada T, Castro Canarte G, Magne Ventura R et al (2016) Molecular epidemiology of carbapenem-resistant *Acinetobacter baumannii* in South America. *J Med Microbiol* 65(10):1088–1091
- Nowak J, Zander E, Stefanik D, Higgins PG, Roca I, Vila J et al (2017) High incidence of pandrug-resistant *Acinetobacter baumannii* isolates collected from patients with ventilator-associated pneumonia in Greece, Italy and Spain as part of the MagicBullet clinical trial. *J Antimicrob Chemother* 72(12):3277–3282
- Lob SH, Biedenbach DJ, Badal RE, Kazmierczak KM, Sahm DF (2015) Antimicrobial resistance and resistance mechanisms of Enterobacteriaceae in ICU and non-ICU wards in Europe and North America: SMART 2011–2013. *J Glob Antimicrob Resist* 3(3):190–197
- Bonomo RA, Burd EM, Conly J, Limbago BM, Poirel L, Segre JA et al (2018) Carbapenemase-producing organisms: a global scourge. *Clin Infect Dis* 66(8):1290–1297
- Rosenthal VD, Al-Abdely HM, El-Kholy AA, AlKhawaja SAA, Leblebicioglu H, Mehta Y et al (2016) International nosocomial infection control consortium report, data summary of 50 countries for 2010–2015: device-associated module. *Am J Infect Control* 44(12):1495–1504
- Weiner LM, Webb AK, Limbago B, Dudeck MA, Patel J, Kallen AJ et al (2016) Antimicrobial-resistant pathogens associated with healthcare-associated infections: summary of data reported to the National Healthcare Safety Network at the Centers for Disease Control and Prevention, 2011–2014. *Infect Control Hosp Epidemiol* 37(11):1288–1301
- Gao L, Lyu Y, Li Y (2017) Trends in drug resistance of *Acinetobacter baumannii* over a 10-year period: nationwide data from the China surveillance of antimicrobial resistance program. *Chin Med J (Engl)* 130(6):659–664
- Bonell A, Azarrafy R, Huong VTL, Viet TL, Phu VD, Dat VQ et al (2018) A systematic review and meta-analysis of ventilator associated pneumonia in adults in Asia; an analysis of national income level on incidence and etiology. *Clin Infect Dis*. <https://doi.org/10.1093/cid/ciy543>
- Hu FP, Guo Y, Zhu DM, Wang F, Jiang XF, Xu YC et al (2016) Resistance trends among clinical isolates in China reported from CHINET surveillance of bacterial resistance, 2005–2014. *Clin Microbiol Infect* 22(Suppl 1):S9–S14
- Bitterman R, Hussein K, Leibovici L, Carmeli Y, Paul M (2016) Systematic review of antibiotic consumption in acute care hospitals. *Clin Microbiol Infect* 22(6):561
- Holmes AH, Moore LS, Sundsfjord A, Steinbakk M, Regmi S, Karkey A et al (2016) Understanding the mechanisms and drivers of antimicrobial resistance. *Lancet* 387(10014):176–187
- Barbier F, Luyt CE (2016) Understanding resistance. *Intensive Care Med* 42(12):2080–2083
- Ruppé E, Woerther PL, Barbier F (2015) Mechanisms of antimicrobial resistance in Gram-negative bacilli. *Ann Intensive Care* 5:21
- Abdallah M, Badawi M, Amirah MF, Rasheed A, Mady AF, Alodat M et al (2017) Impact of carbapenem restriction on the antimicrobial susceptibility pattern of *Pseudomonas aeruginosa* isolates in the ICU. *J Antimicrob Chemother* 72(11):3187–3190
- Zagorianou A, Sianou E, Iosifidis E, Dimou V, Protonotariou E, Miyakis S et al (2012) Microbiological and molecular characteristics of carbapenemase-producing *Klebsiella pneumoniae* endemic in a tertiary Greek hospital during 2004–2010. *Euro Surveill* 17(7):20088
- Meyer E, Schwab F, Schroeren-Boersch B, Gastmeier P (2010) Dramatic increase of third-generation cephalosporin-resistant *E. coli* in German intensive care units: secular trends in antibiotic drug use and bacterial resistance, 2001–2008. *Crit Care* 14(3):R113
- Bassetti M, Cruciani M, Righi E, Rebesch B, Fasce R, Costa A et al (2006) Antimicrobial use and resistance among Gram-negative bacilli in an Italian intensive care unit. *J Chemother* 18(3):261–267
- Samonis G, Korbila IP, Maraki S, Michailidou I, Vardakas KZ, Kofteridis D et al (2014) Trends in isolation of intrinsically resistant to colistin Enterobacteriaceae and association with colistin use in a tertiary hospital. *Eur J Clin Microbiol Infect Dis* 33(9):1505–1510
- Jacoby TS, Kuchenbecker RS, Dos Santos RP, Magedanz L, Guzzato P, Moreira LB (2010) Impact of hospital-wide infection rate, invasive procedures use and antimicrobial consumption on bacterial resistance inside an intensive care unit. *J Hosp Infect* 75(1):23–27
- Fihman V, Messika J, Hajage D, Tournier V, Gaudry S, Magdoud F et al (2015) Five-year trends for ventilator-associated pneumonia: correlation between microbiological findings and antimicrobial drug consumption. *Int J Antimicrob Agents* 46(5):518–525

26. Kaki R, Elligsen M, Walker S, Simor A, Palmay L, Daneman N (2011) Impact of antimicrobial stewardship in critical care: a systematic review. *J Antimicrob Chemother* 66(6):1223–1230
27. Bell BG, Schellevis F, Stobberingh E, Goossens H, Pringle M (2014) A systematic review and meta-analysis of the effects of antibiotic consumption on antibiotic resistance. *BMC Infect Dis* 14:13
28. Nijssen S, Fluit AC, van de Vijver D, Top J, Willems R, Bonten MJM (2010) Effects of reducing beta-lactam antibiotic pressure on intestinal colonization of antibiotic-resistant Gram-negative bacteria. *Intensive Care Med* 36(3):512–519
29. Ruppe E, Burdet C, Grall N, de Lastours V, Lescure FX, Andremon A et al (2018) Impact of antibiotics on the intestinal microbiota needs to be re-defined to optimize antibiotic usage. *Clin Microbiol Infect* 24(1):3–5
30. Pamer EG (2016) Resurrecting the intestinal microbiota to combat antibiotic-resistant pathogens. *Science* 352(6285):535–538
31. Razazi K, Derde LP, Verachten M, Legrand P, Lesprit P, Brun-Buisson C (2012) Clinical impact and risk factors for colonization with extended-spectrum beta-lactamase-producing bacteria in the intensive care unit. *Intensive Care Med* 38(11):1769–1778
32. Poignant S, Guinard J, Guignon A, Bret L, Poisson D-M, Boulain T et al (2015) Risk factors and outcomes of intestinal carriage of AmpC-hyperproducing *Enterobacteriaceae* in ICU patients. *Antimicrob Agents Chemother* 60(3):1883–1887
33. Hilliquin D, Le Guern R, Thepot Seegers V, Neulier C, Lomont A, Marie V et al (2018) Risk factors for acquisition of OXA-48-producing *Klebsiella pneumoniae* among contact patients: a multicentre study. *J Hosp Infect* 98(3):253–259
34. Papadimitriou-Olivgeris M, Marangos M, Fligou F, Christofidou M, Bartzavali C, Anastassiou ED et al (2012) Risk factors for KPC-producing *Klebsiella pneumoniae* enteric colonization upon ICU admission. *J Antimicrob Chemother* 67(12):2976–2981
35. Tan BK, Vivier E, Ait Bouziad K, Zahar JR, Pommier C, Parmeland L et al (2018) A hospital-wide intervention replacing ceftriaxone with cefotaxime to reduce rate of healthcare-associated infections caused by extended-spectrum beta-lactamase-producing *Enterobacteriaceae* in the intensive care unit. *Intensive Care Med* 44(5):672–673
36. Grohs P, Kerneis S, Sabatier B, Lavollay M, Carboneille E, Rostane H et al (2014) Fighting the spread of AmpC-hyperproducing *Enterobacteriaceae*: beneficial effect of replacing ceftriaxone with cefotaxime. *J Antimicrob Chemother* 69(3):786–789
37. Woerther PL, Lepeule R, Burdet C, Decousser JW, Ruppe E, Barbier F (2018) Carbapenems and alternative beta-lactams for the treatment of infections due to ESBL-producing *Enterobacteriaceae*: what impact on intestinal colonization resistance? *Int J Antimicrob Agents* 52:762–770
38. Kao KC, Chen CB, Hu HC, Chang HC, Huang CC, Huang YC (2015) Risk factors of methicillin-resistant *Staphylococcus aureus* infection and correlation with nasal colonization based on molecular genotyping in medical intensive care units: a prospective observational study. *Medicine (Baltimore)* 94(28):e1100
39. Ziakas PD, Anagnostou T, Mylonakis E (2014) The prevalence and significance of methicillin-resistant *Staphylococcus aureus* colonization at admission in the general ICU setting: a meta-analysis of published studies. *Crit Care Med* 42(2):433–444
40. Raman G, Avendano EE, Chan J, Merchant S, Puzniak L (2018) Risk factors for hospitalized patients with resistant or multidrug-resistant *Pseudomonas aeruginosa* infections: a systematic review and meta-analysis. *Antimicrob Resist Infect Control* 7:79
41. Zahar JR, Blot S, Nordmann P, Martischang R, Timsit JF, Harbarth S et al (2018) Screening for intestinal carriage of ESBL-producing *Enterobacteriaceae* in critically ill patients: expected benefits and evidence-based controversies. *Clin Infect Dis*. <https://doi.org/10.1093/cid/ciy864>
42. Mazzeffi M, Gammie J, Taylor B, Cardillo S, Haldane-Lutterodt N, Amoroso A et al (2017) Healthcare-associated infections in cardiac surgery patients with prolonged intensive care unit stay. *Ann Thorac Surg* 103(4):1165–1170
43. van Vught LA, Klein Klouwenberg PM, Spitoni C, Scicluna BP, Wiewel MA, Horn J et al (2016) Incidence, risk factors, and attributable mortality of secondary infections in the intensive care unit after admission for sepsis. *JAMA* 315(14):1469–1479
44. Tabah A, Koulenti D, Laupland K, Misset B, Valles J, de Bruzzi Carvalho F et al (2012) Characteristics and determinants of outcome of hospital-acquired bloodstream infections in intensive care units: the EURO-BACT International Cohort Study. *Intensive Care Med* 38(12):1930–1945
45. Sterling SA, Puskasich MA, Glass AF, Guirgis F, Jones AE (2017) The impact of the Sepsis-3 septic shock definition on previously defined septic shock patients. *Crit Care Med* 45(9):1436–1442
46. Ferrer R, Martinez ML, Goma G, Suarez D, Alvarez-Rocha L, de la Torre MV et al (2018) Improved empirical antibiotic treatment of sepsis after an educational intervention: the ABIS-Edusepsis study. *Crit Care* 22(1):167
47. Liu VX, Fielding-Singh V, Greene JD, Baker JM, Iwashyna TJ, Bhattacharya J et al (2017) The timing of early antibiotics and hospital mortality in sepsis. *Am J Respir Crit Care Med* 196(7):856–863
48. Seymour CW, Gesten F, Prescott HC, Friedrich ME, Iwashyna TJ, Phillips GS et al (2017) Time to treatment and mortality during mandated emergency care for sepsis. *N Engl J Med* 376(23):2235–2244
49. Rhodes A, Evans LE, Alhazzani W, Levy MM, Antonelli M, Ferrer R et al (2017) Surviving Sepsis Campaign: international guidelines for management of sepsis and septic shock: 2016. *Intensive Care Med* 43(3):304–377
50. Laupland KB, Zahar JR, Adrie C, Schwebel C, Goldgran-Toledano D, Azoulay E et al (2012) Determinants of temperature abnormalities and influence on outcome of critical illness. *Crit Care Med* 40(1):145–151
51. Lam SW, Bauer SR, Fowler R, Duggal A (2018) Systematic review and meta-analysis of procalcitonin-guidance versus usual care for antimicrobial management in critically ill patients: focus on subgroups based on antibiotic initiation, cessation, or mixed strategies. *Crit Care Med* 46(5):684–690
52. Schuetz P, Wirz Y, Sager R, Christ-Crain M, Stolz D, Tamm M et al (2018) Effect of procalcitonin-guided antibiotic treatment on mortality in acute respiratory infections: a patient level meta-analysis. *Lancet Infect Dis* 18(1):95–107
53. van Someren Greve F, Juffermans NP, Bos LDJ, Binnekade JM, Braber A, Cremer OL et al (2018) Respiratory viruses in invasively ventilated critically ill patients—a prospective multicenter observational study. *Crit Care Med* 46(1):29–36
54. Loubet P, Voiriot G, Houhou-Fidouh N, Neuville M, Bouadma L, Lescure FX et al (2017) Impact of respiratory viruses in hospital-acquired pneumonia in the intensive care unit: a single-center retrospective study. *J Clin Virol* 91:52–57
55. Singer M, Deutschman CS, Seymour CW, Shankar-Hari M, Annane D, Bauer M et al (2016) The third international consensus definitions for sepsis and septic shock (Sepsis-3). *JAMA* 315(8):801–810
56. Fernando SM, Tran A, Taljaard M, Cheng W, Rochwerf B, Seely AJE et al (2018) Prognostic accuracy of the quick sequential organ failure assessment for mortality in patients with suspected infection: a systematic review and meta-analysis. *Ann Intern Med* 168(4):266–275
57. Williams JM, Greenslade JH, McKenzie JV, Chu K, Brown AFT, Lipman J (2017) Systemic inflammatory response syndrome, quick sequential organ function assessment, and organ dysfunction: insights from a prospective database of ED patients with infection. *Chest* 151(3):586–596
58. Klein Klouwenberg PM, Cremer OL, van Vught LA, Ong DS, Frencken JF, Schultz MJ et al (2015) Likelihood of infection in patients with presumed sepsis at the time of intensive care unit admission: a cohort study. *Crit Care* 19:319
59. Hranjec T, Rosenberger LH, Swenson B, Metzger R, Flohr TR, Politano AD et al (2012) Aggressive versus conservative initiation of antimicrobial treatment in critically ill surgical patients with suspected intensive-care-unit-acquired infection: a quasi-experimental, before and after observational cohort study. *Lancet Infect Dis* 12(10):774–780
60. Parlato M, Cavillon JM (2015) Host response biomarkers in the diagnosis of sepsis: a general overview. *Methods Mol Biol* 1237:149–211
61. Parlato M, Philippart F, Rouquette A, Moucadel V, Puchos V, Blein S et al (2018) Circulating biomarkers may be unable to detect infection at the early phase of sepsis in ICU patients: the CAPTAIN prospective multicenter cohort study. *Intensive Care Med* 44(7):1061–1070
62. Layios N, Lambermont B, Canivet JL, Morimont P, Preiser JC, Garweg C et al (2012) Procalcitonin usefulness for the initiation of antibiotic treatment in intensive care unit patients. *Crit Care Med* 40(8):2304–2309
63. Mearelli F, Fiotti N, Giansante C, Casarsa C, Orso D, De Helmersen M et al (2018) Derivation and validation of a biomarker-based clinical


- algorithm to rule out sepsis from noninfectious systemic inflammatory response syndrome at emergency department admission: a multi-center prospective study. *Crit Care Med* 46(9):1421–1429
64. Kalil AC, Syed A, Rupp ME, Chambers H, Vargas L, Maskin A et al (2015) Is bacteremic sepsis associated with higher mortality in transplant recipients than in nontransplant patients? A matched case-control propensity-adjusted study. *Clin Infect Dis* 60(2):216–222
 65. Kalil AC, Sandkovsky U, Florescu DF (2018) Severe infections in critically ill solid organ transplant recipients. *Clin Microbiol Infect* 24:1257–1263
 66. NCCN (2018) Clinical practice guidelines in oncology. prevention and treatment of cancer-related infections. version 1.2018. https://www.nccn.org/store/login/login.aspx?ReturnURL=https://www.nccn.org/professionals/physician_gls/pdf/infections.pdf. Accessed 1 Sept 2018
 67. Schnell D, Azoulay E, Benoit D, Clouzeau B, Demaret P, Ducassou S et al (2016) Management of neutropenic patients in the intensive care unit (NEWBORNS EXCLUDED) recommendations from an expert panel from the French Intensive Care Society (SRLF) with the French Group for Pediatric Intensive Care Emergencies (GFRUP), the French Society of Anesthesia and Intensive Care (SFAR), the French Society of Hematology (SFH), the French Society for Hospital Hygiene (SF2H), and the French Infectious Diseases Society (SPLIF). *Ann Intensive Care* 6(1):90
 68. European AIDS Clinical Society guidelines. Updated yearly. <http://www.eacsociety.org/Guidelines.aspx>. Accessed 1 Sept 2018
 69. Singh JA, Cameron C, Noorbaloochi S, Cullis T, Tucker M, Christensen R et al (2015) Risk of serious infection in biological treatment of patients with rheumatoid arthritis: a systematic review and meta-analysis. *Lancet* 386(9990):258–265
 70. Venet F, Monneret G (2018) Advances in the understanding and treatment of sepsis-induced immunosuppression. *Nat Rev Nephrol* 14(2):121–137
 71. Martin S, Perez A, Aldecoa C (2017) Sepsis and immunosenescence in the elderly patient: a review. *Front Med* 4:20
 72. Ramanan P, Bryson AL, Binnicker MJ, Pritt BS, Patel R (2018) Syndromic panel-based testing in clinical microbiology. *Clin Microbiol Rev* 31(1):e00024-17
 73. van de Groep K, Bos MP, Savelkoul PHM, Rubenjan A, Gazenbeek C, Melchers WJG et al (2018) Development and first evaluation of a novel multiplex real-time PCR on whole blood samples for rapid pathogen identification in critically ill patients with sepsis. *Eur J Clin Microbiol Infect Dis* 37(7):1333–1344
 74. Ruppe E, Baud D, Schicklin S, Guigon G, Schrenzel J (2016) Clinical metagenomics for the management of hospital- and healthcare-acquired pneumonia. *Future Microbiol* 11(3):427–439
 75. Charalampous T, Richardson H, Kay GL, Baldan R, Jeanes C, Rae D et al (2018) Rapid diagnosis of lower respiratory infection using nanopore-based clinical metagenomics. *BioRxiv*. <https://doi.org/10.1101/387548>
 76. Pendleton KM, Erb-Downward JR, Bao Y, Branton WR, Falkowski NR, Newton DW et al (2017) Rapid pathogen identification in bacterial pneumonia using real-time metagenomics. *Am J Respir Crit Care Med* 196(12):1610–1612
 77. Langelier C, Zinter MS, Kalantar K, Yanik GA, Christenson S, O'Donovan B et al (2018) Metagenomic sequencing detects respiratory pathogens in hematopoietic cellular transplant patients. *Am J Respir Crit Care Med* 197(4):524–528
 78. Le Dorze M, Gault N, Foucrier A, Ruppe E, Mourvillier B, Woerther PL et al (2015) Performance and impact of a rapid method combining mass spectrometry and direct antimicrobial susceptibility testing on treatment adequacy of patients with ventilator-associated pneumonia. *Clin Microbiol Infect* 21(5):468
 79. Lutgring JD, Bittencourt C, McElvania TeKippe E, Cavuoti D, Hollaway R, Burd EM (2018) Evaluation of the accelerate pheno system: results from two academic medical centers. *J Clin Microbiol* 56(4):e01672-17
 80. Timbrook TT, Morton JB, McConeghy KW, Caffrey AR, Mylonakis E, LaPlante KL (2017) The effect of molecular rapid diagnostic testing on clinical outcomes in bloodstream infections: a systematic review and meta-analysis. *Clin Infect Dis* 64(1):15–23
 81. Roberts JA, Paul SK, Akova M, Bassetti M, De Waele JJ, Dimopoulos G et al (2014) DALI: defining antibiotic levels in intensive care unit patients: are current beta-lactam antibiotic doses sufficient for critically ill patients? *Clin Infect Dis* 58(8):1072–1083
 82. Roberts JA, Taccone FS, Lipman J (2016) Understanding PK/PD. *Intensive Care Med* 42(11):1797–1800
 83. De Waele JJ, Lipman J, Carlier M, Roberts JA (2015) Subtleties in practical application of prolonged infusion of beta-lactam antibiotics. *Int J Antimicrob Agents* 45(5):461–463
 84. Bergen PJ, Bulitta JB, Kirkpatrick CM, Rogers KE, McGregor MJ, Wallis SC et al (2017) Substantial impact of altered pharmacokinetics in critically ill patients on the antibacterial effects of meropenem evaluated via the dynamic hollow-fiber infection model. *Antimicrob Agents Chemother* 61(5):e02642-16
 85. Tangden T, Ramos Martin V, Felton TW, Nielsen EI, Marchand S, Bruggemann RJ et al (2017) The role of infection models and PK/PD modelling for optimising care of critically ill patients with severe infections. *Intensive Care Med* 43(7):1021–1032
 86. Huttner A, Harbarth S, Hope WW, Lipman J, Roberts JA (2015) Therapeutic drug monitoring of the beta-lactam antibiotics: what is the evidence and which patients should we be using it for? *J Antimicrob Chemother* 70(12):3178–3183
 87. Duszynska W, Taccone FS, Hurkacz M, Kowalska-Krochmal B, Wiela-Hojenska A, Kubler A (2013) Therapeutic drug monitoring of amikacin in septic patients. *Crit Care* 17(4):R165
 88. Brasseur A, Hites M, Roisin S, Cotton F, Vincent JL, De Backer D et al (2016) A high-dose aminoglycoside regimen combined with renal replacement therapy for the treatment of MDR pathogens: a proof-of-concept study. *J Antimicrob Chemother* 71(5):1386–1394
 89. Pajot O, Burdet C, Couffignal C, Massias L, Armand-Lefevre L, Foucrier A et al (2015) Impact of imipenem and amikacin pharmacokinetic/pharmacodynamic parameters on microbiological outcome of Gram-negative bacilli ventilator-associated pneumonia. *J Antimicrob Chemother* 70(5):1487–1494
 90. Prybylski JP (2015) Vancomycin trough concentration as a predictor of clinical outcomes in patients with *Staphylococcus aureus* bacteremia: a meta-analysis of observational studies. *Pharmacotherapy* 35(10):889–898
 91. Wong G, Brinkman A, Benefield RJ, Carlier M, De Waele JJ, El Helali N et al (2014) An international, multicentre survey of beta-lactam antibiotic therapeutic drug monitoring practice in intensive care units. *J Antimicrob Chemother* 69(5):1416–1423
 92. Mouton RW, Muller AE, Canton R, Giske CG, Kahlmeter G, Turnidge J (2018) MIC-based dose adjustment: facts and fables. *J Antimicrob Chemother* 73(3):564–568
 93. Beumier M, Casu GS, Hites M, Wolff F, Cotton F, Vincent JL et al (2015) Elevated beta-lactam concentrations associated with neurological deterioration in ICU septic patients. *Minerva Anestesiol* 81(5):497–506
 94. Imani S, Buscher H, Marriott D, Gentili S, Sandaradura I (2017) Too much of a good thing: a retrospective study of beta-lactam concentration-toxicity relationships. *J Antimicrob Chemother* 72(10):2891–2897
 95. Torres A, Zhong N, Pachi J, Timsit JF, Kollef M, Chen Z et al (2018) Ceftazidime-avibactam versus meropenem in nosocomial pneumonia, including ventilator-associated pneumonia (REPROVE): a randomised, double-blind, phase 3 non-inferiority trial. *Lancet Infect Dis* 18(3):285–295
 96. Harris PNA, Tambyah PA, Lye DC, Mo Y, Lee TH, Yilmaz M et al (2018) Effect of piperacillin-tazobactam vs meropenem on 30-day mortality for patients with *E. coli* or *Klebsiella pneumoniae* bloodstream infection and ceftriaxone resistance: a randomized clinical trial. *JAMA* 320(10):984–994
 97. Bertolini G, Nattino G, Tascini C, Poole D, Viaggi B, Carrara G et al (2018) Mortality attributable to different *Klebsiella* susceptibility patterns and to the coverage of empirical antibiotic therapy: a cohort study on patients admitted to the ICU with infection. *Intensive Care Med* 44(10):1709–1719
 98. Murray KP, Zhao JJ, Davis SL, Kullar R, Kaye KS, Lephart P et al (2013) Early use of daptomycin versus vancomycin for methicillin-resistant *Staphylococcus aureus* bacteremia with vancomycin minimum inhibitory concentration > 1 mg/L: a matched cohort study. *Clin Infect Dis* 56(11):1562–1569
 99. Wunderink RG, Niederman MS, Kollef MH, Shorr AF, Kunkel MJ, Baruch A et al (2012) Linezolid in methicillin-resistant *Staphylococcus aureus* nosocomial pneumonia: a randomized, controlled study. *Clin Infect Dis* 54(5):621–629

100. Abbas M, Paul M, Huttner A (2017) New and improved? A review of novel antibiotics for Gram-positive bacteria. *Clin Microbiol Infect* 23(10):697–703
101. Tamma PD, Cosgrove SE, Maragakis LL (2012) Combination therapy for treatment of infections with gram-negative bacteria. *Clin Microbiol Rev* 25(3):450–470
102. Paul M, Lador A, Grozinsky-Glasberg S, Leibovici L (2014) Beta lactam antibiotic monotherapy versus beta lactam-aminoglycoside antibiotic combination therapy for sepsis. *Cochrane Database Syst Rev* 1:CD003344
103. Kumar A, Safdar N, Kethireddy S, Chateau D (2010) A survival benefit of combination antibiotic therapy for serious infections associated with sepsis and septic shock is contingent only on the risk of death: a meta-analytic/meta-regression study. *Crit Care Med* 38(8):1651–1664
104. Ripa M, Rodriguez-Nunez O, Cardozo C, Naharro-Abellan A, Almela M, Marco F et al (2017) Influence of empirical double-active combination antimicrobial therapy compared with active monotherapy on mortality in patients with septic shock: a propensity score-adjusted and matched analysis. *J Antimicrob Chemother* 72(12):3443–3452
105. Daikos GL, Tsaousi S, Tzouveleki LS, Anyfantis I, Psychogiou M, Argyropoulou A et al (2014) Carbapenemase-producing *Klebsiella pneumoniae* bloodstream infections: lowering mortality by antibiotic combination schemes and the role of carbapenems. *Antimicrob Agents Chemother* 58(4):2322–2328
106. Tumbarello M, Trecarichi EM, De Rosa FG, Giannella M, Giacobbe DR, Bassetti M et al (2015) Infections caused by KPC-producing *Klebsiella pneumoniae*: differences in therapy and mortality in a multicentre study. *J Antimicrob Chemother* 70(7):2133–2143
107. Falcone M, Russo A, Iacovelli A, Restuccia G, Ceccarelli G, Giordano A et al (2016) Predictors of outcome in ICU patients with septic shock caused by *Klebsiella pneumoniae* carbapenemase-producing *K. pneumoniae*. *Clin Microbiol Infect* 22(5):444–450
108. Gutierrez-Gutierrez B, Salamanca E, de Cueto I, Hsueh PR, Viale P, Pano-Pardo JR et al (2017) Effect of appropriate combination therapy on mortality of patients with bloodstream infections due to carbapenemase-producing Enterobacteriaceae (INCREMENT): a retrospective cohort study. *Lancet Infect Dis* 17(7):726–734
109. Tumbarello M, Viale P, Viscoli C, Trecarichi EM, Tumietto F, Marchese A et al (2012) Predictors of mortality in bloodstream infections caused by *Klebsiella pneumoniae* carbapenemase-producing *K. pneumoniae*: importance of combination therapy. *Clin Infect Dis* 55(7):943–950
110. Aydemir H, Akduman D, Piskin N, Comert F, Horuz E, Terzi A et al (2013) Colistin vs. the combination of colistin and rifampicin for the treatment of carbapenem-resistant *Acinetobacter baumannii* ventilator-associated pneumonia. *Epidemiol Infect* 141(6):1214–1222
111. Durante-Mangoni E, Signorile G, Andini R, Mattei A, De Cristoforo M, Murino P et al (2013) Colistin and rifampicin compared with colistin alone for the treatment of serious infections due to extensively drug-resistant *Acinetobacter baumannii*: a multicenter, randomized clinical trial. *Clin Infect Dis* 57(3):349–358
112. Sirijatuphat R, Thamlikitkul V (2014) Preliminary study of colistin versus colistin plus fosfomycin for treatment of carbapenem-resistant *Acinetobacter baumannii* infections. *Antimicrob Agents Chemother* 58(9):5598–5601
113. Paul M, Daikos GL, Durante-Mangoni E, Yahav D, Carmeli Y, Benattar YD et al (2018) Colistin alone versus colistin plus meropenem for treatment of severe infections caused by carbapenem-resistant gram-negative bacteria: an open-label, randomised controlled trial. *Lancet Infect Dis* 18(4):391–400
114. Zusman O, Altunin S, Koppel F, Dishon Benattar Y, Gedik H, Paul M (2017) Polymyxin monotherapy or in combination against carbapenem-resistant bacteria: systematic review and meta-analysis. *J Antimicrob Chemother* 72(1):29–39
115. Makris D, Petinaki E, Tsolaki V, Manoulakas E, Mantzaris K, Apostolopoulou O et al (2018) Colistin versus colistin combined with ampicillin-sulbactam for multidrug-resistant *Acinetobacter baumannii* ventilator-associated pneumonia treatment: an open-label prospective study. *Indian J Crit Care Med* 22(2):67–77
116. Vardakas KZ, Mavroudis AD, Georgiou M, Falagas ME (2018) Intravenous colistin combination antimicrobial treatment vs. monotherapy: a systematic review and meta-analysis. *Int J Antimicrob Agents* 51(4):535–547
117. Sime FB, Roberts MS, Roberts JA (2015) Optimization of dosing regimens and dosing in special populations. *Clin Microbiol Infect* 21(10):886–893
118. Vardakas KZ, Voulgaris GL, Maliaros A, Samonis G, Falagas ME (2018) Prolonged versus short-term intravenous infusion of antipseudomonal beta-lactams for patients with sepsis: a systematic review and meta-analysis of randomised trials. *Lancet Infect Dis* 18(1):108–120
119. Hao JJ, Chen H, Zhou JX (2016) Continuous versus intermittent infusion of vancomycin in adult patients: a systematic review and meta-analysis. *Int J Antimicrob Agents* 47(1):28–35
120. Tabah A, Cotta MO, Garnacho-Montero J, Schouten J, Roberts JA, Lipman J et al (2016) A systematic review of the definitions, determinants, and clinical outcomes of antimicrobial de-escalation in the intensive care unit. *Clin Infect Dis* 62(8):1009–1017
121. Weiss E, Zahar JR, Lesprit P, Ruppe E, Leone M, Chastre J et al (2015) Elaboration of a consensual definition of de-escalation allowing a ranking of beta-lactams. *Clin Microbiol Infect* 21(7):649
122. Silva BN, Andriolo RB, Atallah AN, Salomao R (2013) De-escalation of antimicrobial treatment for adults with sepsis, severe sepsis or septic shock. *Cochrane Database Syst Rev* 3:CD007934
123. Mokart D, Slehofer G, Lambert J, Sannini A, Chow-Chine L, Brun JP et al (2014) De-escalation of antimicrobial treatment in neutropenic patients with severe sepsis: results from an observational study. *Intensive Care Med* 40(1):41–49
124. Paul M, Dickstein Y, Raz-Pasteur A (2016) Antibiotic de-escalation for bloodstream infections and pneumonia: systematic review and meta-analysis. *Clin Microbiol Infect* 22(12):960–967
125. Leone M, Bechis C, Baumstarck K, Lefrant JY, Albanese J, Jaber S et al (2014) De-escalation versus continuation of empirical antimicrobial treatment in severe sepsis: a multicenter non-blinded randomized noninferiority trial. *Intensive Care Med* 40(10):1399–1408
126. Weiss E, Zahar JR, Garrouste-Orgeas M, Ruckly S, Essaiad W, Schwebel C et al (2016) De-escalation of pivotal beta-lactam in ventilator-associated pneumonia does not impact outcome and marginally affects MDR acquisition. *Intensive Care Med* 42(12):2098–2100
127. De Bus L, Denys W, Catteeuw J, Gadeyne B, Vermeulen K, Boelens J et al (2016) Impact of de-escalation of beta-lactam antibiotics on the emergence of antibiotic resistance in ICU patients: a retrospective observational study. *Intensive Care Med* 42(6):1029–1039
128. Ruppe E, Martin-Loeches I, Rouze A, Levast B, Ferry T, Timsit JF (2018) What's new in restoring the gut microbiota in ICU patients? Potential role of faecal microbiota transplantation. *Clin Microbiol Infect* 24(8):803–805
129. D'Agata EM, Magal P, Olivier D, Ruan S, Webb GF (2007) Modeling antibiotic resistance in hospitals: the impact of minimizing treatment duration. *J Theor Biol* 249(3):487–499
130. Chastre J, Wolff M, Fagon JY, Chevret S, Thomas F, Wermert D et al (2003) Comparison of 8 vs 15 days of antibiotic therapy for ventilator-associated pneumonia in adults: a randomized trial. *JAMA* 290(19):2588–2598
131. Klompas M, Li L, Menchaca JT, Gruber S (2017) Ultra-short-course antibiotics for patients with suspected ventilator-associated pneumonia but minimal and stable ventilator settings. *Clin Infect Dis* 64(7):870–876
132. Sandberg T, Skoog G, Hermansson AB, Kahlmeter G, Kuylenstierna N, Lannergard A et al (2012) Ciprofloxacin for 7 days versus 14 days in women with acute pyelonephritis: a randomised, open-label and double-blind, placebo-controlled, non-inferiority trial. *Lancet* 380(9840):484–490
133. Chotiprasitsakul D, Han JH, Cosgrove SE, Harris AD, Lautenbach E, Conley AT et al (2018) Comparing the outcomes of adults with enterobacteriaceae bacteremia receiving short-course versus prolonged-course antibiotic therapy in a multicenter, propensity score-matched cohort. *Clin Infect Dis* 66(2):172–177
134. Montravers P, Tubach F, Lescot T, Veber B, Esposito-Faresse M, Seguin P et al (2018) Short-course antibiotic therapy for critically ill patients treated for postoperative intra-abdominal infection: the DURAPOP randomised clinical trial. *Intensive Care Med* 44(3):300–310
135. Royer S, DeMerle KM, Dickson RP, Prescott HC (2018) Shorter versus longer courses of antibiotics for infection in hospitalized patients: a systematic review and meta-analysis. *J Hosp Med* 13(5):336–342

136. Hanretty AM, Gallagher JC (2018) Shortened courses of antibiotics for bacterial infections: a systematic review of randomized controlled trials. *Pharmacotherapy* 38(6):674–687
137. Huang DT, Yealy DM, Filbin MR, Brown AM, Chang CH, Doi Y et al (2018) Procalcitonin-guided use of antibiotics for lower respiratory tract infection. *N Engl J Med* 379(3):236–249
138. Mazuski JE, Tessier JM, May AK, Sawyer RG, Nadler EP, Rosengart MR et al (2017) The surgical infection society revised guidelines on the management of intra-abdominal infection. *Surg Infect* 18(1):1–76
139. Woodhead M, Blasi F, Ewig S, Garau J, Huchon G, Ieven M et al (2011) Guidelines for the management of adult lower respiratory tract infections—full version. *Clin Microbiol Infect* 17(Suppl 6):E1–E59
140. Mandell LA, Wunderink RG, Anzueto A, Bartlett JG, Campbell GD, Dean NC et al (2007) Infectious Diseases Society of America/American Thoracic Society consensus guidelines on the management of community-acquired pneumonia in adults. *Clin Infect Dis* 44(Suppl 2):S27–S72
141. Gupta K, Hooton TM, Naber KG, Wullt B, Colgan R, Miller LG et al (2011) International clinical practice guidelines for the treatment of acute uncomplicated cystitis and pyelonephritis in women: a 2010 update by the Infectious Diseases Society of America and the European Society for Microbiology and Infectious Diseases. *Clin Infect Dis* 52(5):e103–e120
142. Torres A, Niederman MS, Chastre J, Ewig S, Fernandez-Vandellos P, Hanberger H et al (2017) International ERS/ESICM/ESCMID/ALAT guidelines for the management of hospital-acquired pneumonia and ventilator-associated pneumonia: Guidelines for the management of hospital-acquired pneumonia (HAP)/ventilator-associated pneumonia (VAP) of the European Respiratory Society (ERS), European Society of Intensive Care Medicine (ESICM), European Society of Clinical Microbiology and Infectious Diseases (ESCMID) and Asociacion Latinoamericana del Torax (ALAT). *Eur Respir J* 50(3):1700582
143. Klein EY, Van Boeckel TP, Martinez EM, Pant S, Gandra S, Levin SA et al (2018) Global increase and geographic convergence in antibiotic consumption between 2000 and 2015. *Proc Natl Acad Sci USA* 115(15):E3463–E3470
144. ECDC. Summary of the latest data on antibiotic consumption in the European Union ESAC-Net surveillance data November 2017. https://ecdc.europa.eu/sites/portal/files/documents/Final_2017_EAAD_ESAC-Net_Summary-edited%20-%20FINALwith%20erratum.pdf2017. Accessed 13 Oct 2018
145. Kollef MH, Chastre J, Clavel M, Restrepo MI, Michiels B, Kaniga K et al (2012) A randomized trial of 7-day doripenem versus 10-day imipenem-cilastatin for ventilator-associated pneumonia. *Crit Care* 16(6):R218
146. Schein M, Marshall J (2004) Source control for surgical infections. *World J Surg* 28(7):638–645
147. Martinez ML, Ferrer R, Torrents E, Guillaumat-Prats R, Goma G, Suarez D et al (2017) Impact of source control in patients with severe sepsis and septic shock. *Crit Care Med* 45(1):1–19
148. Chao WN, Tsai CF, Chang HR, Chan KS, Su CH, Lee YT et al (2013) Impact of timing of surgery on outcome of *Vibrio vulnificus*-related necrotizing fasciitis. *Am J Surg* 206(1):32–39
149. Karvellas CJ, Abalde JG, Zepeda-Gomez S, Moffat DC, Mirzanejad Y, Vazquez-Grande G et al (2016) The impact of delayed biliary decompression and anti-microbial therapy in 260 patients with cholangitis-associated septic shock. *Aliment Pharmacol Ther* 44(7):755–766
150. Bloos F, Ruddel H, Thomas-Ruddel D, Schwarzkopf D, Pausch C, Harbarth S et al (2017) Effect of a multifaceted educational intervention for anti-infectious measures on sepsis mortality: a cluster randomized trial. *Intensive Care Med* 43(11):1602–1612
151. Bloos F, Thomas-Ruddel D, Ruddel H, Engel C, Schwarzkopf D, Marshall JC et al (2014) Impact of compliance with infection management guidelines on outcome in patients with severe sepsis: a prospective observational multi-center study. *Crit Care* 18(2):R42
152. Azuhata T, Kinoshita K, Kawano D, Komatsu T, Sakurai A, Chiba Y et al (2014) Time from admission to initiation of surgery for source control is a critical determinant of survival in patients with gastrointestinal perforation with associated septic shock. *Crit Care* 18(3):R87
153. Boyer A, Vargas F, Coste F, Saubusse E, Castaing Y, Gbikpi-Benissan G et al (2009) Influence of surgical treatment timing on mortality from necrotizing soft tissue infections requiring intensive care management. *Intensive Care Med* 35(5):847–853
154. Karanika S, Paudel S, Grigoras C, Kalbasi A, Mylonakis E (2016) Systematic review and meta-analysis of clinical and economic outcomes from the implementation of hospital-based antimicrobial stewardship programs. *Antimicrob Agents Chemother* 60(8):4840–4852
155. Baur D, Gladstone BP, Burkert F, Carrara E, Foschi F, Dobe S et al (2017) Effect of antibiotic stewardship on the incidence of infection and colonisation with antibiotic-resistant bacteria and *Clostridium difficile* infection: a systematic review and meta-analysis. *Lancet Infect Dis* 17(9):990–1001
156. Dyar OJ, Huttner B, Schouten J, Pulcini C (2017) What is antimicrobial stewardship? *Clin Microbiol Infect* 23(11):793–798
157. Pulcini C, Binda F, Lamkang AS, Trett A, Charani E, Goff DA et al (2018) Developing core elements and checklist items for global hospital antimicrobial stewardship programmes: a consensus approach. *Clin Microbiol Infect* 25(1):20–25
158. Kollef MH, Bassetti M, Francois B, Burnham J, Dimopoulos G, Garnacho-Montero J et al (2017) The intensive care medicine research agenda on multidrug-resistant bacteria, antibiotics, and stewardship. *Intensive Care Med* 43(9):1187–1197
159. De Waele JJ, Akova M, Antonelli M, Canton R, Carlet J, De Backer D et al (2018) Antimicrobial resistance and antibiotic stewardship programs in the ICU: insistence and persistence in the fight against resistance. A position statement from ESICM/ESCMID/WAAAR round table on multi-drug resistance. *Intensive Care Med* 44(2):189–196
160. Flottorp SA, Oxman AD, Krause J, Musila NR, Wensing M, Godycki-Cwirko M et al (2013) A checklist for identifying determinants of practice: a systematic review and synthesis of frameworks and taxonomies of factors that prevent or enable improvements in healthcare professional practice. *Implement Sci* 8:35
161. Bailly S, Meyfroidt G, Timsit JF (2018) What's new in ICU in 2050: big data and machine learning. *Intensive Care Med* 44(9):1524–1527
162. Salluh JF, Chiche JD, Reis CE, Soares M (2018) New perspectives to improve critical care benchmarking. *Ann Inten Care* 8(1):17
163. Naidus E, Celi LA (2016) Big data in healthcare: are we close to it? *Rev Bras Ter Intensiva* 28(1):8–10
164. Bremmer DN, Trienski TL, Walsh TL, Moffa MA (2018) Role of technology in antimicrobial stewardship. *Med Clin N Am* 102(5):955–963
165. Emberger J, Tassone D, Stevens MP, Markley JD (2018) The current state of antimicrobial stewardship: challenges, successes, and future directions. *Curr Infect Dis Rep* 20(9):31
166. Doernberg SB, Chambers HF (2017) Antimicrobial stewardship approaches in the intensive care unit. *Infect Dis Clin N Am* 31(3):513–534
167. Teerawattanapong N, Kengkla K, Dilokthornsakul P, Saokaew S, Apisarnthanarak A, Chaiyakunapruk N (2017) Prevention and control of multidrug-resistant Gram-negative bacteria in adult intensive care units: a systematic review and network meta-analysis. *Clin Infect Dis* 64(suppl_2):S51–S60
168. Ruppe E, Lisboa T, Barbier F (2018) The gut microbiota of critically ill patients: first steps in an unexplored world. *Intensive Care Med* 44:1561–1564
169. de Gunzburg J, Ghazlane A, Ducher A, Le Chatelier E, Duval X, Ruppe E et al (2018) Protection of the human gut microbiome from antibiotics. *J Infect Dis* 217(4):628–636
170. Haak BW, Wiersinga WJ (2017) The role of the gut microbiota in sepsis. *Lancet Gastroenterol Hepatol* 2(2):135–143
171. World Health Organization. Antimicrobial resistance—Global report of surveillance. <http://www.who.int/antimicrobial-resistance/publications/surveillancereport/en/>. 2014
172. European Center for Disease Control and Prevention. European Antimicrobial Resistance Surveillance Network (EARS-Net)—Annual report. http://www.ecdc.europa.eu/en/healthtopics/antimicrobial_resistance. 2016
173. Luyt CE, Brechot N, Trouillet JL, Chastre J (2014) Antibiotic stewardship in the intensive care unit. *Crit Care* 18(5):480
174. Alobaid AS, Hites M, Lipman J, Taccone FS, Roberts JA (2016) Effect of obesity on the pharmacokinetics of antimicrobials in critically ill patients: a structured review. *Int J Antimicrob Agents* 47(4):259–268

ORIGINAL

Cardiovascular clusters in septic shock combining clinical and echocardiographic parameters: a post hoc analysis

Guillaume Geri^{1,2,3} , Philippe Vignon^{4,5,6}, Alix Aubry^{1,2}, Anne-Laure Fedou⁴, Cyril Charron¹, Stein Silva⁷, Xavier Repesse¹ and Antoine Vieillard-Baron^{1,2,3*}

© 2019 Springer-Verlag GmbH Germany, part of Springer Nature

Abstract

Purpose: Mechanisms of circulatory failure are complex and frequently intricate in septic shock. Better characterization could help to optimize hemodynamic support.

Methods: Two published prospective databases from 12 different ICUs including echocardiographic monitoring performed by a transesophageal route at the initial phase of septic shock were merged for post hoc analysis. Hierarchical clustering in a principal components approach was used to define cardiovascular phenotypes using clinical and echocardiographic parameters. Missing data were imputed.

Findings: A total of 360 patients (median age 64 [55; 74]) were included in the analysis. Five different clusters were defined: patients well resuscitated (cluster 1, $n=61$, 16.9%) without left ventricular (LV) systolic dysfunction, right ventricular (RV) failure or fluid responsiveness, patients with LV systolic dysfunction (cluster 2, $n=64$, 17.7%), patients with hyperkinetic profile (cluster 3, $n=84$, 23.3%), patients with RV failure (cluster 4, $n=81$, 22.5%) and patients with persistent hypovolemia (cluster 5, $n=70$, 19.4%). Day 7 mortality was 9.8%, 32.8%, 8.3%, 27.2%, and 23.2%, while ICU mortality was 21.3%, 50.0%, 23.8%, 42.0%, and 38.6% in clusters 1, 2, 3, 4, and 5, respectively ($p<0.001$ for both).

Conclusion: Our clustering approach on a large population of septic shock patients, based on clinical and echocardiographic parameters, was able to characterize five different cardiovascular phenotypes. How this could help physicians to optimize hemodynamic support should be evaluated in the future.

Keywords: Septic shock, Hemodynamic failure, Cluster

Introduction

In the past, it was considered that hemodynamic alterations in septic shock occur in different phases, an early phase with a low flow state related to hypovolemia, a second phase after the initial resuscitation with a hyperdynamic state, and finally a third phase with cardiac failure

leading to multiorgan failure and death [1]. Since the landmark study by Parker et al. [2], it has been progressively accepted that depressed left ventricular (LV) systolic function may develop at the early phase of septic shock. For a long time, the assessment of hemodynamic instability in septic shock was based on right heart catheterization, while alternatives are now well recognized, including critical care echocardiography (CCE) [3]. The high incidence of early myocardial alterations has been confirmed by echocardiographic studies [4, 5] and one found an incidence of LV systolic dysfunction of 39% during the first day [6]. There is then an urgent need for better characterization of cardiovascular phenotypes

*Correspondence: antoine.vieillard-baron@aphp.fr

¹ Medical-Surgical Intensive Care Unit, Ambroise Paré University Hospital, APHP, 9 avenue Charles de Gaulle, 92100 Boulogne-Billancourt, France
Full author information is available at the end of the article

in order to propose targeted/personalized medicine for hemodynamic support [7], as for the adequate need for fluids [8, 9] or inotropic support. Although CCE may detect combined mechanisms of circulatory failure in septic shock (i.e., vasoplegia, hypovolemia, LV systolic dysfunction, and right ventricular (RV) failure) [10], it failed to extend such an integrative and individualized approach in daily clinical practice. Cardiovascular phenotypes are insufficiently characterized and rely on too simplistic and inadequate definitions, as reflected by the binary approach mostly based on the value of LV ejection fraction (EF) to identify sepsis-induced LV systolic dysfunction [11].

We therefore hypothesized that the application of a clustering approach to a large database of septic shock patients monitored by CCE could help to better characterize the different cardiovascular phenotypes.

Materials and methods

Study design

The databases of two recently published prospective, observational, multicenter studies using CCE during early resuscitation of patients with septic shock were merged. The Hemosepsis study (inclusion January 2011–December 2013) compared the identification of hemodynamic profiles using both CCE and transpulmonary thermodilution in septic shock patients in sinus rhythm [12]. The Hemopred study included patients (November 2012–November 2014) with shock of any origin, mostly related to sepsis, to compare the accuracy of the different parameters of fluid responsiveness [13]. In both cohorts, we excluded patients with a history of chronic heart failure. Overall, the patients included came from 12 different ICUs. Septic shock was not defined by the Sepsis-3 definition as Hemopred and Hemosepsis were designed before its publication. Diagnosis was based on a suspected infection responsible for sustained hypotension despite adequate fluid loading that required vasopressors, with associated clinical signs of tissue hypoperfusion (mottled skin, encephalopathy, oliguria for more than 2 h) that were biologically confirmed (pH < 7.38 and base deficit > −5 mmol/L or lactate > 2 mmol/L or central venous oxygen saturation < 70%).

CCE

CCE was performed using a transesophageal route (TEE) in all patients during the first 12 h following the diagnosis of septic shock, after initial fluid resuscitation and vasopressor infusion. Patients were all intubated, sedated, and perfectly adapted to the respirator, as no spontaneous effort was observed during the echo procedure. Views, recorded parameters, and measurements were extensively described previously following the same

Take-home message

Using a clustering approach including clinical and echocardiographic parameters, 5 different hemodynamic phenotypes were identified in 360 septic shock patients, left ventricular (LV) systolic dysfunction, LV hyperkinesia, still hypovolemia, right ventricular failure and well-resuscitated phenotype.

prospective procedure [12, 13]. Echo parameters are all well recognized in the literature as good/adequate parameters of cardiovascular status in sepsis. Briefly, we systematically measured parameters of LV systolic function, i.e., LVEF and LV fractional area change (FAC) [14], and of LV diastolic function, i.e., maximal mitral Doppler *E* wave velocity and maximal tissue Doppler velocity of the lateral aspect of the mitral annulus at early diastole (*E'*) [15]. RV function was evaluated by the RV/LV end-diastolic area (EDA) ratio [16]. Fluid responsiveness was assessed using the superior vena cava collapsibility index (Δ SVC) [17]. We measured the velocity time integral (VTI) in the LV outflow tract and the diameter of the aortic annulus, which allowed us to calculate LV stroke volume and cardiac index (CI) [18]. The ultrasound systems used in the two cohorts were the same in each participating center as a result of the relatively narrow period of time encompassing the two studies. Images were all obtained and interpreted by intensivists trained in advanced level CCE, as mentioned in the two original studies. Images were not “validated” by independent experts to increase the external validity of our results which correspond to hemodynamic data obtained during daily echocardiographic assessment on clinical grounds. Importantly, no a priori criteria of “abnormality” were applied for the different parameters to best take into account inter-individual variabilities, allowing better characterization of cardiovascular phenotypes.

Patient characteristics and clinical hemodynamic evaluation

We calculated the sequential organ failure assessment (SOFA) score and the simplified acute physiology score (SAPS II). Mortality at day 7 and in the ICU was also recorded, as was the origin of infection.

In each patient, several clinical hemodynamic parameters were prospectively recorded at the time of the CCE: heart rate, invasive systolic (SAP), diastolic (DAP), and mean arterial pressure (MAP); central venous pressure (CVP) and central venous oxygen saturation (ScVO₂) were measured through a catheter placed in the internal jugular or subclavian vein; serum lactate level, volume of initial filling, presence of epinephrine, dobutamine, or norepinephrine and if so respective doses were also

recorded. We also recorded blood gas analysis and respiratory settings.

Statistical analysis

Baseline characteristics were reported as median [interquartile range] and n (%) for quantitative and qualitative variables, respectively. Quantitative variables were compared using nonparametric tests, the Mann–Whitney test or the Kruskal–Wallis test, as appropriate. Qualitative variables were compared using Pearson’s Chi-square test or Fisher’s exact test, as appropriate.

A two-step clustering approach was used to (1) reduce the dimensionality of the dataset and (2) to perform hierarchical clustering. This approach, so-called hierarchical clustering on principal components (HCPC), was performed using the *factoMineR* package in R [19]. We first performed a principal component analysis including hemodynamic parameters (i.e., LVEF, LVFAC, mitral Doppler *E* wave velocity, lateral mitral tissue Doppler *E'* velocity, aortic VTI, RV/LV EDA, Δ SVC, systolic arterial blood pressure, diastolic arterial blood pressure, heart rate, norepinephrine and epinephrine infusion doses). Variables were standardized as they were measured in different units. The HCPC procedure allows one, after the hierarchical clustering step is performed, to choose the number of clusters based on the hierarchical tree and to perform a K-means clustering to improve the initial partition obtained from the hierarchical clustering [20]. Agglomerative hierarchical clustering used the Ward’s criterion and an Euclidean metric. Data on the internal validity and stability of the analysis are shown in the supplementary material (Fig. S1). Missing data were imputed using iterative principal component analysis (implemented in the *imputePCA* R function), as previously described. Briefly, this method starts using a mean imputation, performs principal component analysis on the completed dataset, and missing values are then updated by the fitted values using a predefined number of dimensions [21]. Multiple imputations were also performed to visualize the variability related to the imputation process (Fig. S2).

We then compared variables (hemodynamic and non-hemodynamic-related variables) according to clusters. We chose not to include hemodynamic variables whose proportion of missing values was higher than 10%.

Last, we evaluated the diagnostic performance of the three most important variables for each cluster using two methods: (1) evaluation of the area under the receiver operating curve (AUROC) of a multivariable logistic regression with a binary variable of being in each cluster (yes/no) as the dependent variable and these three variables as independent variables and (2) calculation of

sensitivity, specificity, and negative and positive predictive values for the combination of these three variables above or below the thresholds. These thresholds were picked up from the description of the hemodynamic parameters we described across clusters as follows: the first interquartile (Q1) when the mean of the cluster was lower than the overall mean and the third interquartile (Q3) when the mean of the cluster was higher than the overall mean; 95% confidence intervals were calculated for all these results.

A p value lower than 0.05 was considered significant. All statistical analyses were performed using RStudio (Version 1.1.414—2009–2018 RStudio, Inc.).

Role of the funding source

The Hemosepsis study was financially supported by the Programme de Recherche Clinique Inter-régional (academic financial support provided by the French Ministry of Health). The Hemopred study was financially supported by the CIC-P 1435, CHU Limoges. Neither sponsor was involved in any step of the present work.

Results

Among 432 patients from both cohorts, 360 were analyzed (Fig. S3). We excluded from the analysis 50 patients with a history of chronic heart failure, as it was not at all characterized. In most patients (82%), there was no missing data (Fig. S4). Characteristics of the population and mortality are reported in Table 1. Median age was 64 [interquartile 55; 74], SOFA score 10 [7; 12], and SAPS II 57 [45; 70]. No difference was observed between patients initially included in the Hemosepsis or Hemopred cohort regarding severity scores (Table S1). Twenty-one patients (5.8%) were in atrial fibrillation at the time of CCE and 42 patients (11.7%) already received inotropic drug (dobutamine, $n = 17$ or epinephrine, $n = 25$). Day 7 and in-ICU mortality were 20.1% and 35%, respectively.

Analysis in clusters characterized five distinct cardiovascular phenotypes (Tables 1, 2; Fig. 1). Sixty-one patients (16.9%) could be considered as “well resuscitated” (cluster 1), since we observed neither LV systolic dysfunction and RV failure nor fluid responsiveness. Both CI and ScvO₂ were within the normal range. Sixty-four patients (17.7%) had an “LV systolic dysfunction” phenotype (cluster 2). These patients exhibited low LVEF, LVFAC, and CI (29% [22; 40], 26% [18; 33], and 2.2 L/min/m² [1.7; 2.5], respectively), had higher lactate level, required a higher dose of norepinephrine, and were not fluid responders. Only nine of these patients had a LVEF higher than 45%. ScvO₂ remained within the normal range. Similarly, LV filling pressure as reflected by *E/E'* ratio remained non-elevated despite cardiac failure. Eighty-four patients (23.3%) had a phenotype reflecting a

Table 1 Baseline characteristics of the patients included in the analysis according to cluster partition

	All patients	Cluster					p value
		1	2	3	4	5	
	N = 360	n = 61	n = 64	n = 84	n = 81	n = 70	
Demographics							
Age, years	64 [55; 74]	59.0 [50.0; 68.0]	64.0 [54.0; 75.0]	66.0 [59.0; 75.0]	63.0 [55.0; 73.0]	64.0 [55.0; 76.0]	0.011
Male gender	233 (64.7)	35 (57.4)	40 (62.5)	62 (73.8)	54 (66.7)	42 (60.0)	0.245
Chronic respira- tory failure	52 (14.4)	5 (1.4)	12 (3.3)	12 (3.3)	12 (3.3)	11 (3.1)	0.561
Atrial fibrillation at time of CCE	21 (5.8)	4 (1.1)	6 (1.7)	4 (1.1)	3 (0.8)	4 (1.1)	0.670**
SAPS II	57 [45; 70]	50.0 [39.0; 63.0]	62.0 [51.0; 74.0]	55.0 [42.0; 67.0]	59.5 [48.0; 72.0]	56.5 [44.0; 70.0]	0.004
SOFA score	10 [7, 12]	10.0 [8.0; 12.0]	10.0 [8.0; 12.5]	10.0 [7.0; 11.0]	11.0 [7.5; 13.0]	9.0 [7.0; 11.0]	0.089
Arterial blood lactate level, mmol/L	2.5 [1.5; 4.3]	2.7 [1.6; 4.1]	3.1 [2.1; 6.6]	1.9 [1.3; 3.0]	2.6 [1.6; 4.0]	2.7 [1.6; 4.7]	< 0.001
Non-hemodynamic parameters							
PaCO ₂ , mmHg	40 [34; 47]	42.0 [35.9; 49.0]	40.0 [33.5; 44.5]	40.3 [35.0; 48.0]	40.0 [33.5; 47.3]	38.5 [34.0; 43.0]	0.376
PaO ₂ /FiO ₂ , mmHg	184 [113; 262]	170.5 [111.9; 258.8]	174.0 [92.9; 241.8]	195.5 [120.3; 269.4]	153.8 [105.5; 231.5]	204.0 [125.0; 296.0]	0.099
PaO ₂ /FiO ₂ ratio Berlin classifica- tion*							0.628
> 300 mmHg, n (%)	57 (15.9)	12 (20.0)	8 (12.5)	13 (15.5)	9 (11.2)	15 (21.4)	
200–300 mmHg, n (%)	100 (27.9)	14 (23.3)	20 (31.2)	26 (31.0)	20 (25.0)	20 (28.6)	
< 200 mmHg, n (%)	201 (56.1)	34 (56.7)	36 (56.2)	45 (53.6)	51 (63.8)	35 (50.0)	
Site of infection							0.001
Lung, n (%)	169 (46.9)	22 (36.1)	23 (35.9)	43 (51.2)	48 (59.3)	33 (47.1)	
Urinary tract, n (%)	25 (6.9)	3 (4.9)	7 (10.9)	7 (8.3)	4 (4.9)	4 (5.7)	
GI tract, n (%)	109 (30.3)	20 (32.8)	26 (40.6)	21 (25.0)	13 (16.0)	29 (41.4)	
Skin, n (%)	24 (6.7)	8 (13.1)	4 (6.2)	7 (8.3)	2 (2.5)	3 (4.3)	
Others, n (%)	33 (9.2)	8 (13.1)	4 (6.2)	6 (7.1)	14 (17.3)	1 (1.4)	
Outcome							
Day 7 mortality, n (%)	72 (20.1)	6 (9.8)	21 (32.8)	7 (8.3)	22 (27.2)	16 (23.2)	< 0.001
ICU mortality, n (%)	126 (35.0)	13 (21.3)	32 (50.0)	20 (23.8)	34 (42.0)	27 (38.6)	0.001

*Two missing FiO₂

**Exact Fisher's test

“hyperkinetic” state (cluster 3). In these patients, LV systolic function (LVEF 60% [52.5; 66]) and CI (3.3 L/min/m² [2.3; 4.3]) were increased compared to other clusters, and patients exhibited no sign of fluid responsiveness. Eighty-one patients (22.5%) had a hemodynamic profile consistent with underlying “RV failure” (cluster 4). These patients exhibited a markedly high RV/LV EDA ratio (0.8 [0.6; 0.9]) with a normal or supranormal LV systolic function (LVEF 57% [46; 64]) and no more fluid responsiveness. In this subset of patients, more patients had a PaO₂/

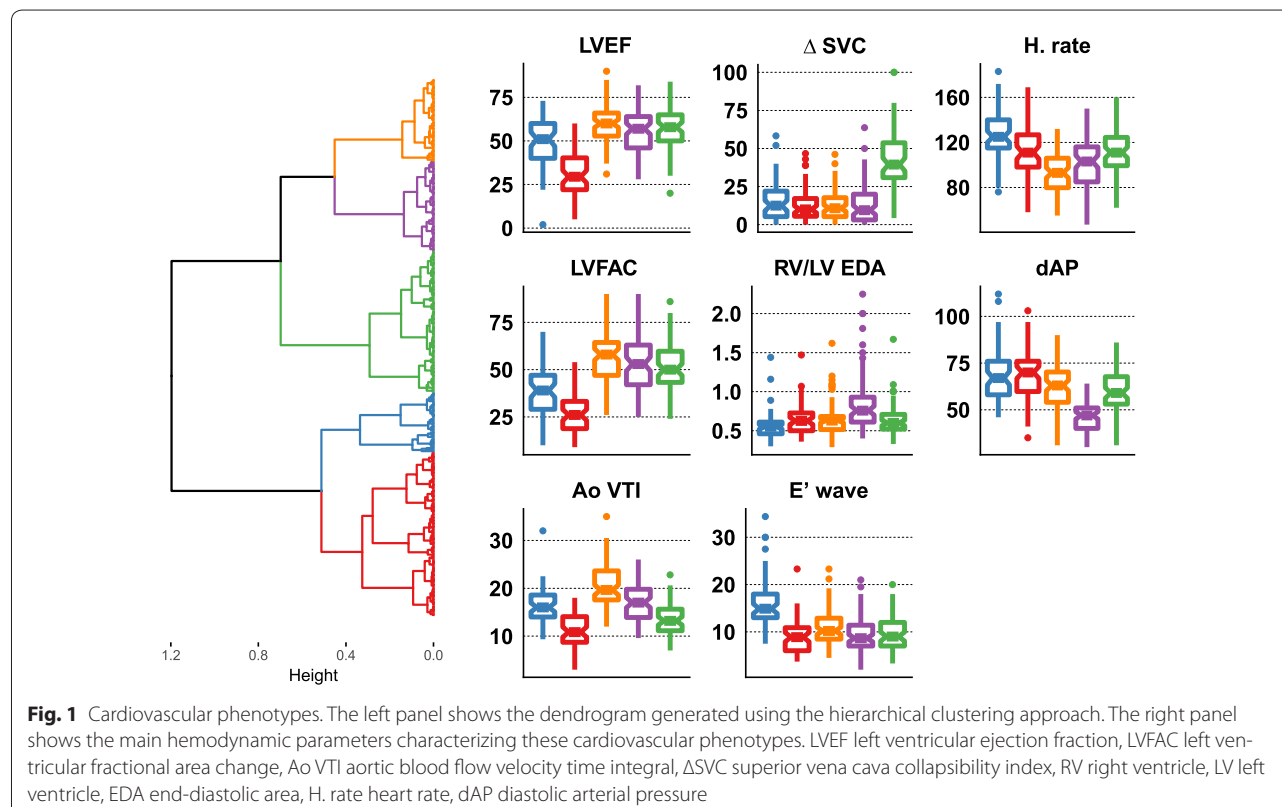
FiO₂ lower than 200 mmHg. Finally, 70 patients (19.4%) belonged to the last cardiovascular phenotype which could be named “still hypovolemic” (cluster 5). These patients exhibited a low CI (2.6 L/min/m² [1.9; 3.1]) despite an increased LV systolic function (LVEF 58% [50; 65]) due to sustained fluid responsiveness, as reflected by markedly elevated ΔSVC (39% [31; 54]), and low preload (CVP 8 mmHg [5; 12]). Interestingly, patients in this cluster received significantly more fluids before CCE than the others, 2762 mL [2500; 4000] and 2000 mL [1000; 3433],

Table 2 Hemodynamic data of the 360 patients included in the study according to the cluster partition

	All patients	Cluster					p value
		1	2	3	4	5	
	N = 360	n = 61	n = 64	n = 84	n = 81	n = 70	
Hemodynamic parameters							
Heart rate, per minute	107 [90; 124]	125.0 [115.0; 140.0]	111.0 [98.0; 127.5]	93.0 [79.5; 106.0]	103.0 [85.0; 116.0]	111.0 [99.0; 125.0]	< 0.001
Systolic arterial blood pressure, mmHg	112 [96; 129]	129.0 [115.0; 141.0]	111.5 [101.5; 124.0]	127.0 [114.0; 139.0]	90.0 [80.0; 100.0]	105.0 [95.0; 120.0]	< 0.001
Diastolic arterial blood pressure, mmHg	59 [51; 70]	67.0 [58.0; 76.0]	70.0 [59.5; 76.0]	63.0 [54.0; 70.5]	47.0 [40.0; 51.0]	59.0 [53.0; 68.0]	< 0.001
Mean arterial blood pressure, mmHg	77 [67; 88]	84.0 [77.0; 97.0]	81.5 [72.5; 92.0]	84.5 [75.0; 91.0]	61.0 [52.0; 67.0]	74.0 [68.0; 82.0]	< 0.001
Cardiac index, L/min/m ²	2.9 [2.1; 3.8]	3.8 [3.0; 4.4]	2.2 [1.7; 2.5]	3.3 [2.3; 4.3]	3.2 [2.4; 3.9]	2.6 [1.9; 3.1]	< 0.001
ScvO ₂ , %	79 [71; 85]	80.0 [72.1; 86.2]	78.0 [66.0; 84.1]	82.3 [75.0; 85.0]	77.7 [69.1; 84.0]	77.0 [69.4; 84.0]	0.038
Central venous pressure, mmHg	10 [7, 13]	11.0 [8.0; 13.5]	10.0 [9.0; 14.0]	10.0 [7.5; 12.0]	9.0 [6.0; 13.0]	8.5 [5.5; 12.0]	0.041
Fluid expansion before CCE, mL	2000 [1000; 3500]	2000 [1228; 3433]	2000 [1000; 3500]	2000 [1500; 3000]	2000 [1000; 3289]	2762 [2500; 4000]	0.175
Hemodynamic treatments							
Epinephrine infusion, n (%)	25 (6.9)	7 (11.5)	11 (17.2)	6 (7.1)	0 (0.0)	1 (1.4)	< 0.001
Epinephrine infusion rate, mg/h	0.0 [0.0; 0.0]	0.0 [0.0; 1.0]	0.0 [0.0; 0.0]	0.0 [0.0; 0.0]	0.0 [0.0; 0.0]	0.0 [0.0; 0.0]	< 0.001
Norepinephrine infusion, n (%)	309 (85.8)	58 (95.1)	56 (87.5)	64 (76.2)	70 (86.4)	61 (87.1)	0.027
Norepinephrine infusion rate, mg/h	1.9 [0.6; 4.0]	2.2 [1.0; 4.0]	3.0 [1.5; 5.5]	1.0 [0.2; 2.3]	2.1 [0.7; 5.0]	1.8 [0.8; 3.5]	< 0.001
Dobutamine infusion, n (%)	17 (4.7)	1 (0.3)	6 (1.7)	2 (0.6)	5 (1.4)	3 (0.8)	0.246*
Dobutamine infusion rate, µg/kg/min	5 [5; 7.5]	5**	5 [5; 6.9]	6 [5.5; 6.5]	8 [5; 10]	5 [3.7; 5.5]	0.551
Echocardiographic parameters							
LVEF, %	54 [40; 64]	51.0 [40.0; 60.0]	29.5 [22.0; 40.5]	60.0 [52.5; 66.0]	57.0 [46.0; 64.0]	57.9 [50.0; 65.0]	< 0.001
LVFAC, %	46 [33; 58]	39.0 [29.0; 47.0]	26.0 [18.5; 33.4]	58.0 [46.9; 64.6]	53.0 [42.0; 63.0]	50.0 [43.0; 60.0]	< 0.001
Mitral E wave, cm/s	68 [54; 87]	90.0 [78.0; 105.0]	56.5 [44.0; 64.5]	77.0 [64.0; 89.5]	68.0 [56.0; 88.0]	51.0 [44.5; 67.0]	< 0.001
Mitral E' wave, cm/s	10 [7.5; 13.6]	14.9 [13.0; 18.0]	8.9 [6.0; 10.9]	10.2 [8.3; 12.9]	8.7 [7.0; 11.4]	9.0 [7.0; 12.0]	< 0.001
E/E' ratio	6.8 [5.3; 9.3]	6.0 [4.6; 7.4]	6.6 [5.3; 9.4]	7.3 [5.5; 9.3]	8.4 [5.2; 11.1]	6.3 [4.3; 8.8]	0.003
Aortic VTI, cm	15.4 [12.8, 19]	16.0 [14.0; 18.6]	10.9 [8.7; 14.2]	19.6 [17.6; 23.8]	17.0 [13.9; 19.8]	13.2 [11.1; 15.7]	< 0.001
RV/LV EDA	0.6 [0.5; 0.8]	0.6 [0.5; 0.6]	0.6 [0.5; 0.7]	0.6 [0.5; 0.7]	0.8 [0.6; 0.9]	0.6 [0.5; 0.7]	< 0.001
ΔSVC, %	13.2 [6; 29.2]	12.5 [5.3; 22.0]	10.0 [5.4; 17.3]	10.8 [5.3; 17.8]	9.5 [3.1; 20.0]	39.4 [30.8; 54.0]	< 0.001

*Exact Fisher's test

**Only one patient received dobutamine



respectively, $p < 0.001$. ICU mortality was 21.3 [95% CI 13.0; 33.1], 50.0 [38.1; 61.9], 23.8 [16.0; 33.9], 42 [31.8; 52.8], and 38.6 [28.0; 50.3] % in clusters 1, 2, 3, 4, and 5, respectively ($p < 0.001$). Figure 2 and Table 3 report the distribution of parameters which characterize clusters as well as the respective importance of each of them in the partition process. Figure 3 reports the overall good performance of the three most important variables in each cluster, with a very high specificity but a quite low sensitivity.

Discussion

The clustering approach combining echocardiographic parameters (LVEF, LVFAC, aortic VTI, RV/LV EDA, Δ SVC, mitral E wave velocity, and E' wave velocity) and clinical parameters (heart rate, blood pressure, type and dose of catecholamine) allowed us to characterize five distinct cardiovascular phenotypes, the hemodynamic profiles of which correspond to “well-resuscitated” patients (16.9%, cluster 1), patients with LV systolic dysfunction (17.7%, cluster 2), hyperkinetic profile (23.3%, cluster 3), RV failure (22.5%, cluster 4), and sustained hypovolemia (19.4%, cluster 5).

This approach in clustering without any a priori criteria was able to distinguish different phenotypes between all the expected alterations of the macrocirculation. In the

LV systolic dysfunction phenotype (cluster 2), patients had a higher serum lactate level, a lower CI, and a higher dose of norepinephrine. While differences in lactate and CI probably reflect the severity of underlying septic cardiomyopathy, the higher dose of norepinephrine could have participated in the development of LV failure. It has been reported that in animal models alteration in LV intrinsic contraction is constant in sepsis [22] and that the level of LV afterload especially alters LVEF in this abnormal heart [23]. Accordingly, the increased dose of norepinephrine administered in this subset of patients could have unmasked, or even participated in, the observed LV systolic dysfunction by increasing afterload. Jardin et al. and Parker et al. reported higher systemic vascular resistance in patients with LV systolic dysfunction [2, 24]. Boissier et al. [25] recently reported a negative correlation between echocardiographic parameters of LV systolic function and those of LV afterload. Patients with this cardiovascular phenotype also failed to exhibit increased E/E' , which is widely considered as a surrogate of LV filling pressure [26]. The absence of elevation in LV filling pressure has been reported as a specific characteristic of this hemodynamic profile, not only when evaluated by the E/E' but also when measured in the past using a pulmonary artery catheter [2, 24]. It was suggested to be related to an increase in LV compliance

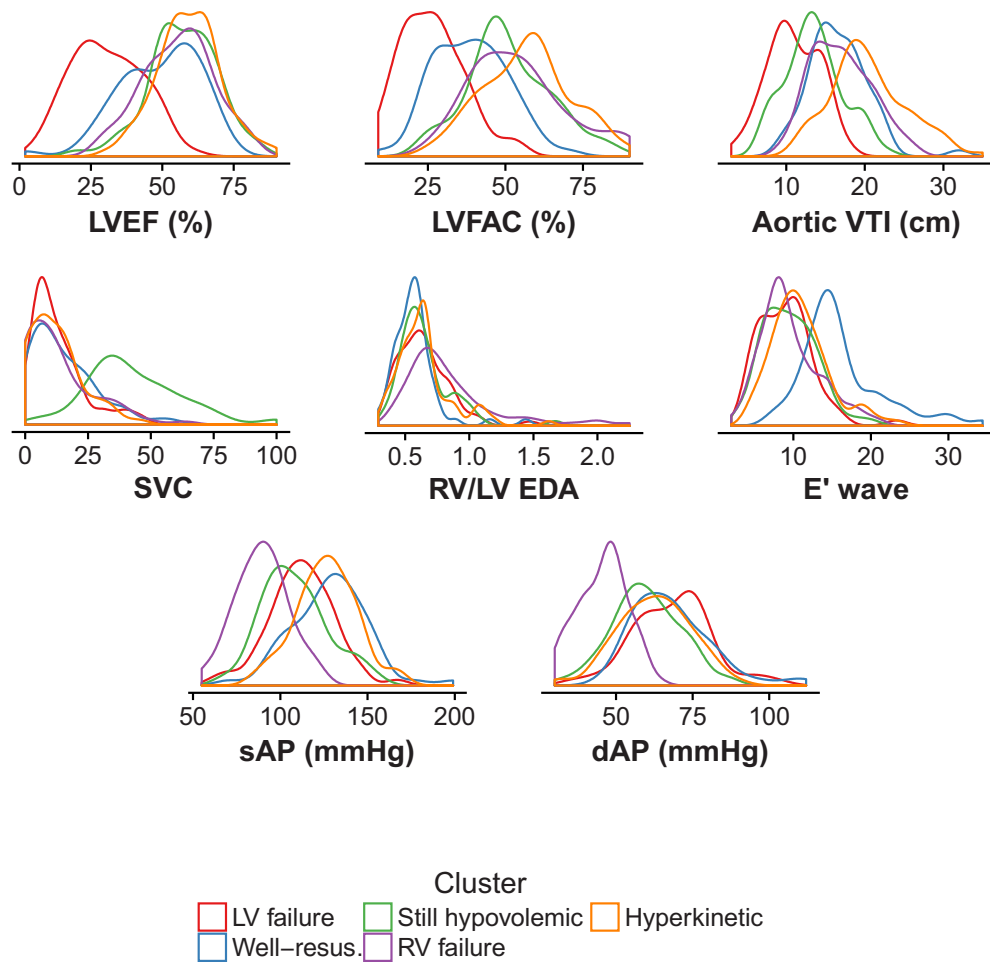


Fig. 2 Distribution of the factors contributing most in the clustering approach in the five cardiovascular phenotypes. LVEF left ventricular ejection fraction, LVFAC left ventricular fractional area change, Ao VTI aortic blood flow velocity time integral, Δ SVC superior vena cava collapsibility, RV right ventricle, LV left ventricle, EDA end-diastolic area, dAP diastolic arterial pressure, sAP systolic arterial pressure

Table 3 Variables that significantly contributed to the cluster

Variable	Cluster 1	Cluster 2	Cluster 3	Cluster 4	Cluster 5
LVEF (%)	–	31/51 (4×10^{-30})	59/51 (1×10^{-7})	56/51 (2×10^{-3})	57/51 (2×10^{-4})
LVFAC (%)	39/47 (10×10^{-5})	26/47 (9×10^{-25})	57/47 (4×10^{-10})	54/47 (2×10^{-5})	51/47 (1×10^{-2})
Aortic VTI (cm)	–	11/16 (4×10^{-18})	20/16 (5×10^{-20})	–	14/16 (8×10^{-6})
Mitral E wave (cm/s)	92/71 (2×10^{-14})	58/71 (1×10^{-6})	78/71 (2×10^{-3})	–	55/71 (3×10^{-10})
E' wave (cm/s)	16/11 (5×10^{-22})	9/11 (4×10^{-4})	–	10/11 (4×10^{-3})	10/11 (7×10^{-3})
RV/LV EDA	0.57/0.68 (8×10^{-4})	–	–	0.85/0.68 (2×10^{-10})	–
Δ SVC (%)	–	13/19 (2×10^{-3})	12/19 (8×10^{-5})	13/19 (2×10^{-3})	43/19 (2×10^{-35})
sBP (mmHg)	128/112 (4×10^{-9})	–	126/112 (3×10^{-10})	89/112 (1×10^{-23})	–
dBp (mmHg)	68/60 (9×10^{-7})	68/60 (5×10^{-7})	–	45/60 (1×10^{-26})	–
Heart rate (/min)	127/108 (2×10^{-11})	–	93/108 (2×10^{-10})	101/108 (9×10^{-3})	–
Norepinephrine infusion rate (mg/h)	–	3.8/2.6 (7×10^{-5})	1.6/2.6 (7×10^{-5})	3.1/2.6 (3×10^{-2})	–
Epinephrine infusion rate (mg/h)	–	0.3/0.1 (1×10^{-2})	–	0/0.1 (2×10^{-2})	–

Bold numbers show the three most important variables for each cluster. Values shown in each cell are mean in cluster/overall mean (p value)

LVEF left ventricular ejection fraction, LVFAC left ventricular fractional area change, Ao VTI aortic blood flow velocity time integral, Δ SVC superior vena cava collapsibility, RV right ventricle, LV left ventricle, EDA end-diastolic area, dAP diastolic arterial pressure, sAP systolic arterial pressure

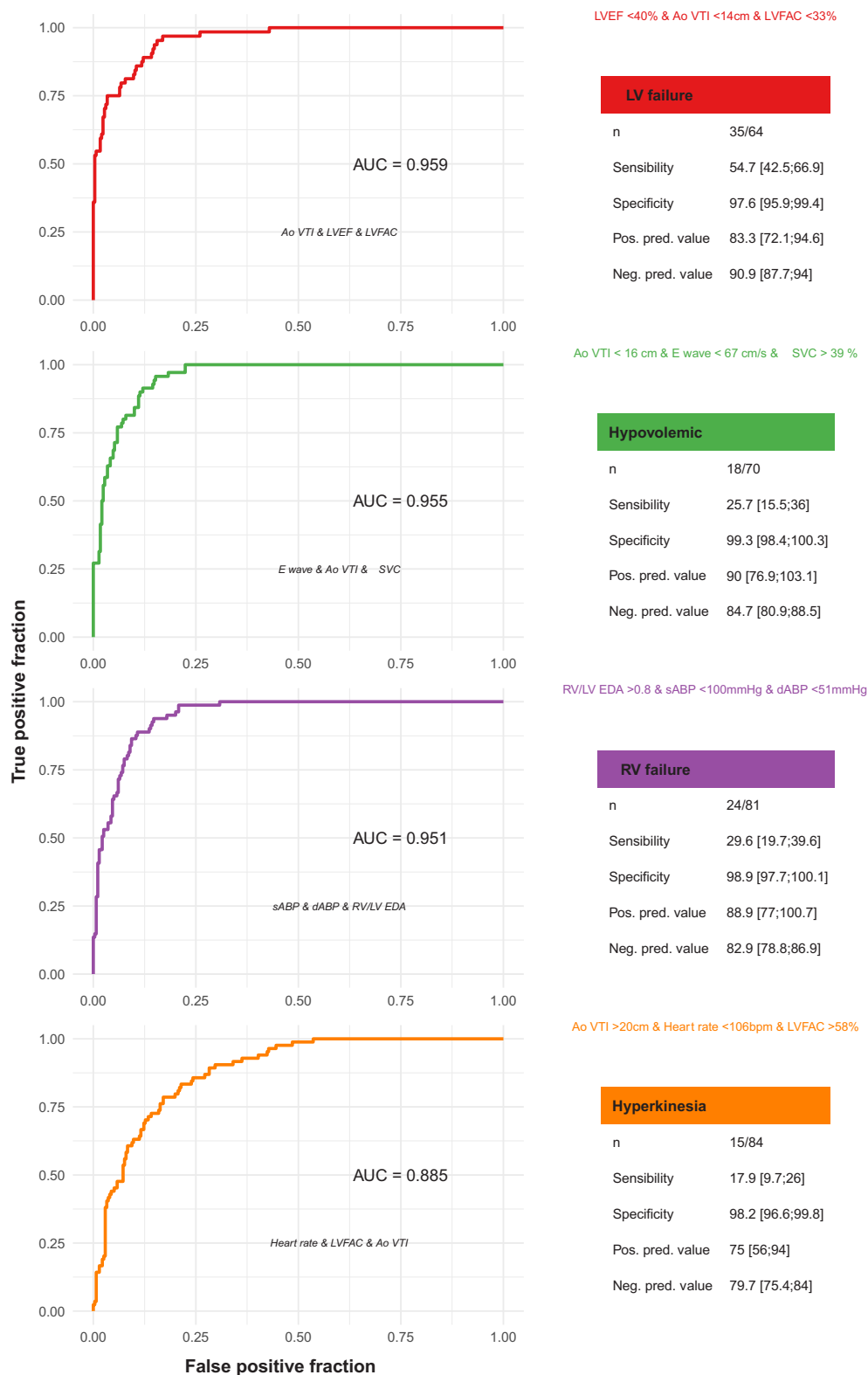


Fig. 3 Performance of the combination of the three most important variables in the clustering approach for each cluster. The ROC curves on the left of the figure are drawn from a multivariable logistic regression with being in a cluster (binary variable yes/no) as the dependent variable and the three most important variables as independent variables. The areas under the curve and the three variables are listed on the plot. On the right of the figure are presented the diagnostic performance of the combination of the three variables for each cluster

due to sepsis [27]. Our clustering approach was also able to differentiate the “still hypovolemic” cardiovascular phenotype (cluster 5) from the “hyperkinetic” (cluster 3), both conditions being characterized by the presence of global LV hyperkinesia. Nevertheless, CI, aortic VTI (a surrogate of LV stroke volume), and CVP were lower in patients with persistent hypovolemia when compared to their counterparts, and Δ SVC was consistent with fluid responsiveness, as opposed to patients with a hyperkinetic hemodynamic profile. Patients who had a cardiovascular phenotype consistent with RV failure (cluster 4) were the only ones to exhibit a large increase in RV/LV EDA, which was recently considered to define RV failure [16]. It is noteworthy that these patients had a higher proportion of $\text{PaO}_2/\text{FiO}_2$ level lower than 200 mmHg, suggesting that RV failure was potentially secondary to the development of ARDS-related septic shock. Finally, we failed to identify a cardiovascular phenotype corresponding to patients with isolated LV diastolic dysfunction, as previously reported [15]. In contrast, as indicated by the median E' maximal velocity, LV diastolic dysfunction was uniformly distributed in all phenotypes, with the exception of the well resuscitated patients (cluster 1).

Our approach may have potential interest for optimizing hemodynamic support using CCE after the very early resuscitation phase since personalized medicine has gained more and more importance [7]. A recent pilot study suggested that early vasopressor infusion could restrict fluid volume without detrimental effect on prognosis [9]. If our approach is able to better characterize patients who really need more fluids after the early phase, this could significantly change management and prognosis. While fluid overload is suggested to be detrimental [28], inappropriate use of vasopressors in still hypovolemic situations may lead to increase in tissue hypoperfusion and severe ischemia of vital organs [29]. The need for inotropic infusion in septic shock is still an issue of controversy. Indeed no study ever reported a relationship between LV systolic dysfunction and mortality and a beneficial effect of inotrope infusion. In a recent single-center randomized controlled trial, infusion of levosimendan did not modify prognosis but has deleterious cardiovascular effects [14]. However, the authors did not select their population at all on the basis of a pre-existing LV systolic dysfunction phenotype [14]. Our clustering approach, defining LV systolic dysfunction without a binary and simplistic threshold value of LVEF but by combining the usual echocardiographic and clinical hemodynamic variables as done in daily practice, could allow better characterization of these patients. How these patients could benefit from dobutamine infusion should be evaluated in the future. Finally, it is now well

recognized that the right ventricle may fail in septic shock, especially when associated with ARDS, and that it may induce low flow state [30–32]. It was suggested that this profile may induce false positive pulse pressure variations [33, 34], a parameter frequently used to manage fluids. Accurately diagnosing RV failure could also help physicians to adequately optimize hemodynamic and respiratory support [30].

Our study suffers from limitations. First, even if one patient could only be statistically classified into one cardiovascular phenotype, interquartiles show a substantial between-cluster overlap for most studied parameters. This was expected since clusters were more defined by a specific association of parameters than by the presence of a single specific one. A soft clustering approach that addresses the overlapping issues might be of interest in future research [35]. Moreover, application of clustering in clinical practice is therefore not so obvious, even though we report in Fig. 3 and Table 3 the most valuable parameters that counted in the determination of each cluster and their value distribution. Last, the explained variance in the principal component analysis was quite low, reflecting the potential unmeasured factors we did not have to better describe these phenotypes. Second, we were unable to provide the exact timing of CCE following admission, which could alter our results, especially for the cluster 5 “still hypovolemic”. However, we reported that these patients did not receive less fluid than the others before CCE performance, and even more, which may suggest a particular profile of capillary linkage. Third, we did not repeat the hemodynamic evaluation by CCE, as we only evaluated our cohort in the first 12 h after initial resuscitation of septic shock patients. It is of interest to assess in future studies whether the transition from one cardiovascular phenotype to another during the first 2–3 days could alter outcome, and whether it is related to spontaneous progression or induced by therapy. Fourth, variabilities of echo parameters were not calculated in the present cohort. However, the intraobserver and interobserver reproducibility of SVC collapsibility index were calculated as good to excellent in the Hemopred study [13]. The same authors, using the same route (TEE), in the same population (septic shock) previously reported interobserver variability of AoVTI, RV/LV EDA, LV volumes, and areas [10] which were below 10–11%. Fifth, we decided not to include in the analysis the 50 patients with a history of chronic heart failure. Indeed, we did not have any characterization of this failure; in particular, we did not know whether it was systolic, diastolic, valvular, injuring the right ventricle, and even the treatment. Finally, all CCE evaluations were performed by highly trained intensivists, which limits the external validity of our results, even though patients came from 12 different

ICUs and we reported in the past that the learning curve of TEE hemodynamic evaluation was steep and skills quickly achieved [36].

In conclusion, using a clustering approach in a large cohort of patients with septic shock evaluated early by CCE, we identified five distinct cardiovascular phenotypes which could help physicians to individualize the hemodynamic support. How this better characterization could change management and prognosis should be evaluated in the future.

Electronic supplementary material

The online version of this article (<https://doi.org/10.1007/s00134-019-05596-z>) contains supplementary material, which is available to authorized users.

Author details

¹ Medical-Surgical Intensive Care Unit, Ambroise Paré University Hospital, APHP, 9 avenue Charles de Gaulle, 92100 Boulogne-Billancourt, France. ² UFR des Sciences de la Santé Simone Veil, Université Versailles Saint Quentin, Versailles, France. ³ INSERM UMR1018, Team Kidney and Heart, CESP, Villejuif, France. ⁴ Medical-Surgical Intensive Care Unit, Limoges University Hospital, Limoges, France. ⁵ Faculty of Medicine, University of Limoges, Limoges, France. ⁶ INSERM CIC 1435, Limoges University Hospital, Limoges, France. ⁷ Medical-Surgical Intensive Care Unit, Teaching Hospital of Toulouse, Toulouse, France.

Acknowledgements

The authors thank Mr. David Marsh for his English editing and Dr. Ana Catalina Hernandez Padilla for her help in the data management.

Author's contribution

GG, PV, AA and AVB designed the study and drafted the manuscript. PV, AA, ALF, CC, SS, XR and AVB collected the data. GG conducted the statistical analysis. AA, ALF, CC, SS and XR carefully revised the manuscript.

Funding

The Hemosepsis study was financially supported by the Programme de Recherche Clinique Inter-régional (academic financial support provided by the French Ministry of Health). The Hemopred study was financially supported by the CIC-P 1435, CHU Limoges.

Compliance with ethical standards

Conflicts of interest

GG, PV, AU, ALF, CC, SS, XR declared no conflict of interest. AVB has received grant from GSK for conducting clinical research and is a member of the scientific advisory board.

Ethical approval

Both cohorts (Hemosepsis and Hemopred) received Limoges ethics committee approval.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Received: 13 November 2018 Accepted: 6 March 2019

Published online: 19 March 2019

References


- Hess ML, Hastillo A, Greenfield LJ (1981) Spectrum of cardiovascular function during gram-negative sepsis. *Prog Cardiovasc Dis* 23:279–298

- Parker MM, Shelhamer JH, Bacharach SL et al (1984) Profound but reversible myocardial depression in patients with septic shock. *Ann Intern Med* 100:483–490
- De Backer D, Bakker J, Cecconi M et al (2018) Alternatives to the Swan–Ganz catheter. *Intensive Care Med* 44:730–741. <https://doi.org/10.1007/s00134-018-5187-8>
- Viellard-Baron A, Schmitt JM, Beauchet A et al (2001) Early preload adaptation in septic shock? A transesophageal echocardiographic study. *Anesthesiology* 94:400–406
- Bégot E, Dalmay F, Etchecopar C et al (2015) Hemodynamic assessment of ventilated ICU patients with cardiorespiratory failure using a miniaturized multiplane transesophageal echocardiography probe. *Intensive Care Med* 41:1886–1894. <https://doi.org/10.1007/s00134-015-3998-4>
- Viellard-Baron A, Caille V, Charron C et al (2008) Actual incidence of global left ventricular hypokinesia in adult septic shock. *Crit Care Med* 36:1701–1706. <https://doi.org/10.1097/CCM.0b013e318174db05>
- Coopersmith CM, De Backer D, Deutschman CS et al (2018) Surviving Sepsis Campaign: research priorities for sepsis and septic shock. *Intensive Care Med* 44:1400–1426. <https://doi.org/10.1007/s00134-018-5175-z>
- Perner A, Cecconi M, Cronhjort M et al (2018) Expert statement for the management of hypovolemia in sepsis. *Intensive Care Med* 44:791–798. <https://doi.org/10.1007/s00134-018-5177-x>
- Macdonald SPJ, Keijzers G, Taylor DM et al (2018) Restricted fluid resuscitation in suspected sepsis associated hypotension (REFRESH): a pilot randomised controlled trial. *Intensive Care Med* 44:2070–2078. <https://doi.org/10.1007/s00134-018-5433-0>
- Viellard-Baron A, Prin S, Chergui K et al (2003) Hemodynamic instability in sepsis: bedside assessment by Doppler echocardiography. *Am J Respir Crit Care Med* 168:1270–1276. <https://doi.org/10.1164/rccm.200306-816CC>
- Åneman A, Viellard-Baron A (2016) Cardiac dysfunction in sepsis. *Intensive Care Med* 42:2073–2076. <https://doi.org/10.1007/s00134-016-4503-4>
- Vignon P, Bégot E, Mari A et al (2018) Hemodynamic assessment of patients with septic shock using transpulmonary thermodilution and critical care echocardiography: a comparative study. *Chest* 153:55–64. <https://doi.org/10.1016/j.chest.2017.08.022>
- Vignon P, Repesse X, Bégot E et al (2017) Comparison of echocardiographic indices used to predict fluid responsiveness in ventilated patients. *Am J Respir Crit Care Med* 195:1022–1032. <https://doi.org/10.1164/rccm.201604-0844OC>
- Huang SJ, Nalos M, McLean AS (2013) Is early ventricular dysfunction or dilatation associated with lower mortality rate in adult severe sepsis and septic shock? A meta-analysis. *Crit Care* 17:R96. <https://doi.org/10.1186/cc12741>
- Landesberg G, Gilon D, Merozy Y et al (2012) Diastolic dysfunction and mortality in severe sepsis and septic shock. *Eur Heart J* 33:895–903. <https://doi.org/10.1093/eurheartj/ehs351>
- Viellard-Baron A, Naeije R, Haddad F et al (2018) Diagnostic workup, etiologies and management of acute right ventricle failure. *Intensive Care Med* 44:774–790. <https://doi.org/10.1007/s00134-018-5172-2>
- Viellard-Baron A, Chergui K, Rabiller A et al (2004) Superior vena caval collapsibility as a gauge of volume status in ventilated septic patients. *Intensive Care Med* 30:1–6. <https://doi.org/10.1007/s00134-004-2361-y>
- Wetterslev M, Møller-Sørensen H, Johansen RR, Perner A (2016) Systematic review of cardiac output measurements by echocardiography vs. thermodilution: the techniques are not interchangeable. *Intensive Care Med* 42:1223–1233. <https://doi.org/10.1007/s00134-016-4258-y>
- Lê S, Josse J, Husson F (2008) FactoMineR: an R package for multivariate analysis. *J Stat Soft* 25:1–18
- Husson F, Josse J, Pagès J (2010) Principal component methods—hierarchical clustering—partitional clustering: why would be need to choose for visualizing data? Technical report, pp 1–17
- Josse J, Pagès J, Husson F (2011) Multiple imputation in principal component analysis. *Adv Data Anal Classif* 5:231–246. <https://doi.org/10.1007/s11634-011-0086-7>
- Barraud D, Faivre V, Damy T et al (2007) Levosimendan restores both systolic and diastolic cardiac performance in lipopolysaccharide-treated rabbits: comparison with dobutamine and milrinone. *Crit Care Med* 35:1376–1382. <https://doi.org/10.1097/01.CCM.0000261889.18102.84>

23. Robotham JL, Takata M, Berman M, Harasawa Y (1991) Ejection fraction revisited. *Anesthesiology* 74:172–183. <https://doi.org/10.1097/00000542-199101000-00026>
24. Jardin F, Brun-Ney D, Auvert B et al (1990) Sepsis-related cardiogenic shock. *Crit Care Med* 18:1055–1060. <https://doi.org/10.1097/00003246-199010000-00001>
25. Boissier F, Razazi K, Seemann A et al (2017) Left ventricular systolic dysfunction during septic shock: the role of loading conditions. *Intensive Care Med* 43:633–642. <https://doi.org/10.1007/s00134-017-4698-z>
26. Vignon P, Allot V, Lesage J et al (2007) Diagnosis of left ventricular diastolic dysfunction in the setting of acute changes in loading conditions. *Crit Care* 11:R43. <https://doi.org/10.1186/cc5736>
27. Suffredini AF, Fromm RE, Parker MM et al (1989) The cardiovascular response of normal humans to the administration of endotoxin. *N Engl J Med* 321:280–287. <https://doi.org/10.1056/NEJM198908033210503>
28. Perner A, Vieillard-Baron A, Bakker J (2015) Fluid resuscitation in ICU patients: Quo vadis? *Intensive Care Med* 41:1667–1669. <https://doi.org/10.1007/s00134-015-3900-4>
29. Murakawa K, Kobayashi A (1988) Effects of vasopressors on renal tissue gas tensions during hemorrhagic shock in dogs. *Crit Care Med* 16:789–792
30. Vieillard-Baron A, Naeije R, Haddad F et al (2018) Diagnostic workup, etiologies and management of acute right ventricle failure. *Intensive Care Med* 44:774–790. <https://doi.org/10.1007/s00134-018-5172-2>
31. Harjola V-P, Mebazaa A, Čelutkienė J et al (2016) Contemporary management of acute right ventricular failure: a statement from the Heart Failure Association and the Working Group on Pulmonary Circulation and Right Ventricular Function of the European Society of Cardiology. *Eur J Heart Fail* 18:226–241. <https://doi.org/10.1002/ehf.478>
32. Lahm T, Douglas IS, Archer SL et al (2018) Assessment of right ventricular function in the research setting: knowledge gaps and pathways forward. An official American Thoracic Society Research Statement. *Am J Respir Crit Care Med* 198:e15–e43. <https://doi.org/10.1164/rccm.201806-1160ST>
33. Vieillard-Baron A, Matthay M, Teboul JL et al (2016) Experts' opinion on management of hemodynamics in ARDS patients: focus on the effects of mechanical ventilation. *Intensive Care Med* 42:739–749. <https://doi.org/10.1007/s00134-016-4326-3>
34. Mahjoub Y, Pila C, Friggeri A et al (2009) Assessing fluid responsiveness in critically ill patients: false-positive pulse pressure variation is detected by Doppler echocardiographic evaluation of the right ventricle. *Crit Care Med* 37:2570–2575. <https://doi.org/10.1097/CCM.0b013e3181a380a3>
35. Peters G, Crespo F, Lingras P, Weber R (2013) Soft clustering—fuzzy and rough approaches and their extensions and derivatives. *Int J Approx Reason* 54:307–322. <https://doi.org/10.1016/j.ijar.2012.10.003>
36. Charron C, Vignon P, Prat G et al (2013) Number of supervised studies required to reach competence in advanced critical care transesophageal echocardiography. *Intensive Care Med* 39:1019–1024. <https://doi.org/10.1007/s00134-013-2838-7>

ORIGINAL

Fever control in critically ill adults. An individual patient data meta-analysis of randomised controlled trials

Paul J. Young^{1,2*} , Rinaldo Bellomo³, Gordon R. Bernard⁴, Daniel J. Niven⁵, Frederique Schortgen⁶, Manoj Saxena^{7,9}, Richard Beasley² and Mark Weatherall⁸

© 2019 Springer-Verlag GmbH Germany, part of Springer Nature

Abstract

Purpose: One potential way to protect patients from the physiological demands that are a consequence of fever is to aim to prevent fever and to treat it assiduously when it occurs. Our primary hypothesis was that more active fever management would increase survival among patient subgroups with limited physiological reserves such as older patients, patients with higher illness acuity, and those requiring organ support.

Methods: We conducted an individual-level patient data meta-analysis of randomised controlled trials to compare the outcomes of ICU patients who received more active fever management with the outcomes of patients who received less active fever management. The primary outcome variable of interest was the unadjusted time to death after randomisation.

Results: Of 1413 trial participants, 707 were assigned to more active fever management and 706 were assigned to less active fever management. There was no statistically significant heterogeneity in the effect of more active compared with less active fever management on survival in any of the pre-specified subgroups that were chosen to identify patients with limited physiological reserves. Overall, more active fever management did not result in a statistically significant difference in survival time compared with less active fever management [hazard ratio 0.91; (95% CI 0.75–1.10), $P=0.32$].

Conclusions: Our findings do not support the hypothesis that more active fever management increases survival compared with less active fever management overall or in patients with limited physiological reserves.

Keywords: Sepsis, Fever, Septic shock, Paracetamol, Non-steroidal anti-inflammatory drugs, Physical cooling

Introduction

Fever occurs commonly in intensive care (ICU) patients and increases metabolic demand [1]. Increasing metabolic demand has important physiological consequences on oxygen consumption and cardiac output [1]. One

potential way to protect patients from the physiological demands that are a consequence of fever is to aim to prevent fever and to treat it assiduously when it develops [2]. This strategy is an attractive candidate intervention to improve outcomes in the ICU setting because patients with a range of critical illnesses including major trauma, infection, acute myocardial infarction, and pancreatitis develop fever [3–5], and many such patients have limited physiological reserves.

Body temperature can be manipulated in ICU patients with medicines [6, 7] and physical cooling devices [8] allowing for more or less active approaches to fever

*Correspondence: Paul.Young@ccdhb.org.nz

¹ Intensive Care Unit, Wellington Hospital, Capital and Coast District Health Board, Wellington, New Zealand

Full author information is available at the end of the article

management. In a recent systematic review and aggregate data meta-analysis evaluating the effect of fever management on all-cause mortality in ICU patients, we found that more active fever management neither increased nor decreased mortality in critically ill adults compared with less active fever management [9]. However, despite these findings, it is plausible that the balance of risks and benefits of active fever management in ICU patients varies based on the physiological reserves of the patients being treated and the nature of their illness [10]. In this context, we submit that physiological reserves are reasonably defined as the capacity of a patient to cope with the physiological demands associated with fever and depend on patients' physiology, illness severity, and the organ support they require.

Our primary hypothesis was that more active fever management would increase survival among patient subgroups with limited physiological reserves such as older patients, patients with higher illness acuity, and those with very high body temperature (≥ 39.5 °C). Because fever is part of the adaptive host response to infection [11], we further hypothesised that more active fever management would improve survival in the absence of infection but not in the presence of infection.

Methods

Study design

To address our hypotheses, we conducted an individual-level patient data meta-analysis (IPDMA) using available data from randomised controlled trials identified in our recent systematic review and aggregate data meta-analysis [9]. We contacted lead investigators for all randomised controlled trials identified in the recent systematic review and requested access to individual patient-level data (see ESM for details). The search strategy used in our systematic review has been published previously [9]; however, in brief, we searched major databases for randomised controlled trials evaluating fever management in adult ICU patients excluding trials where the intervention involved therapeutic hypothermia. We included trials that evaluated any treatment administered commonly to febrile patients to reduce body temperature. The protocol for this IPDMA was posted online on 29 June 2018 at <http://wellingtonicu.com/PubResPres/Protocols/> in advance of analyses being undertaken.

Data extraction and cleaning for analysis

Pre-randomisation (baseline) data points were extracted from individual study databases. These were age, gender, invasively ventilated (yes or no), receiving inotropes and/or vasopressors at baseline (yes or no), suspected infection at baseline (yes or no), Acute Physiology And Chronic Health Evaluation (APACHE) II score [12],

Take-home message

We conducted an individual level patient data meta-analysis of randomised controlled trials to compare the outcomes of ICU patients who received more active fever management with the outcomes of patients who received less active fever management. Our findings do not support the hypothesis that more active fever management increases survival compared with less active fever management overall, or in patients with limited physiological reserves.

mean arterial pressure (mmHg), heart rate (beats per min), serum creatinine ($\mu\text{mol/l}$), and body temperature (°C).

We sought to compare the outcomes of patients who received more active fever management with the outcomes of patients who received less active fever management. Accordingly, where studies compared an antipyretic drug with placebo, the patients allocated to the antipyretic drug were considered to have received more active fever management. Where studies compared different thresholds for temperature treatment, patients allocated to the group with the lowest body temperature target were considered to have received more active fever management.

Time to death after randomisation was defined as the difference between time zero (T0) and the date and time of death. T0 was generally defined as the date and time of randomisation. In one study, where the date and time of randomisation were not recorded, the date and time of administration of the first dose of study medication defined T0. Where no time, only a date, was available to define either T0 or the time of death, the time(s) were assumed to be 12:00 p.m. The time for censored participants (those who did not die) was defined as the last time of observation in relation to the time of randomisation as described above. All patients who died on or before the date of ICU discharge were defined as dead for the purposes of evaluating the end point 'mortality at ICU discharge'. ICU and hospital length of stay were defined as the difference between T0 as described above and ICU and hospital discharge, respectively. Where no time, only a date, was available to define either T0 or the time of discharge, the time(s) were assumed to be 12:00 p.m. Body temperature at 6, 12, 24, 48, and 72 h after randomisation were included in the IPDMA database. One study reported temperature data at 4 h and 8 h after randomisation [7]. For this study, the 6-h temperature data point was calculated by averaging the values from the 4–8 h time points.

Outcomes

The primary outcome variable of interest was the time to death after randomisation. This outcome was chosen

Table 1 Baseline characteristics

Characteristic	More active fever control (n = 707)	Less active fever control (n = 706)
Age (years)	56.7 ± 17.2 (n = 705)	56.5 ± 17.2 (n = 705)
Male sex, n/N (%)	415/707 (58.7)	397/706 (56.2)
Intensive care support, n/N (%)		
Invasive mechanical ventilation	498/707 (70.4)	500/706 (70.8)
Receiving inotropes and/or vasopressors	358/707 (50.6)	377/706 (53.4)
APACHE-II score ^a	17.1 ± 7.1 (n = 605)	17.3 ± 7.4 (n = 605)
SAPS score ^b	76.9 ± 13.9 (n = 101)	78.3 ± 14.4 (n = 99)
Biochemistry and physiology		
Heart rate (beats per min)	106 ± 22 (n = 703)	107 ± 23 (n = 705)
Mean arterial pressure (mmHg)	78 ± 15 (n = 704)	77 ± 14 (n = 705)
Temperature (°C)	38.5 ± 1.0 (n = 703)	38.5 ± 0.9 (n = 706)
Serum creatinine (μmol/l)	134 ± 128 (n = 536)	129 ± 123 (n = 535)
Study, n/N (%)		
Bernard et al.	224/707 (31.7)	231/706 (32.7)
Niven et al.	14/707 (2.0)	12/706 (1.7)
Saxena et al.	21/707 (3.0)	20/706 (2.8)
Schortgen et al.	101/707 (14.3)	99/706 (14.0)
Young et al.	347/707 (49.1)	344/706 (48.7)
Suspected infection at baseline	672/707 (95.1)	673/706 (95.3)

Plus/minus values are mean ± SD

APACHE Acute Physiology and Chronic Health Evaluation, ICU intensive care unit, SAPS Simplified Acute Physiology Score

^a Scores on the APACHE II range from 0 to 71, with higher scores indicating more severe disease and a higher risk of death. APACHE II scores were not collected for participants in the Schortgen et al. trial

^b Scores on the SAPS III range from 0 to 263, with higher scores indicating more severe disease and a higher risk of death. SAPS III was only collected for participants in the Schortgen et al. trial

as the primary outcome because it allowed multiple trials with different durations of follow-up to be combined without loss of data. Secondary outcomes were mortality at ICU discharge, ICU and hospital length of stay, and body temperature at 6, 12, 24, 48, and 72 h following randomisation.

Statistical analyses

Data summaries by treatment group are frequency and proportions expressed as percentages for categorical data and mean ± standard deviation for continuous data.

Survival times use log-rank tests and are shown as Kaplan-Meier curves and relative survival estimated with a Cox proportional-hazards model. The primary analysis model was adjusted for study as fixed effect; however, a sensitivity analysis adjusted for baseline covariates of age, sex, and APACHE-II score was also performed incorporating individual study as a fixed effect. Because the APACHE-II score was not available for one study [8] two post hoc sensitivity analyses were used: one in which the APACHE-II score was not included in the adjusted model and another in which illness severity scores based on Simplified Acute Physiology Score (SAPS3) [13] were

used in place of APACHE-II scores for the study that did not include APACHE-II data (see ESM for details).

ICU mortality was compared by treatment group using logistic regression with analyses performed in a similar fashion to those performed for survival. Mortality data by treatment group are reported as frequencies with proportions expressed as percentages with treatment effects reported as odds ratios.

ICU and hospital length of stay were highly skewed and were analysed on the logarithm transformed scale. These variables are summarised as geometric mean with treatment effects expressed as a ratio of geometric means. A post hoc analysis evaluating whether ICU length of stay differed in relation to randomised treatment for patients who did or did not die in the ICU used an ANCOVA-based interaction model.

Interaction analyses were also used to explore whether survival, ICU mortality, ICU length of stay, or hospital length of stay varied in pre-specified subgroups based on pre-randomisation characteristics. The subgroups of interest were: invasively ventilated or not; receiving inotropes and/or vasopressors or not; both invasively ventilated and receiving inotropes and/or vasopressors

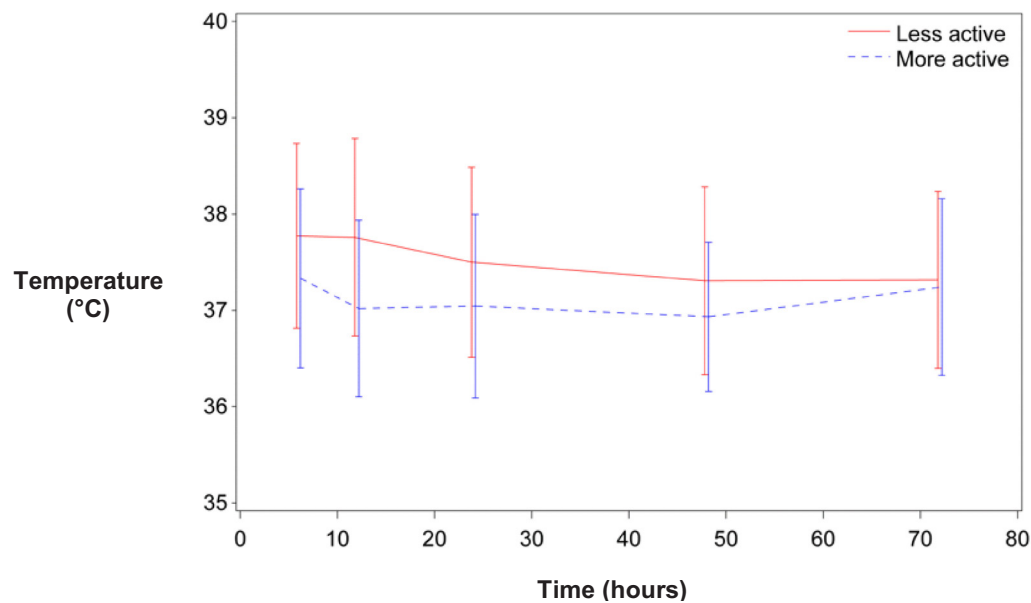


Fig. 1 Body temperature over time with more active fever management vs. less active fever management. **P* value for the interaction term evaluating temperature difference by time from randomisation was <0.001 indicating a statistically significant variation in the temperature difference by treatment over time

or not; infection present or not; high fever (≥ 39.5 °C or < 39.5 °C); age (≥ 75 years or < 75 years); APACHE-II score ≥ 25 or < 25 ; and physical cooling included in the study intervention or physical cooling not included in the study intervention.

Temperature was analysed using a mixed linear model with a power exponential structure for the correlation between repeated measurements and the time by randomisation interaction term used to estimate temperature differences between randomised treatments at each time point.

A two-sided *P* value < 0.05 was considered to indicate statistical significance and estimates are shown with 95% confidence intervals (CI). No adjustment was made for multiple comparisons.

SAS version 9.4 (SAS Institute Inc., Cary, NC, USA) was used for analyses.

Results

Data sources

Individual-level patient data were obtained for 5 [6–8, 14, 15] of 13 randomised controlled trials identified in

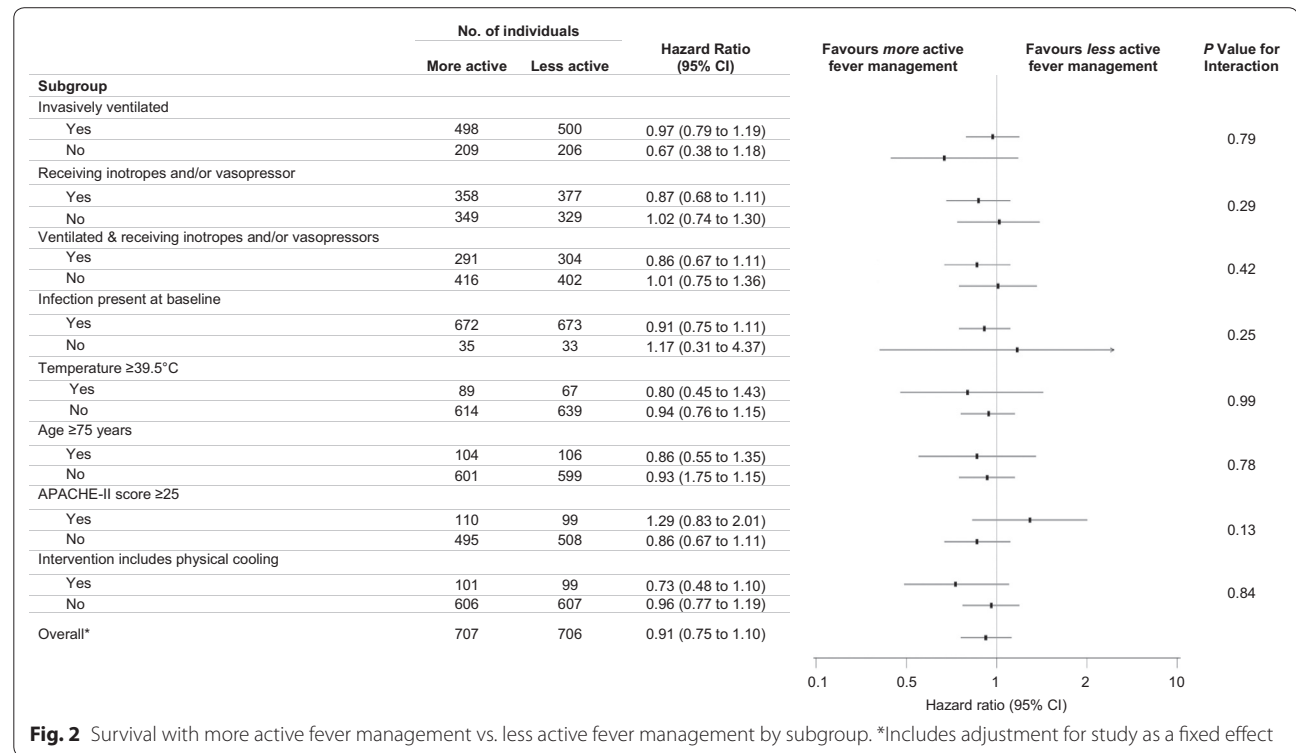
the systematic review conducted for our aggregate data meta-analysis [9]. This included the three largest trials [6–8] and resulted in data from 1413 of 1780 (79.4%) of the participants in the original trials being included in this analysis. Details of included trials and those trials from which data could not be obtained are shown in the ESM.

Patient characteristics

Of the 1413 participants included in this analysis, 707 were assigned to more active fever management and 706 were assigned to less active fever management. The study groups had similar characteristics at baseline (Table 1). More than 95% of participants were suspected to have an infection at baseline.

Effects of fever management on body temperature

Patients assigned to more active fever management had statistically significantly lower body temperature than patients assigned to less active fever management (Fig. 1). The effect of study treatment on body temperature varied with time with a maximum temperature difference

**Table 2 Outcomes**

	More active fever control (n = 707)	Less active fever control (n = 706)	Treatment effect estimate ^a (95% CI)	P value
Outcomes				
			Hazard ratio	
Survival (days), median (95% CI)	202 (146–N/A)	212 (143–390)	0.91 (0.75–1.10)	0.32
			Adjusted hazard ratio ^b	
			0.96 (0.77–1.19)	0.69
			Odds ratio	
ICU mortality—n/N (%)	98/707 (13.9)	121/706 (17.1)	0.78 (0.58–1.04)	0.09
			Adjusted odds ratio ^a	
			0.76 (0.52–1.09)	0.13
			Ratio of geometric means (95% CI)	
ICU length of stay (days), geometric mean (95% CI)	5.7 (5.2–6.2)	5.6 (5.2–6.2)	1.01 (0.89–1.14)	0.92
			Ratio of geometric means (95% CI)	
Hospital length of stay (days), geometric mean (95% CI)	14.6 (13.5–15.9)	13.5 (12.3–14.8)	1.08 (0.96–1.23)	0.19

IQR interquartile range, CI confidence interval

^a The hazard ratio from the primary survival analysis includes an adjustment for study as a fixed effect. The widths of the confidence intervals for secondary analyses have not been adjusted for multiplicity and the intervals should not be used to infer definitive differences between the groups

^b Adjusted for age, sex, study, and APACHE-II score; APACHE-II data were not available for the Schortgen et al. trial. Additional adjusted analyses are presented in the ESM

of 0.73°C (95% CI 0.63 – 0.83°C) between temperature groups evident at 12 h post randomisation.

Survival and mortality

In the analysis addressing the primary hypothesis that more active fever management would increase survival among patient subgroups with limited physiological

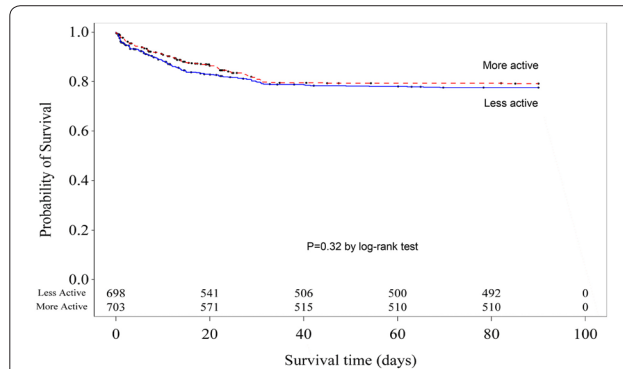


Fig. 3 Kaplan-Meier survival estimates of the probability of survival. Because the number of observations beyond day 90 is small, this figure is truncated at day 90 with data censored at day 90 if death had not occurred by then. An expanded Kaplan-Meier survival plot including all available data points is provided with the electronic supplementary material

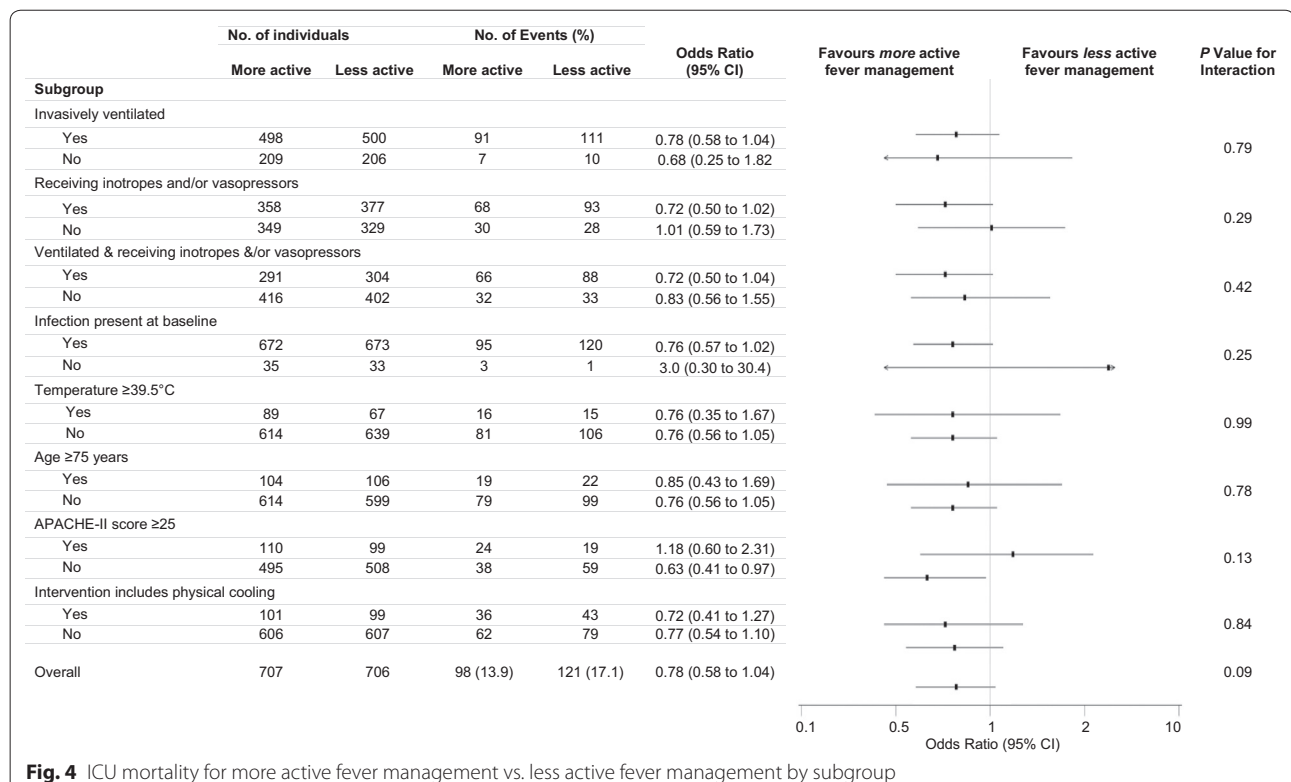
reserves such as older patients, patients with higher illness acuity, and those requiring organ support, we found no statistically significant heterogeneity of treatment effect in any of the pre-specified subgroups (Fig. 2). Similarly, there was no heterogeneity of survival response by treatment allocation in patients with and without infections (Fig. 2). Overall, more active fever management did

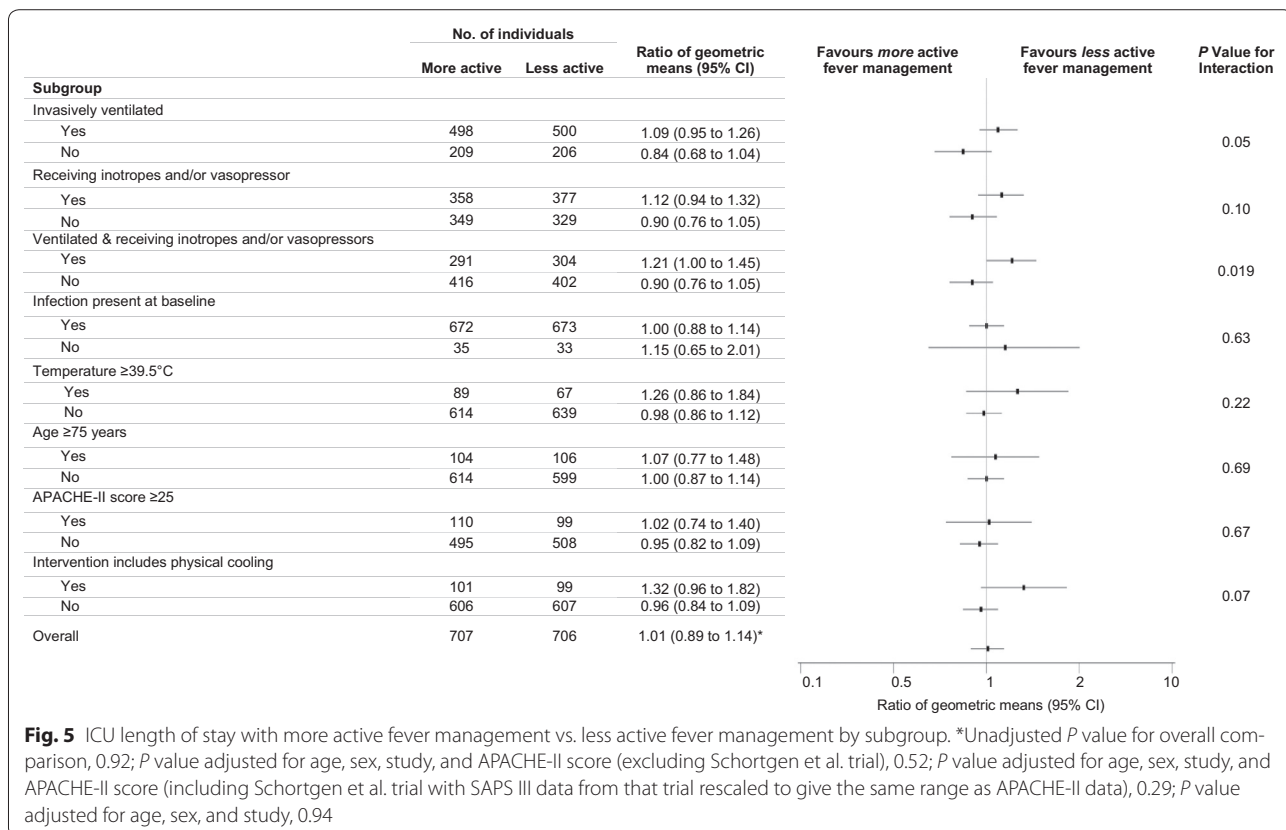
not result in a statistically significant difference in survival time compared with less active fever management [hazard ratio; 0.91; (95% CI 0.75–1.10), $P=0.32$] (Table 2, Fig. 3, Fig. S1 ESM). Findings were similar in analyses adjusting for pre-specified baseline covariates (Table 2 and Fig. 3) and in sensitivity analyses (ESM). Moreover, the estimates of the hazard ratios related to survival from the sensitivity analyses were more-or-less identical treating the studies as fixed effects (as was pre-specified) or as random effects.

There was no statistically significant heterogeneity in the effect of more active compared with less active fever management on ICU mortality in any of the pre-specified subgroups (Fig. 4). A total of 98 of 707 patients (13.9%) assigned to more active fever management and 121 of 706 patients (17.1%) assigned to less active fever management died in the ICU [absolute mortality difference, -3.3% points (95% CI -7.1 to 0.5% points); odds ratio, 0.78 (95% CI 0.58–1.04), $P=0.09$] (Table 2). Findings were similar in analyses adjusting for pre-specified baseline covariates (Table 2) and in sensitivity analyses (ESM).

Length of stay

Overall, ICU length of stay and hospital length of stay were similar between treatment groups (Table 2).





However, there was statistically significant heterogeneity of response to treatment in relation to length of stay variables in some subgroups (Fig. 5 and Fig. S2, ESM). In each case where significant heterogeneity in response to treatment was observed, the length of stay was statistically significantly shorter in a subgroup where the point estimate for ICU mortality risk favoured more active fever management. In a post hoc interaction analysis of ICU and hospital length of stay by treatment allocation, there was statistically significant heterogeneity in response in survivors compared with non-survivors (ESM). Compared with less active fever management, more active fever management was associated with longer ICU and hospital length of stay in patients who died in ICU and with shorter ICU and hospital length of stay in patients who survived ICU.

Discussion

In this individual-level patient data meta-analysis of randomised controlled trials, more active fever management did not increase survival compared with less active fever management in critically ill adults either overall or in those with limited physiological reserves. Survival by treatment group was similar in a range of subgroup pairs that divided the study population into groups based on

age, illness severity, on receipt of specific organ supports, and in the presence or absence of high fever at baseline.

Overall, effect size estimates in relation to ICU mortality based on the 95% CI were consistent with an absolute effect on ICU mortality with active fever management ranging from a decrease of 7.1% points to an increase of 0.5% points. Although we observed statistically significant heterogeneity of treatment effect in relation to ICU and hospital length of stay effects, the interpretation of these findings is complicated because length of stay can be reduced by more rapid recovery or by early death. Moreover, a reduction in mortality, even a non-statistically significant one, can be associated with a statistically significant increase in length of stay when survivors have longer average lengths of stay than non-survivors. In patients who were receiving invasive mechanical ventilation, those receiving inotropes and/or vasopressors, or those receiving both of these, a relative increase in hospital length of stay was associated with lower ICU mortality based on point estimates. We also found that, compared with less active temperature management, more active temperature was associated with reduced ICU and hospital length of stay in patients who survived ICU and with increased length of stay in patients who died in ICU.

Our study is consistent with two recent aggregate data metaanalyses [9, 16] evaluating fever control in adult ICU patients; however, it extends their findings because the use of individual-level patient data allowed us to conduct analyses adjusting for important baseline covariates and to accurately evaluate subgroups of interest defined based on pre-randomisation characteristics.

Our study has a number of limitations. Because our analyses were not adjusted for multiple comparisons, they should be considered exploratory and should not be used to infer definitive treatment effects. Although we did not demonstrate statistically significant heterogeneity of treatment effect on survival for subgroups of interest, confidence intervals around hazard ratios were generally wide and the possibility of clinically important differences in survival responses by subgroup cannot be excluded. In particular, as nearly all patients were suspected of having an infection at baseline, our findings effectively neither confirm nor refute the hypothesis that the presence of infection is an important factor in determining the efficacy of active fever management [17]. We were only able to obtain data from 5 out of 13 trials identified in our recent systematic review. However, the three largest trials [6–8] conducted were included in our analysis and 79.4% of all potential data from prior randomised controlled trials were analysed. Most of the studies where data were not available were small single-centre studies. The studies included in our analysis used a variety of different therapies and it is not known whether these therapies have equivalent effects on patient outcomes. Nevertheless, as all therapies evaluated are given to patients to treat fever, we submit that combining trials in an IPDMA has both face validity and clinical relevance.

In conclusion, our findings do not support the hypothesis that more active fever management increases survival compared with less active fever management in patients with limited physiological reserves. However, as point estimates for the effect of active fever management on ICU mortality encompass potentially clinically important effects, further clinical trials are justified. The significant heterogeneity in treatment effects on length of stay in subgroups based on the receipt of organ support, combined with the finding that more active fever management increases ICU and hospital length in patients who die in ICU, and reduces length of stay in patient who survive ICU, suggests that further research in patients receiving organ support may be of interest.

Electronic supplementary material

The online version of this article (<https://doi.org/10.1007/s00134-019-05553-w>) contains supplementary material, which is available to authorized users.

Author details

¹ Intensive Care Unit, Wellington Hospital, Capital and Coast District Health Board, Wellington, New Zealand. ² Medical Research Institute of New Zealand, Wellington, New Zealand. ³ Austin Hospital, Heidelberg, VIC, Australia. ⁴ Vanderbilt University Medical Center, Nashville, TN, USA. ⁵ Department of Critical Care Medicine, Cumming School of Medicine, University of Calgary, Calgary, AB, Canada. ⁶ Adult Intensive Care Unit, Centre Hospitalier Intercommunal de Créteil, Créteil, France. ⁷ The George Institute for Global Health, Sydney, Australia. ⁸ University of Otago, Wellington, New Zealand. ⁹ Bankstown Hospital, University of New South Wales, South Western Sydney Local Health District, Sydney, NSW, Australia.

Acknowledgements

This research was conducted during the tenure of a Clinical Practitioner Research Fellowship from the Health Research Council of New Zealand held by PY. The Medical Research Institute of New Zealand is supported by independent research organisation funding from the Health Research Council of New Zealand.

Compliance with ethical standards

Conflicts of interest

Dr. Paul Young and Dr. Manoj Saxena report receiving speaker's fees from Bard Medical Pty. Dr. Gordon Bernard reports having an equity stake and is on the Board of Directors for Cumberland Pharmaceuticals, Nashville, TN, makers of intravenous ibuprofen (Caldolor).

Ethical approval

An approval by an ethics committee was not applicable.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Received: 6 December 2018 Accepted: 29 January 2019

Published online: 11 February 2019


References

1. Golding R, Taylor D, Gardner H, Wilkinson JN (2016) Targeted temperature management in intensive care—do we let nature take its course? *JICS* 17:154–159
2. Manthous CA, Hall JB, Olson D, Singh M, Chatila W, Pohlman A, Kushner R, Schmidt GA, Wood LD (1995) Effect of cooling on oxygen consumption in febrile critically ill patients. *Am J Respir Crit Care Med* 151:10–14
3. Laupland KB, Shahpori R, Kirkpatrick AW, Ross T, Gregson DB, Stelfox HT (2008) Occurrence and outcome of fever in critically ill adults. *Crit Care Med* 36:1531–1535
4. Laupland KB, Zahar JR, Adrie C, Schwebel C, Goldgran-Toledano D, Azoulay E, Garrouste-Orgeas M, Cohen Y, Jamali S, Souweine B, Darmon M, Timsit JF (2012) Determinants of temperature abnormalities and influence on outcome of critical illness. *Crit Care Med* 40:145–151
5. Niven DJ, Stelfox HT, Shahpori R, Laupland KB (2013) Fever in adult ICUs: an interrupted time series analysis. *Crit Care Med* 41:1863–1869
6. Young P, Saxena M, Bellomo R, Freebairn R, Hammond N, van Haren F, Holliday M, Henderson S, Mackle D, McArthur C, McGuinness S, Myburgh J, Weatherall M, Webb S, Beasley R (2015) Acetaminophen for fever in critically ill patients with suspected infection. *N Engl J Med* 373:2215–2224
7. Bernard GR, Wheeler AP, Russell JA, Schein R, Summer WR, Steinberg KP, Fulkerson WJ, Wright PE, Christman BW, Dupont WD, Higgins SB, Swindell BB (1997) The effects of ibuprofen on the physiology and survival of patients with sepsis. The Ibuprofen in Sepsis Study Group. *N Engl J Med* 336:912–918
8. Schortgen F, Clabault K, Katsahian S, Devaquet J, Mercat A, Deye N, Del-lamonica J, Bouadma L, Cook F, Beji O, Brun-Buisson C, Lemaire F, Brochard L (2012) Fever control using external cooling in septic shock: a randomized controlled trial. *Am J Respir Crit Care Med* 185:1088–1095
9. Dallimore J, Ebmeier S, Thayabaran D, Bellomo R, Bernard G, Schortgen F, Saxena M, Beasley R, Weatherall M, Young P (2018) Effect of active

- temperature management on mortality in intensive care unit patients. *Crit Care Resusc* 20:150–163
10. Young PJ, Nielsen N, Saxena M (2017) Fever control. *Intensive Care Med* 44:227–230
 11. Young PJ, Saxena M (2014) Fever management in intensive care patients with infections. *Crit Care* 18:206
 12. Knaus WA, Draper EA, Wagner DP, Zimmerman JE (1985) APACHE II: a severity of disease classification system. *Crit Care Med* 13:818–829
 13. Metnitz PG, Moreno RP, Almeida E, Jordan B, Bauer P, Campos RA, Iapichino G, Edbrooke D, Capuzzo M, Le Gall JR (2005) SAPS 3—from evaluation of the patient to evaluation of the intensive care unit. Part 1: Objectives, methods and cohort description. *Intensive Care Med* 31:1336–1344
 14. Niven DJ, Stelfox HT, Leger C, Kubes P, Laupland KB (2013) Assessment of the safety and feasibility of administering antipyretic therapy in critically ill adults: a pilot randomized clinical trial. *J Crit Care* 28:296–302
 15. Saxena MK, Taylor C, Billot L, Bompont S, Gowardman J, Roberts JA, Lipman J, Myburgh J (2015) The effect of paracetamol on core body temperature in acute traumatic brain injury: a randomised, controlled clinical trial. *PLoS One* 10:e0144740
 16. Drewry AM, Ablordeppey EA, Murray ET, Stoll CRT, Izadi SR, Dalton CM, Hardi AC, Fowler SA, Fuller BM, Colditz GA (2017) Antipyretic therapy in critically ill septic patients: a systematic review and meta-analysis. *Crit Care Med* 45:806–813
 17. Young PJ, Saxena M, Beasley R, Bellomo R, Bailey M, Pilcher D, Finfer S, Harrison D, Myburgh J, Rowan K (2012) Early peak temperature and mortality in critically ill patients with or without infection. *Intensive Care Med* 38:437–444

ORIGINAL

Early PREDiction of sepsis using leukocyte surface biomarkers: the ExPRES-sepsis cohort study

Manu Shankar-Hari^{1,2*} , Deepankar Datta³, Julie Wilson^{1,2}, Valentina Assi^{4,5}, Jacqueline Stephen⁵, Christopher J. Weir^{4,5}, Jillian Rennie³, Jean Antonelli⁴, Anthony Bateman⁶, Jennifer M. Felton³, Noel Warner^{7,8}, Kevin Judge^{7,8}, Jim Keenan^{7,8}, Alice Wang^{7,8}, Tony Burpee^{7,8}, Alun K. Brown², Sion M. Lewis², Tracey Mare², Alistair I. Roy^{7,8}, John Wright⁹, Gillian Hulme¹⁰, Ian Dimmick¹⁰, Alasdair Gray^{4,11}, Adriano G. Rossi³, A. John Simpson¹², Andrew Conway Morris¹³ and Timothy S. Walsh^{3,4,5}

© 2018 Springer-Verlag GmbH Germany, part of Springer Nature and ESICM

Abstract

Purpose: Reliable biomarkers for predicting subsequent sepsis among patients with suspected acute infection are lacking. In patients presenting to emergency departments (EDs) with suspected acute infection, we aimed to evaluate the reliability and discriminant ability of 47 leukocyte biomarkers as predictors of sepsis (Sequential Organ Failure Assessment score ≥ 2 at 24 h and/or 72 h following ED presentation).

Methods: In a multi-centre cohort study in four EDs and intensive care units (ICUs), we standardised flow-cytometric leukocyte biomarker measurement and compared patients with suspected acute infection (cohort-1) with two comparator cohorts: ICU patients with established sepsis (cohort-2), and ED patients without infection or systemic inflammation but requiring hospitalization (cohort-3).

Results: Between January 2014 and February 2016, we recruited 272, 59 and 75 patients to cohorts 1, 2, and 3, respectively. Of 47 leukocyte biomarkers, 14 were non-reliable, and 17 did not discriminate between the three cohorts. Discriminant analyses for predicting sepsis within cohort-1 were undertaken for eight neutrophil (cluster of differentiation antigens (CD) CD15; CD24; CD35; CD64; CD312; CD11b; CD274; CD279), seven monocyte (CD35; CD64; CD312; CD11b; HLA-DR; CD274; CD279) and a CD8 T-lymphocyte biomarker (CD279). Individually, only higher neutrophil CD279 [OR 1.78 (95% CI 1.23–2.57); $P=0.002$], higher monocyte CD279 [1.32 (1.03–1.70); $P=0.03$], and lower monocyte HLA-DR [0.73 (0.55–0.97); $P=0.03$] expression were associated with subsequent sepsis. With logistic regression the optimum biomarker combination was increased neutrophil CD24 and neutrophil CD279, and reduced monocyte HLA-DR expression, but no combination had clinically relevant predictive validity.

Conclusions: From a large panel of leukocyte biomarkers, immunosuppression biomarkers were associated with subsequent sepsis in ED patients with suspected acute infection.

Clinical trial registration: NCT02188992.

Keywords: Sepsis, Infection, Mortality, Cohort study, Biomarker, risk prediction

*Correspondence: manu.shankar-hari@kcl.ac.uk

² Guy's and St Thomas' NHS Foundation Trust, London SE17EH, UK
Full author information is available at the end of the article

Introduction

Sepsis is life-threatening organ dysfunction caused by a dysregulated host response to infection [1]. Host immune responses result from leukocytes sensing pathogen- and tissue damage-associated danger signals [2, 3]. Sepsis-related immune responses involve both humoral and leukocyte components of the innate and adaptive immune systems, with excessive inflammation and immunosuppression occurring simultaneously in most patients [2, 3]. These are thought to influence the resulting clinical phenotypes and outcomes [3, 4].

Leukocyte responses in sepsis measured using flow cytometry detect leukocyte biomarkers, including surface markers and/or leukocyte subsets [5]. Previous flow cytometry-based leukocyte biomarker studies in sepsis were mostly small, single-centre studies in patients with sepsis, typically focusing on a limited panel of biomarkers. These studies rarely evaluated biomarker reliability and reproducibility, which is methodologically and clinically relevant as it influences diagnostic validity [6]. In addition, few studies used robust unbiased designs to assess predictive ability for clinically relevant outcomes in unselected populations with suspected infections *prior* to developing organ dysfunction and established sepsis.

We hypothesized that among patients with clinically suspected acute infection, but without established sepsis, leukocyte biomarkers would identify patients who subsequently deteriorate clinically and develop sepsis, when measured within a few hours of presentation to the emergency department (ED). Our study objectives were: (1) to identify reliable leukocyte biomarkers; (2) to ascertain which of the reliable biomarkers could discriminate [6] acutely unwell patients with suspected infection from patients with community acquired sepsis-related critical illness in the intensive care unit (ICU) and/or ED patients with non-infective acute illness requiring hospitalisation; and (3) to ascertain whether any of the reliable biomarkers with cross-cohort discrimination could predict which patients with suspected infection in the ED subsequently develop sepsis. We also undertook a post hoc extreme phenotype analysis [7], to compare the biomarker profiles between acutely unwell patients with suspected infection who subsequently developed most severe illness with those who recovered rapidly.

Methods

Study sites and ethics

We performed a prospective, multi-centre, observational cohort study at four sites in the United Kingdom. Ethical approval was granted by the Scotland A/Oxford C Research Ethics Committees (13/SS/0023;13/

Take-home message

In this first study of standardised multi-site flow cytometry in acutely unwell patients with suspected infections attending emergency departments, we explored which of 47 leukocyte biomarkers reliably discriminates which patients develop sepsis over the next 3 days, defined according to the Sepsis-3 sepsis criteria.

After highlighting the importance of test reliability (14 biomarkers lacked measurement reliability) and comparator cohorts (a further 17 biomarkers did not discriminate acutely unwell patients with suspected infection from patients with established sepsis-related critical illness and/or non-infective acute illness), we found that none of the remaining 16 biomarkers had clinically relevant predictive ability for subsequent sepsis or other important clinical outcomes. However, markers of early immune suppression (neutrophil and monocyte CD274 and CD279; monocyte HLA-DR) had the strongest associations with clinical outcomes. The optimum biomarker combination associated with clinical deterioration to sepsis was increased neutrophil CD24 and CD279 and reduced monocyte HLA-DR expression.

SC/0266). Consent was provided by patients or surrogate decision-makers according to capacity. We registered the study (NCT02188992) and published the protocol including the analysis plan [8].

Cohort definitions and eligibility criteria

We recruited three distinct patient cohorts using an a priori sampling method to achieve similar age and sex profiles across the ED cohorts. Detailed inclusion/exclusion criteria are listed in the electronic supplement and published protocol (emethods-1) [8]. Cohort-1 comprised acutely unwell patients with suspected infection and systemic inflammation presenting to ED and formed the “discovery cohort”. Patients considered by clinical teams to already have established severe sepsis and/or require ICU admission when screened were excluded. Cohort-2 comprised ICU patients with established community acquired sepsis-related critical illness and formed the “true positive” cohort. Cohort-3 comprised acutely ill patients presenting to ED without infection or systemic inflammation, but requiring hospitalization and formed the “true negative” cohort. Inclusion criteria used throughout the study were based on the sepsis definitions by Levy et al. [9], as our study was designed prior to the Sepsis-3 definitions [1, 10]. All ED patients were enrolled within 12 h of hospital presentation. For all cohorts, we excluded patients with acute pancreatitis, haematological malignancy, chemotherapy in the past 2 weeks, myelodysplastic syndromes, known neutropenia, HIV infection, viral hepatitis infection, pregnancy, blood transfusion > 4 units in the past week, oral corticosteroids for > 24 h prior to enrolment, or a decision not to have active therapy/for palliative care [8].

Leukocyte surface biomarkers and cross-site standardization of flow cytometry

We devised five separate flow cytometry panels to assess 47 leukocyte biomarkers with biological plausibility for having predictive validity for subsequent sepsis (eMethods-1; eTable-1; eFigure-1). We developed, standardized and harmonized flow cytometry procedures across all four study sites [8]. We performed flow cytometry within 4 h of sample acquisition. All anti-human antibodies conjugated to fluorochromes for flow cytometry were from the same batch and clones [all Becton–Dickinson Biosciences (BDB)], standardized on the same platform (FACSCanto II; BDB, San Jose, CA, USA), using a common batch of Cytometer Setup and Tracking beads with the same beads for daily internal quality controls, at all clinical sites. All flow cytometry standard (FCS) files were read by expert technicians using standardized gating procedures developed for each biomarker prior to analysis. The gating strategy for estimating median fluorescence intensity (MFI) or proportions is reported in eMethods-1. All FCS analysis technicians were blinded from clinical data.

Sample size

We based sample size estimates on the confidence interval (CI) widths for positive and negative predictive values (PPV and NPV). The initial design had a primary outcome of septic shock, with an estimated event rate of 5–10% in cohort-1 [11, 12]. For a range in test performance for PPV/NPV of 50–90% we planned a sample size of: cohort-1, $n=300$; cohort-2, $n=100$; and cohort-3, $n=100$, to give a CI width between $\pm 4.6\%$ to $\pm 9.8\%$ for PPV and $\pm 3.4\%$ to $\pm 6.3\%$ for NPV. At an interim analysis of clinical event rates, the incidence of septic shock was substantially lower than anticipated. We decided by consensus to change the primary outcome to severe sepsis (and subsequently adopted the sepsis-3 sepsis criteria [1] of Sequential Organ Failure Assessment (SOFA) score ≥ 2), with critical care admission a key secondary outcome, to ensure adequate clinically relevant events in the discriminant analyses. These changes occurred *prior to* study completion and were reported in the published protocol [8].

Statistical analysis

The primary study cohort was cohort-1. The primary exposure was suspected infection. The cohorts-2 and 3 were comparator populations for cross-cohort discrimination and biomarker selection.

Outcomes

The primary outcome was sepsis, defined as SOFA score ≥ 2 at 24 h and/or 72 h following presentation

to hospital in patients with suspected infection in the ED (cohort-1) [1]. Secondary outcomes were: critical care admission or death within 72 h of presentation; SOFA ≥ 4 at 24 h and/or 72 h following presentation to hospital; development of septic shock; discharge home within 72 h; discharge to home or in hospital with no organ failure within 72 h; death from sepsis; confirmed infection and length of hospital stay [8]. All cohort-1 data are based on blood samples taken in the ED after recruitment.

Biomarkers selection strategy

Our analytic approach to discover biomarkers with potential diagnostic discrimination for risk of subsequent sepsis occurred in three a priori planned stages and one post hoc analysis.

Stage one: reliability

Inter- and intra-reader reliability for 47 different biomarkers was established according to the protocol [8]. To be included in subsequent evaluation stages, biomarkers needed to demonstrate both inter- and intra-reader reliability at the pre-defined intra-class correlation coefficient (ICC) between readers ≥ 0.9 ; see Fig. 1 and eMethods-2; eTable-2). For intra-reader reliability the ICC for each reader was calculated as the ratio of within-reader variability to the total variance (within-reader plus residual variance) from the normal linear mixed model. For inter-reader reliability the ICC was calculated as the ratio of between-reader variability to the total variance (between-reader plus residual variance) from the normal linear mixed model. Reliability analyses were done prior to linking leukocyte biomarkers data and clinical outcome data.

Stage two: cross-cohort discrimination

For reliable biomarkers, statistically significant inter-group differences between the three cohorts were explored using Kruskal–Wallis analysis of variance (ANOVA) tests (eTable-3) and visual inspection of data. Biomarkers that discriminated between cohort-1 and either cohort-2 (true-positive) and/or cohort-3 (true negative) and had variability within cohort-1 consistent with potential to discriminate clinical outcomes were selected for Stage-3 analysis. Other factors considered were cell counts, the magnitude of MFI, and potential linkage and co-linearity between groups of biomarkers. This was done in consensus meetings by researchers blinded from clinical outcomes within cohort-1.

Stage three: prediction of clinical outcomes in cohort-1

Within cohort-1 patients, the ability of the selected biomarkers to predict the primary and secondary outcomes

was calculated using univariate logistic regression. For the secondary outcomes of death from sepsis, septic shock and length of stay, we provided a descriptive summary as per the analysis plan [8]. The odds ratio (OR) for the outcome per standard deviation increase in biomarker, receiver operating characteristic (ROC) curves, and area under ROC curve (AUROC) were used to assess predictive ability. Youden's index identified the optimal cut-off point for each marker [13]. Candidate biomarkers that showed consistent inclusion were then taken forward for multivariable modelling.

We used best subsets regression [14] to identify optimal combinations of predictive markers. Specifically, models containing a given number of biomarkers were fitted for all potential biomarker combinations. The five best-fitting models of a given size, according to the Chi squared score statistic, were identified. Biomarkers that consistently appeared in the best-fitting models were selected for the final model. The change in Chi squared score statistic between the best fitting models containing different numbers of biomarkers was used to determine the number of biomarkers to be included in the final model. Linearity of biomarker associations on the logistic scale was checked using plots of deviance residuals. Based on consistency and model fit we identified optimal combinations of predictive markers.

Post hoc extreme phenotype comparison

On the recommendation of a pre-planned independent expert group (see eTable-4), we compared biomarker profiles between sub-populations within cohort-1 with extreme clinical phenotypes of organ dysfunction and outcome to further explore associations for the biomarkers evaluated. We defined *well* and *sick* extreme phenotypes [7] by consensus among clinical investigators using clinical data without knowledge of group differences in biomarkers (eFigure-2). The *well* phenotype had no positive microbiology, a SOFA score ≤ 2 at 24 and 72 h post-enrolment and were either discharged home by 72 h or were in hospital but no longer receiving antibiotics. The *sick* phenotype had a confirmed infection, SOFA score ≥ 2 at both 24 and 72 h post-enrolment and were still in hospital and receiving antibiotics at 72 h. We compared biomarker expression between the two phenotypes using two-sample t-tests or Mann-Whitney tests as appropriate, applying Bonferroni correction for multiple testing.

For additional comparison, we also measured C-reactive protein (CRP) and procalcitonin (PCT) concentrations at the same time point for cohort-1 patients, given the widespread clinical use of these biomarkers in assessing infection. We constructed ROC curves for CRP and PCT and estimated similar univariate predictive

performance characteristics of these for outcomes reported, to enable direct comparison of predictive validity with the more novel biomarkers.

Results

Patient characteristics

Between January 2014 and February 2016, we recruited 272, 59 and 75 patients ($N=406$) to cohorts 1, 2, and 3, respectively. The clinical characteristics for the three cohorts and the cohort-1 outcomes are shown in Table 1. Cohorts-1 and 3 had a similar age and sex distribution. Cohort-2 patients tended to be older. The primary outcome in cohort-1, clinical deterioration to sepsis, occurred in 139 patients (51.1%).

Stage one: reliability

The step-wise assessment of intra-reader and then inter-reader reliability resulted in rejection of 14 biomarkers as non-reliable, leaving 33 reliable biomarkers for cross-cohort comparison (Fig. 1; eTable-2).

Stage two: cross-cohort discrimination

Statistical comparison, expert review, and cohort-1 data distribution resulted in rejection of a further 17 biomarkers (Fig. 1; eTable-2; eTable-3). The cross-cohort comparisons plots for the 16 selected biomarkers are shown in eFigure-3. Based on the stage-1 and -2 selections, eight neutrophil biomarkers [cluster of differentiation antigens (CD) CD15; CD24; CD35; CD64; CD312; CD11b; CD274; CD279], seven monocyte biomarkers (CD35; CD64; CD312; CD11b; HLA-DR; CD274; CD279) and one CD8 T-lymphocyte biomarker (CD279) were selected for evaluation of discrimination for clinical outcomes. Biological relevance of these markers in sepsis are summarized in Table 2.

Stage three: prediction of clinical outcomes in cohort-1

Most biomarkers lacked any clinically or statistically significant discrimination for predicting primary and secondary outcomes within cohort-1 patients. Amongst the individual biomarkers, clinical deterioration to sepsis was associated with higher neutrophil CD279 expression, higher monocyte CD279 expression and lower monocyte HLA-DR expression. The optimal MFI cutoff for neutrophil CD279 was 239 [sensitivity 0.88 (95% confidence interval 0.82–0.93); specificity 0.35(0.26–0.43)]; for monocyte CD279 was 141 [sensitivity 0.83(0.77–0.90); specificity 0.39(0.30–0.47)]; and for monocyte HLA-DR was 3572 [sensitivity 0.43(0.34–0.51); specificity 0.69(0.60–0.77)]. Although these associations were statistically significant, discriminant ability was poor and unlikely to be clinically useful in isolation.

Table 1 Cohort characteristics and cohort-1 outcomes

	Cohort-1 (infected ED cohort) N = 272	Cohort-2 (ICU-septic) N = 59	Cohort-3 (non-infected ED controls) N = 75
Cohort characteristics			
Age in years mean (SD)	62.1 (19.1)	67.9 (12.8)	61.6 (20.0)
Female N (%)	133 (48.9%)	23 (39.0%)	33 (44.0%)
FCI Score median (IQR)	2 (1,3)	2 (1,4)	1 (0,2)
White cell count median (IQR)			
Total	13.5 (10.7, 16.2)	16.9 (10.1, 19.6)	7.7 (6.4, 9.1)
Neutrophils	11.2 (8.5, 14.1)	14.1 (8.2, 17.5)	4.9 (4.1, 6.4)
Lymphocytes	0.9 (0.6, 1.4)	0.9 (0.6, 1.3)	1.7 (1.3, 2.1)
C-reactive protein median (IQR)	64 (21,168)	212 (86,309)	13 (2,27)
Procalcitonin Median (IQR)	29.4 (0.0, 337.3)	No data	No data
Confirmed infection	238 (87.5%)	59 (100%)	0
qSOFA score ≥ 2			
At ED presentation	44 (16.2%)	No data	No data
At 24 h	6 (2.2%)		
At 72 h	5 (1.8%)		
APACHE II score median (IQR)	9 (6, 13)	16 (12, 21)	6 (3, 9)
SOFA score median (IQR)	2 (1, 3)	7 (5, 9)	1 (1, 2)
Site of infection N (%)			
Respiratory	124 (45.6%)		
Urinary	44 (16.2%)		
Unknown	40 (14.7%)		
Musculoskeletal, skin and soft tissue	32 (11.7%)		
Abdominal (including biliary)	28 (11.0%)		
Neurological	4 (1.5%)		
Outcomes for Cohort-1			
Primary outcome ¹			
SOFA ≥ 2 at 24 or 72 h	139 (51.1%)		
Secondary outcomes			
ICU admission or death within 72 h of hospitalization	22 (8.1%)		
SOFA ≥ 4 at 24 or 72 h	36 (13.2%)		
Discharged home within 72 h of hospitalization	86 (31.6%)		
Discharged home or in hospital with no organ failure	148 (54.4%)		
Hospital mortality N (%)	1 (0.4%)		
Development of septic shock	1 (0.4%)		
Organ support			
On antibiotics at 72 h	144 (52.9%)		
Vasopressors	2 (0.7%)		
Ventilation invasive	2 (0.7%)		
Ventilation non-invasive	5 (1.8%)		
Hospital length of stay (days) median (IQR)	5 (2, 9)		

FCI functional co-morbidity index, qSOFA quick sepsis organ failure assessment, SOFA sepsis organ failure assessment, APACHE II Acute Physiology And Chronic Health Evaluation II, ICU intensive care unit, ED emergency department

¹ N = 3 missing data for primary outcome

With best subsets logistic regression, the optimum combination for predicting clinical deterioration to sepsis included increased neutrophil CD24; increased neutrophil CD279; and reduced monocyte HLA-DR expression

[sensitivity 0.72(0.64–0.79); specificity 0.56(0.48–0.65)]. With best subsets logistic regression, the optimum combination for predicting the secondary outcome of discharge to home within 72 h, included increased neutrophil CD15,

Table 2 Biological relevance in sepsis patients of the reliable cell surface markers with discriminant value identified in cohort-1

Cell surface markers	Marker positive leukocytes in our study	Biological relevance in sepsis [2, 3, 22, 25, 32, 39–42]	Our key inferences
CD15	Neutrophil	Expressed on all myeloid cells and from the promyelocyte stage onwards on neutrophils. Although monocytes express CD15 at low levels, we were gating CD15 ^{hi} granulocytes	Alongside CD14, CD16, CD11b, CD15, is a marker for myeloid derived suppressor cells [43, 44], which is implicated in suppressing T cell function
CD24	Neutrophil	Expressed on mature granulocytes and B cells; down-regulated on neutrophils in sepsis; induces neutrophil apoptosis which is delayed in sepsis	CD16 low/CD14 negative/CD24 positive myeloid-derived suppressor cells are cytotoxic to T cells [20]. Immature granulocytes in peripheral circulation in sepsis is associated with greater risk of death [45]
CD35	Neutrophil Monocytes	Receptors of complement activation (RCA) family expressed on leukocytes; potentially discriminates sepsis from inflammation	Understanding of major roles of CD35 alterations in sepsis is unclear
CD64	Neutrophil Monocytes	Fc gamma receptor expressed on leukocytes; Patients with sepsis have increased expression of CD64 has been consistently reported	Despite this association, CD64 as a single marker has limited diagnostic performance in sepsis [23, 40].
CD11b	Neutrophil Monocytes	Role in adhesive interactions of monocytes; macrophages and granulocytes; mediating the uptake of complement-coated particles; increased in sepsis following neutrophil activation	Neutrophil and monocyte increase in CD11b is inconsistent in the literature [46, 47]. Tissue resident CD11b positive T cells, secrete interferon gamma and may influence local host defence mechanisms in bacterial infections [48]
CD312	Neutrophil Monocytes	human myeloid-restricted class B seven-span transmembrane (TM7) subfamily of G-protein coupled receptors; acutely altered in sepsis secondary to leukocyte activation	Understanding of major roles of CD312 alterations in sepsis is unclear
CD274	Neutrophil Monocytes	PD-1 and PDL-1 form a checkpoint inhibitor complex and are considered markers of sepsis related immunosuppression. In sepsis, neutrophils, monocytes and lymphocytes express elevated levels of with CD274 and CD279 [2, 30, 31]. In sepsis, neutrophils are thought to impair T cell function through PD-L1 mechanism [33, 49]	Recently, it has been shown that the increasing functional deficit in multiple innate and adaptive immune responses in sepsis-related critical illness could be restored ex vivo in cells treatment with monoclonal antibodies targeting either arm of the PD-1:PD-L1 axis [30, 49]. Thus, measuring cellular levels of PD-1 and PD-L1 could inform trial design
CD279	Neutrophil Monocytes CD8 T cells		
HLA-DR	Monocyte	Consistently reported as a marker of immunosuppression in sepsis and in critically ill patients	Reduced HLA-DR expression on monocytes is associated with increased risk of nosocomial infection due to impaired monocyte competence. Monocyte HLA-DR expression less than 8000 monoclonal antibodies/cell for 2 or more days can be reversed with GM-CSF therapy, with potentially beneficial effects [25]. This is a useful biomarker for enrichment in future clinical trials

CD cluster of differentiation antigens; HLA-DR Human Leukocyte Antigen–antigen D Related

reduced neutrophil CD274 and increased total monocyte HLA-DR expression. No biomarkers had significant discriminant value for the outcome of critical care admission or death within 72 h. The performance of individual and optimized combinations of biomarkers for predicting the primary and secondary outcomes are shown in Table 3. No marked non-linearities in biomarker effects were identified. Overall, although statistically significant associations were demonstrated, discrimination of clinical outcomes was unlikely to be clinically useful (Fig. 1).

Extreme phenotype analysis

From 272 patients in cohort-1, we identified 40 “well” and 52 “sick” phenotypes (eFigure-2). “Sick” phenotype patients were characterized by being older, more often male, with a higher frequency of co-morbidities, more frequently lymphopenic, with higher APACHE II and SOFA scores at baseline. After Bonferroni correction for multiple comparisons, “sick” phenotypes had significantly higher monocyte CD279 and neutrophil CD279 in the ED, but no other biomarkers were different (Table 4; eFigure-4).

For both CRP and PCT, there was also no statistically or clinically significant discrimination for subsequent sepsis with univariate analysis (Table 3).

Discussion

In this multi-site cohort study, we reduced a candidate panel of 47 leukocyte biomarkers to 16 reliable biomarkers with potential for discriminating the risk of developing sepsis in patients with suspected infection presenting to the ED. The combination of higher neutrophil CD24, higher neutrophil CD279, and a lower monocyte HLA-DR expression best predicted the clinical deterioration to sepsis. Consistent with this association, a lower neutrophil CD279 expression and higher monocyte HLA-DR expression were associated with discharge home within 72 h (implying rapid recovery). Although our pre-defined biomarker discovery strategy identified these biomarkers as associated with development of sepsis and more severe illness, their discriminant value was insufficient to suggest utility for decision-making in routine clinical care.

Our findings have potential clinical relevance. The key pathophysiological insight is that leukocyte biomarkers of immunosuppression such as check-point inhibitors (CD279; CD274) and antigen processing ability (HLA-DR) were altered even in patients with *suspected infection* presenting to ED. We also demonstrate the importance of assessing reliability when standardising flow cytometry for large-scale time critical use. The development of clinically useable tests is likely to require a form of cross-platform calibration (such as multiparametric version of the Quantibrite system, BD Bioscience). Our study shows

it is feasible to implement flow cytometry as a means of undertaking precision medicine in sepsis, for example to guide novel therapeutic interventions such as those tested recently in immunotherapy trials [15] and highlighted in recent expert reviews [16, 17]. However, our data suggest that for patients with suspected infection the predictive validity of panels of leukocyte biomarkers are unlikely to be useful as general clinical decision-making tools. Of note, both CRP and PCT also performed poorly.

Strengths of our study were well-defined hypothesis, pre-published protocol [8], internationally accepted primary outcome [1], clinically relevant secondary outcomes and hierarchical analytic approach to reduce biomarker selection bias. Reliability of multi-site flow cytometry is potentially problematic due to measurement error bias [18], which we addressed rigorously with fluorochrome-conjugated antibody titrated for optimal signal and kept constant throughout the study. Using hospitalized non-infected patients and ICU-sepsis patients as comparators during biomarker selection increased the chance of detecting infection related host responses and is superior to using healthy volunteer controls. Our blood sampling time point in the ED was prior to severe illness, before major clinical interventions, and much earlier than in previous studies of sepsis biomarkers, and we excluded patients who clinicians considered to have already established sepsis and/or critical illness. As such, our population was different from other recent studies, which evaluated leukocyte biomarkers for prediction of sepsis trajectory (by including patients with sepsis-2 defined sepsis, severe sepsis and septic shock) [19, 20] and stratified nosocomial infection risk in ICU patients [21] (see eTable-5, which highlights important differences). The post hoc extreme phenotype analysis enhanced face validity by considering multiple clinical variables simultaneously for phenotype definition.

Our study has potential weaknesses. Although we could not include all potential leukocyte biomarkers, we studied a range of leukocyte biomarkers (such as complement pathway receptors (CD35, CD11b), G protein-coupled receptors (CD312), Fc-gamma-receptors (CD64 [22, 23]), factors delaying neutrophil apoptosis (CD24 [22]), check-point molecules (CD274, CD279) [24]; HLA-DR expression [25–27]), that previous studies highlight association with adverse outcomes in established sepsis. We enrolled a smaller sample size than planned due to time and funding constraints. However, this had a limited impact since substantial differences in biomarker levels across cohorts still enabled selection of candidate biomarkers for further discriminant analysis. Supervised classification methods such as classification and regression trees (CART) is a valid alternative analytic approach for this research question. However, CART requires approximately 50 events

Table 3 Candidate biomarkers and combinations for predicting outcomes in cohort-1

Biomarker	Marker expression in cohort-1 as Median MFI (IQR)	Primary outcome [OR (95% CI) per SD increase in MFI; <i>p</i> value]		Secondary outcomes ¹ [OR (95% CI) per SD increase in MFI; <i>p</i> value]				
		SOFA score ≥ 2 at 24 h and/or 72 h following presentation to hospital ²	AUROC (95% CI)	ICU admission or death within 72 h of presentation	SOFA ≥4 at 24 or 72 h after presentation	Discharge home within 72 h of presentation	Discharge home within 72 h of presentation or in-hospital with no organ failure	Confirmed infection
Neutrophils								
CD15	31,148 (22,261, 39,622)	0.94 (0.69–1.28); 0.70	0.50 (0.41–0.59)	1.36 (0.82–2.22); 0.23	1.01 (0.65–1.58); 0.97	1.38 (0.99–1.91); 0.06	1.13 (0.83–1.56); 0.42	0.89 (0.57, 1.41); 0.63
CD24	22,261 (16,398, 28,565)	1.20 (0.94–1.54); 0.15	0.56 (0.49–0.63)	1.26 (0.84–1.90); 0.17	1.48 (1.08–2.05); 0.01	1.00 (0.77–1.30); 1.00	0.79 (0.62–1.02); 0.07	1.31 (0.85, 2.04); 0.22
CD35	17,363 (10,021, 26,452)	0.98 (0.77–1.25); 0.87	0.51 (0.44–0.58)	1.18 (0.76–1.83); 0.45	0.90 (0.62–1.31); 0.59	1.18 (0.91–1.53); 0.21	1.15 (0.90–1.47); 0.28	1.34 (0.88, 2.06); 0.17
CD64	2384 (1353, 5522)	0.95 (0.71–1.29); 0.75	0.49 (0.41–0.58)	0.98 (0.55–1.75); 0.94	0.88 (0.53–1.45); 0.61	0.97 (0.70–1.33); 0.83	0.95 (0.71–1.28); 0.74	1.62 (0.84, 3.12); 0.15
CD312	685 (451, 845)	1.29 (0.99–1.67); 0.06	0.57 (0.50–0.64)	0.74 (0.43–1.29); 0.29	0.82 (0.55–1.23); 0.34	0.79 (0.59–1.06); 0.12	0.85 (0.67–1.09); 0.21	1.10 (0.73, 1.67); 0.64
CD11b	20,583 (13,210, 28,737)	1.25 (0.97–1.62); 0.08	0.56 (0.49–0.63)	1.45 (0.97–2.16); 0.07	1.36 (0.98–1.60); 0.57	1.12 (0.86–1.45); 0.39	0.84 (0.66–1.08); 0.18	1.27 (0.83, 1.96); 0.27
CD274	269 (207, 320)	1.25 (0.96–1.61); 0.10	0.57 (0.50–0.64)	0.91 (0.55–1.49); 0.70	1.16 (0.83–1.61); 0.39	0.70 (0.51–0.95); 0.02	0.77 (0.59–0.99); 0.045	1.56 (0.97, 2.52); 0.07
CD279	569 (300, 640)	1.78 (1.23–2.57); 0.002	0.59 (0.52–0.66)	0.96 (0.57–1.61); 0.86	1.06 (0.77–1.46); 0.72	0.57 (0.39–0.83); 0.003	0.60 (0.41–0.87); 0.007	1.06 (0.68, 1.66); 0.78
Monocyte								
CD35	21,018 (13,818, 28,565)	1.15 (0.89–1.48); 0.28	0.55 (0.48–0.62)	0.99 (0.62–1.57); 0.95	1.33 (0.97–1.83); 0.07	0.91 (0.70–1.20); 0.52	0.94 (0.73–1.20); 0.60	1.19 (0.77, 1.84); 0.44
CD64	30,848 (24,499, 39,622)	1.04 (0.77–1.39); 0.80	0.57 (0.49–0.66)	1.25 (0.77–2.03); 0.36	1.12 (0.74–1.71); 0.59	0.95 (0.69–1.30); 0.73	0.92 (0.69–1.24); 0.58	2.24 (1.11, 4.52); 0.02
CD312	1087 (649, 1617)	0.91 (0.71–1.16); 0.43	0.54 (0.47–0.61)	0.73 (0.41–1.29); 0.29	0.79 (0.52–1.21); 0.28	1.24 (0.96–1.61); 0.09	1.06 (0.83–1.36); 0.64	0.94 (0.65, 1.36); 0.73
CD11b	22,705 (14,413, 28,651)	1.21 (0.94–1.57); 0.14	0.58 (0.51–0.65)	1.25 (0.83–1.88); 0.28	1.27 (0.91–1.76); 0.16	1.15 (0.89–1.49); 0.30	0.87 (0.68–1.12); 0.27	1.24 (0.80, 1.93); 0.33
HLA-DR	4435 (2379, 8001)	0.73 (0.55–0.97); 0.03	0.56 (0.49–0.63)	0.69 (0.34–1.40); 0.30	0.76 (0.46–1.24); 0.27	1.35 (1.04–1.75); 0.02	1.34 (1.00–1.80); 0.052	0.96 (0.66, 1.38); 0.82
CD274	60 (0, 166)	0.90 (0.70–1.16); 0.41	0.50 (0.43–0.56)	1.06 (0.69–1.61); 0.80	1.03 (0.73–1.46); 0.85	0.84 (0.62–1.15); 0.28	0.99 (0.78–1.27); 0.95	0.89 (0.64, 1.23); 0.48
CD279	240 (129, 280)	1.32 (1.03–1.70); 0.03	0.58 (0.51–0.65)	0.89 (0.56–1.43); 0.31	1.21 (0.84–1.75); 0.27	0.68 (0.52–0.90); 0.006	0.80 (0.62–1.02); 0.07	0.98 (0.67, 1.44); 0.92
CD8 T cells CD279	117 (72, 169)	1.16 (0.81–1.66); 0.43	0.48 (0.41–0.55)	0.23 (0.02–2.29); 0.21	0.94 (0.58–1.93); 0.80	0.79 (0.43–1.45); 0.45	0.82 (0.55–1.23); 0.34	2.00 (0.44, 9.06); 0.37
Neutrophil CD24 +		1.48 (1.10–1.98); 0.009	0.64 (0.58–0.71)	*	*	1.32 (0.94–1.85); 0.10	0.65 (0.49–0.87); 0.004	*
Neutrophil CD279		2.23 (1.47–3.38); < 0.001	0.67 (0.60–0.74)			0.59 (0.41–0.86); 0.006	0.47 (0.31–0.71); 0.0004	
Neutrophil CD24 +		1.49 (1.10–2.00); 0.009				1.48 (1.03–2.13); 0.04		
Neutrophil CD279 +		2.37 (1.54–3.64); < 0.001						

Table 3 continued

Biomarker	Marker expression in cohort-1 as Median MFI (IQR)	Primary outcome [OR (95% CI) per SD increase in MFI; <i>p</i> value]		Secondary outcomes ¹ [OR (95% CI) per SD increase in MFI; <i>p</i> value]				
		SOFA score ≥ 2 at 24 h and/or 72 h following presentation to hospital ²	AUROC (95% CI)	ICU admission or death within 72 h of presentation	SOFA ≥ 4 at 24 or 72 h after presentation	Discharge home within 72 h of presentation	Discharge home within 72 h of presentation or in-hospital with no organ failure	Confirmed infection
Monocyte HLA-DR		0.72 (0.53–0.97); 0.03						
Neutrophil CD15 +								
Neutrophil CD274 +								
Monocyte HLA-DR								
Other markers#								
CRP		1.20 (0.94–1.54); <i>p</i> =0.15	0.56 (0.49, 0.63)	0.88 (0.55–1.42); 0.60	0.99 (0.69–1.42); 0.94	0.74 (0.55–0.99); 0.04	0.85 (0.66–1.08); 0.19	1.16 (0.99–2.65); 0.06
PCT		0.94 (0.72–1.21); <i>p</i> =0.61	0.53 (0.46, 0.60)	0.93 (0.54–1.61); 0.57	0.81 (0.48–1.36); 0.42	0.83 (0.60–1.15); 0.27	1.02 (0.79–1.32); 0.89	4.00 (0.78–20.5); 0.10

²SOFA score sequential organ failure assessment score; MFI median fluorescence intensity; CD Cluster of differentiation; ICU intensive care unit

* Best subsets regression did not identify any combination models which provided better fit than individual marker models

¹ As pre-specified, secondary outcomes of hospital mortality, occurrence of septic shock and length of stay are not reported

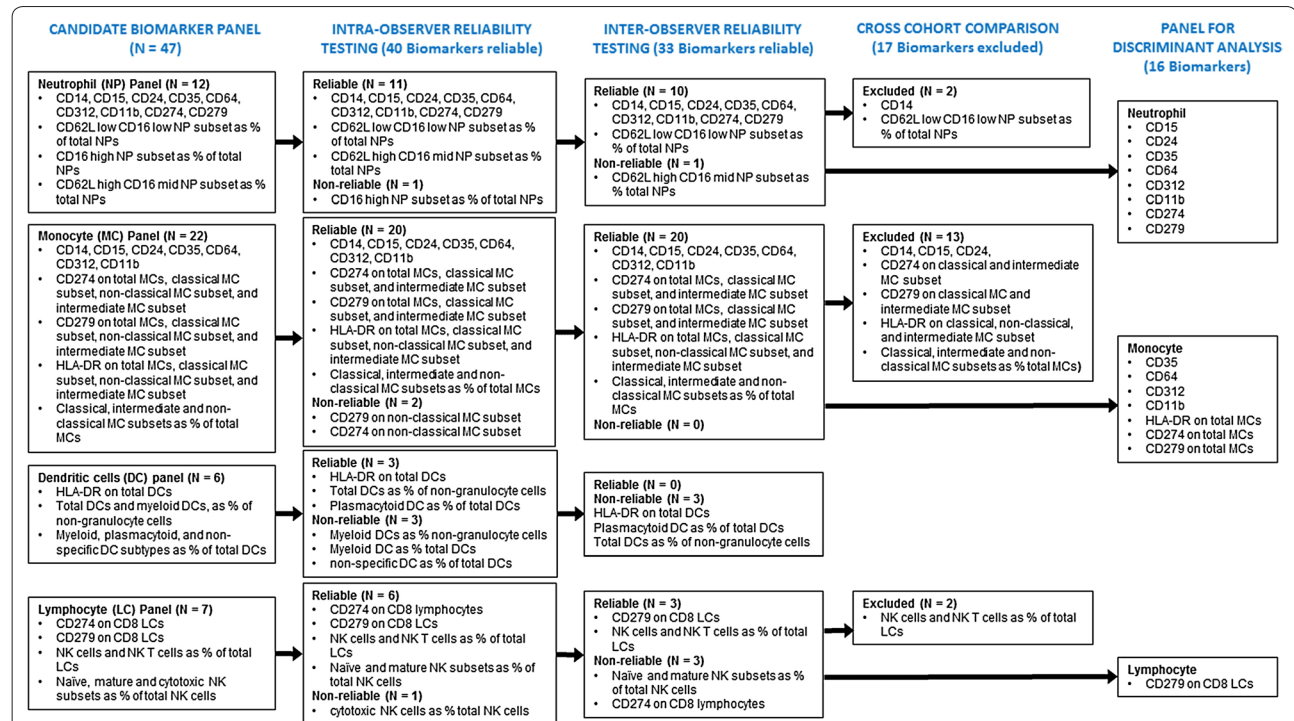


Fig. 1 Overview of selection of leukocyte biomarkers for discriminant analysis through the pre-defined stages of the study. For a detailed description of the rationale for biomarker selection see eMethods-1 and eMethods-2. Non-reliable refers to the analysis of cell populations that are not sufficiently distinct in bimodal FACS plots, are difficult to reliably standardize for a uniform gating approach and need further development. We are proposing that these biomarkers are necessarily of limited value

Table 4 Extreme phenotype description

	Well phenotype (N=40)	Sick phenotype (N=52)	p value
Age, median (IQR)	37.5 (27.3–56.8)	70.0 (56.0–81.0)	<0.001
Female, n (%)	26 (65%)	19 (37%)	0.009
FCI Score Median (IQR)	1 (0–2)	2 (1–3)	0.03
White cell count Median (IQR)			
Total	13.2 (10.3–14.4)	13.1 (9.1–16.5)	0.78
Neutrophils	10.3 (8.1–12.0)	11.2 (7.5–15.1)	0.29
Lymphocytes	1.2 (0.7–1.8)	0.8 (0.5–1.2)	0.01
C-reactive protein Median (IQR)	58.5 (24.0–107.3)	56.0 (16.5–191.0)	0.85
qSOFA score ≥ 2			
At ED presentation	3 (7.5%)	10 (19.2%)	0.11
At 24 h	0	4 (7.7%)	0.07
At 72 h	0	2 (3.8%)	0.22
Source of infection*, n (%)			
Respiratory	13 (40.6%)	30 (57.7%)	0.13
Neurological	1 (3.1%)	2 (3.8%)	0.87
Urinary	2 (6.3%)	7 (13.4%)	0.31
Abdominal	5 (15.6%)	5 (9.6%)	0.41
Skin	9 (28.1%)	3 (5.8%)	0.005
Biliary	0 (0%)	5 (9.6%)	0.005
Sepsis of unknown origin	2 (6.3%)	0 (0%)	0.07
Baseline APACHE 2 score, median (IQR)	4.5 (2–7)	11.5 (9–16)	<0.001
Baseline SOFA, median (IQR)	1 (1–1)	3 (2–4)	<0.001
Discharged home within 72 h, n (%)	32 (80%)	0	<0.001
Admitted to HDU/ICU within 72 h, n (%)	0	14 (26.9%)	<0.001
Neutrophil biomarkers (MFI) median (IQR)			
Neutrophil CD15	30,848 (24,499–45,352)	30,848 (19,116–41,992)	>0.10
Neutrophil CD24	23,815 (18,299–29,261)	24,034 (18,741–30,710)	>0.10
Neutrophil CD35	19,485 (7985–26,580)	15,636 (10,988–25,117)	>0.10
Neutrophil CD64	3098 (1528–6272)	2150 (1693–5378)	>0.10
Neutrophil CD312	565.8 (382.7–712.9)	670.9 (493.6–853.9)	>0.10
Neutrophil CD11b	16,089 (13,664–25,552)	22,154 (13,510–30,737)	>0.10
Neutrophil CD27 s	279.0 (101.4–322.8)	284.3 (233.8–327.7)	>0.10
Neutrophil CD279	326.4 (152.7–584.2)	584.2 (383.7–648.8)	0.005
Monocyte biomarkers (MFI) median (IQR)			
Monocyte CD35	16,556 (9974–27,488)	22,476 (15,067–27,681)	>0.10
Monocyte CD64	29,685 (21,843–45,021)	33,323 (29,405–45,352)	>0.10
Monocyte CD312	1243 (694–2001)	817.0 (470.5–1560.0)	>0.10
Monocyte CD11b	20,205 (12,102–26,644)	26,660 (16,984–32,741)	>0.10
Monocyte CD274	50.7 (0–167.2)	78.6 (0–199.7)	>0.10
Monocyte CD279	151.2 (94.8–262.1)	245.4 (161.1–287.0)	0.05
Monocyte HLA-DR	6172 (3516–11,544)	4016 (2692–7170)	0.12
CD-8 T cell biomarker (MFI) median (IQR)			
CD8 T-Lymphocyte CD279	112.2 (78.7–153.3)	115.6 (58.5–167.9)	>0.10

Categorical variables are given as numbers with percentages. Continuous variables are given as mean with standard deviation where data are parametric, and median with interquartile range otherwise. Comparisons between phenotypes were performed with Fisher exact test between percentages for categorical variables, unpaired t-test for continuous normally-distributed variables, and Mann–Whitney test for other continuous variables

Significant differences are shown in bold (*p*-value of <0.05 taken as significant). For biomarker comparisons, Bonferroni method was used to correct for multiple comparisons and the corrected *p*-values are reported

* For the well phenotype, the denominator for the 'source of infection' variable is 32, as only 32 patients had a final diagnosis of infection. Biomarker comparisons are also reported as dot plots in eFigure-4

per variable when predicting a dichotomised outcome, before predictions become stable and over-optimism is minimised [28]. As our observed number of sepsis events did not reach this threshold we opted to use the best subsets logistic regression approach as pre-specified in our statistical analysis plan [8]. As our cohort-1 inclusion criteria mandated SIRS, we have excluded SIRS negative patients with infection, who could have progressed to develop sepsis. However, this is unlikely to bias the results, as the prevalence of SIRS negative sepsis-3 sepsis in ICUs in England is only 3% [29]. As our objective was to study leukocyte biomarkers at an earlier time point than previously achieved and to identify biomarkers that predict deterioration within 72 h of hospitalisation, we excluded patients planned for direct admission to ICU from the ED at enrolment, which explains the lower than expected event rate for death and septic shock. Findings might be different for more severely ill patients studied later in sepsis, as observed in other recent flowcytometric studies (eTable-5) [19–21].

Our findings have biological plausibility, as the leukocyte biomarkers that best predicted the risk of developing sepsis in our study were on the key innate immune cells, namely neutrophils and monocytes, which are first responders to infection. The strongest biomarker predicting subsequent sepsis and extreme phenotypes was higher levels of CD279 (programmed death receptor 1, PD-1) on monocytes and neutrophils. CD279 expression is associated with neutrophil and monocyte suppressor subsets [30], memory lymphocyte subsets [31], is thought to regulate T cell responses and induce an inhibitory signal characterized by cell cycle arrest and reduced cytokine synthesis [2, 32]. This early role for CD279/PD-1 is consistent with animal models of sepsis [33] and sepsis cohorts [30]. CD279/PD-1 acts in conjunction with its ligand CD274 (PD-L1). In our study, lower CD274, together with lower CD279, higher monocyte HLA-DR, and lower neutrophil CD24, emerged as a predictor for rapid recovery sepsis phenotype. These novel findings require further confirmatory studies.

Although none of the biomarkers we studied had discriminant ability that could be used to guide clinical decision-making, our data imply that immunosuppression in infected patients precedes established sepsis and that higher CD279/PD-1 and lower HLA-DR are potential theragnostic and enrichment markers [34–37] for anti-PD-1/PDL-1 agents and granulocyte-monocyte colony stimulating factor [25], respectively, for carefully designed immunotherapy trials [3, 38].

Conclusions

We conclude that in a population of patients presenting with suspected infection prior to established sepsis, a sequential approach to identifying reliable potential leukocyte biomarkers from a large candidate panel that may predict the subsequent development of sepsis identified only a small number with discriminant properties. These were markers of immune suppression, namely CD279 and HLA-DR, suggesting this may be an early event, prior to development of sepsis.

Electronic supplementary material

The online version of this article (<https://doi.org/10.1007/s00134-018-5389-0>) contains supplementary material, which is available to authorized users.

Author details

¹ School of Immunology & Microbial Sciences, Kings College, London, UK. ² Guy's and St Thomas' NHS Foundation Trust, London SE17EH, UK. ³ MRC Centre for Inflammation Research, University of Edinburgh, 47 Little France Crescent, Edinburgh, UK. ⁴ Centre for Population Health Sciences, Usher Institute, University of Edinburgh, Edinburgh, UK. ⁵ Edinburgh Clinical Trials Unit, University of Edinburgh, Edinburgh, UK. ⁶ Department of Anaesthesia, Critical Care & Pain Medicine, University of Edinburgh, Edinburgh, UK. ⁷ Becton–Dickinson Bioscience, Franklin Lakes, NJ, USA. ⁸ Integrated Critical Care Unit, Sunderland Royal Hospital, Sunderland, UK. ⁹ Emergency Department, Royal Victoria Infirmary, Newcastle Upon Tyne Hospitals NHS Foundation Trust, Newcastle upon Tyne, UK. ¹⁰ Flow Cytometry Core Facility Laboratory, Faculty of Medical Sciences, Centre for Life, Newcastle University, Newcastle upon Tyne, UK. ¹¹ Department of Emergency Medicine, Royal Infirmary of Edinburgh, Edinburgh, UK. ¹² Institute of Cellular Medicine, Newcastle University, Newcastle upon Tyne, UK. ¹³ University Division of Anesthesia, Department of Medicine, Addenbrooke's Hospital, Hills Road, Cambridge, UK.

Acknowledgements

This independent research by Dr. Manu Shankar-Hari is supported by the National Institute for Health Research Clinician Scientist Award (NIHR-CS-2016-16-011). The views expressed in this publication are those of the author(s) and not necessarily those of the NHS, the National Institute for Health Research or the Department of Health. Prof Christopher Weir was supported in this work by NHS Lothian via the Edinburgh Clinical Trials Unit. Dr. Andrew Conway Morris is supported by a Clinical Research Career Development Fellowship from the Wellcome Trust (WT 2055214/Z/16/Z).

Author contributions

Drs. Shankar-Hari, Weir and Walsh had full access to all the data in the study and take responsibility for integrity of data and the accuracy of the data analyses. Concept and design: Walsh, Simpson, Conway Morris, Datta, Weir, Warner. Statistical analysis: Assi, Stephen, Weir, Datta, Wilson, Shankar-Hari. Drafting of manuscript: Shankar-Hari, Weir, Walsh. Acquisition, analysis and interpretation of data: All authors. Critical revision of the manuscript for important intellectual content: All authors. Obtained funding: Walsh, Conway Morris, Brown, Simpson, Warner, Keenan. Administrative, technical, or material support: Walsh, Weir, Warner, Judge, Keenan. Supervision: Walsh, Weir. All authors confirm to the accuracy or integrity of the work.

Funding

The study was funded by Innovate UK (Sepsis 2: 101193). Dr Shankar-Hari is supported by the National Institute for Health Research Clinician Scientist Award (CS-2016-16-011). Dr Conway Morris is supported by a Clinical Research Career Development Fellowship from the Wellcome Trust (WT 2055214/Z/16/Z).

Compliance with ethical standards

Conflicts of interests

Noel Warner, Kevin Judge, Jim Keenen and Alice Wang were all employees of BD biosciences whilst this work was being undertaken, and all four authors hold stock in BD Biosciences. Prof Simpson collaborated with BDB on a Wellcome Trust/Department of Health-funded Healthcare Innovation Challenge Fund (HICF) grant in suspected ventilator-associated pneumonia. He is Director of the NIHR Newcastle In Vitro Diagnostic Evidence Co-operative (formerly the NIHR Newcastle Diagnostic Evidence Co-operative)—these entities exist to evaluate in vitro diagnostics and have worked with (and continue to work with) BDB and other companies in this capacity. All other authors declare that they do have any personal conflict of interest directly related to this manuscript.

Received: 7 August 2018 Accepted: 24 September 2018
Published online: 5 October 2018

References

- Singer M, Deutschman CS, Seymour CW, Shankar-Hari M, Annane D, Bauer M, Bellomo R, Bernard GR, Chiche JD, Cooper-Smith CM, Hotchkiss RS, Levy MM, Marshall JC, Martin GS, Opal SM, Rubenfeld GD, van der Poll T, Vincent JL, Angus DC (2016) The Third International Consensus Definitions for Sepsis and Septic Shock (Sepsis-3). *JAMA* 315:801–810
- Hotchkiss RS, Monneret G, Payen D (2013) Sepsis-induced immunosuppression: from cellular dysfunctions to immunotherapy. *Nat Rev Immunol* 13:862–874
- van der Poll T, van de Veerdonk FL, Scicluna BP, Netea MG (2017) The immunopathology of sepsis and potential therapeutic targets. *Nat Rev Immunol* 17:407–420
- Scicluna BP, van Vught LA, Zwinderman AH, Wiewel MA, Davenport EE, Burnham KL, Nurnberg P, Schultz MJ, Horn J, Cremer OL, Bonten MJ, Hinds CJ, Wong HR, Knight JC, van der Poll T, consortium M (2017) Classification of patients with sepsis according to blood genomic endotype: a prospective cohort study. *Lancet Respir Med* 5:816–826
- Venet F, Guignant C, Monneret G (2011) Flow cytometry developments and perspectives in clinical studies: examples in ICU patients. *Methods Mol Biol* 761:261–275
- Angus DC, Seymour CW, Cooper-Smith CM, Deutschman CS, Klompas M, Levy MM, Martin GS, Osborn TM, Rhee C, Watson RS (2016) A framework for the development and interpretation of different sepsis definitions and clinical criteria. *Crit Care Med* 44:e113–e121
- Shankar-Hari M (2017) How could we enhance translation of sepsis immunology to inform immunomodulation trials in sepsis? *Crit Care* 21:125
- Datta D, Conway Morris A, Antonelli J, Warner N, Brown KA, Wright J, Simpson AJ, Rennie J, Hulme G, Lewis SM, Mare TA, Cookson S, Weir CJ, Dimmick I, Keenan J, Rossi AG, Shankar-Hari M, Walsh TS, Ex PSI (2016) Early PREdiction of Severe Sepsis (ExPRES-Sepsis) study: protocol for an observational derivation study to discover potential leucocyte cell surface biomarkers. *BMJ Open* 6:e011335
- Levy MM, Fink MP, Marshall JC, Abraham E, Angus D, Cook D, Cohen J, Opal SM, Vincent JL, Ramsay G, Sccm/Esicm/Accp/Ats/Sis (2003) 2001 SCCM/ESICM/ACCP/ATS/SIS international sepsis definitions conference. *Crit Care Med* 31:1250–1256
- Shankar-Hari M, Phillips GS, Levy ML, Seymour CW, Liu VX, Deutschman CS, Angus DC, Rubenfeld GD, Singer M, Sepsis Definitions Task F (2016) Developing a new definition and assessing new clinical criteria for septic shock: for the Third International Consensus Definitions for Sepsis and Septic Shock (Sepsis-3). *JAMA* 315:775–787
- Gray A, Ward K, Lees F, Dewar C, Dickie S, McGuffie C, committee Ss (2013) The epidemiology of adults with severe sepsis and septic shock in Scottish emergency departments. *Emerg Med J* 30:397–401
- Glickman SW, Cairns CB, Otero RM, Woods CW, Tsalik EL, Langley RJ, van Velkinburgh JC, Park LP, Glickman LT, Fowler VG Jr, Kingsmore SF, Rivers EP (2010) Disease progression in hemodynamically stable patients presenting to the emergency department with sepsis. *Acad Emerg Med* 17:383–390
- Fluss R, Faraggi D, Reiser B (2005) Estimation of the Youden Index and its associated cutoff point. *Biometrical journal Biometrische Zeitschrift* 47:458–472
- Miller A, Tibshirani R, Cox D, Keiding N, Isham V, Louis T, Tong H, Reid N (2002) Subset selection in regression. Chapman and Hall/CRC, New York
- Francois B, Jeannot R, Daix T, Walton AH, Shotwell MS, Unsinger J, Monneret G, Rimmele T, Blood T, Morre M, Gregoire A, Mayo GA, Blood J, Durum SK, Sherwood ER, Hotchkiss RS (2018) Interleukin-7 restores lymphocytes in septic shock: the IRIS-7 randomized clinical trial. *JCI insight*. <https://doi.org/10.1172/jci.insight.98960>
- Perner A, Gordon AC, Angus DC, Lamontagne F, Machado F, Russell JA, Timsit JF, Marshall JC, Myburgh J, Shankar-Hari M, Singer M (2017) The intensive care medicine research agenda on septic shock. *Intensive Care Med* 43:1294–1305
- Perner A, Rhodes A, Venkatesh B, Angus DC, Martin-Loeches I, Preiser JC, Vincent JL, Marshall J, Reinhart K, Joannidis M, Opal SM (2017) Sepsis: frontiers in supportive care, organisation and research. *Intensive Care Med* 43:496–508
- Mittag A, Tarnok A (2009) Basics of standardization and calibration in cytometry—a review. *J Biophotonics* 2:470–481
- Daix T, Guerin E, Tavernier E, Mercier E, Gissot V, Herault O, Mira JP, Dumas F, Chapuis N, Guitton C, Bene MC, Quenot JP, Tissier C, Guy J, Piton G, Roggy A, Muller G, Legac E, de Prost N, Khellaf M, Wagner-Ballon O, Coudroy R, Dindinaud E, Uhel F, Roussel M, Lafon T, Jeannot R, Vargas F, Fleureau C, Roux M, Allou K, Vignon P, Feuillard J, Francois B, Septiflux Trial G (2018) Multicentric Standardized Flow Cytometry Routine Assessment of Patients With Sepsis to Predict Clinical Worsening. *Chest* 154:617–627
- Guerin E, Orabona M, Raquil MA, Giraudeau B, Bellier R, Gibot S, Bene MC, Lacombe F, Droin N, Solary E, Vignon P, Feuillard J, Francois B (2014) Circulating immature granulocytes with T-cell killing functions predict sepsis deterioration. *Crit Care Med* 42:2007–2018
- Conway Morris A, Datta D, Shankar-Hari M, Stephen J, Weir CJ, Rennie J, Antonelli J, Bateman A, Warner N, Judge K, Keenan J, Wang A, Burpee T, Brown KA, Lewis SM, Mare T, Roy AI, Hulme G, Dimmick I, Rossi AG, Simpson AJ, Walsh TS (2018) Cell-surface signatures of immune dysfunction risk-stratify critically ill patients: INFECTION study. *Intensive Care Med* 44:627–635
- Parlato M, Souza-Fonseca-Guimaraes F, Philippart F, Misset B, Captain Study G, Adib-Conquy M, Cavaillon JM (2014) CD24-triggered caspase-dependent apoptosis via mitochondrial membrane depolarization and reactive oxygen species production of human neutrophils is impaired in sepsis. *J Immunol* 192:2449–2459
- Gros A, Roussel M, Sauvadet E, Gacouin A, Marque S, Chimot L, Lavoue S, Camus C, Fest T, Le Tulzo Y (2012) The sensitivity of neutrophil CD64 expression as a biomarker of bacterial infection is low in critically ill patients. *Intensive Care Med* 38:445–452
- Chang K, Svabek C, Vazquez-Guillamet C, Sato B, Rasche D, Wilson S, Robbins P, Ulbrandt N, Suzich J, Green J, Patera AC, Blair W, Krishnan S, Hotchkiss R (2014) Targeting the programmed cell death 1: programmed cell death ligand 1 pathway reverses T cell exhaustion in patients with sepsis. *Crit Care* 18:R3
- Meisel C, Schefold JC, Pischowski R, Baumann T, Hetzger K, Gregor J, Weber-Carstens S, Hasper D, Keh D, Zuckermann H, Reinke P, Volk HD (2009) Granulocyte-macrophage colony-stimulating factor to reverse sepsis-associated immunosuppression: a double-blind, randomized, placebo-controlled multicenter trial. *Am J Respir Crit Care Med* 180:640–648
- Rangel-Frausto MS, Pittet D, Costigan M, Hwang T, Davis CS, Wenzel RP (1995) The natural history of the systemic inflammatory response syndrome (SIRS). A prospective study. *JAMA* 273:117–123
- Rangel-Frausto MS, Pittet D, Hwang T, Woolson RF, Wenzel RP (1998) The dynamics of disease progression in sepsis: Markov modeling describing the natural history and the likely impact of effective antiseptic agents. *Clin Infect Dis* 27:185–190
- van der Ploeg T, Austin PC, Steyerberg EW (2014) Modern modelling techniques are data hungry: a simulation study for predicting dichotomous endpoints. *BMC Med Res Methodol* 14:137
- Shankar-Hari M, Harrison DA, Rowan KM (2016) Differences in impact of definitional elements on mortality precludes international comparisons of sepsis epidemiology—a Cohort Study illustrating the need for standardized reporting. *Crit Care Med* 44:2223–2230

30. Patera AC, Drewry AM, Chang K, Beiter ER, Osborne D, Hotchkiss RS (2016) Frontline Science: defects in immune function in patients with sepsis are associated with PD-1 or PD-L1 expression and can be restored by antibodies targeting PD-1 or PD-L1. *J Leukoc Biol* 100:1239–1254
31. Wilson JK, Zhao Y, Singer M, Spencer J, Shankar-Hari M (2018) Lymphocyte subset expression and serum concentrations of PD-1/PD-L1 in sepsis—pilot study. *Crit Care* 22:95
32. Boomer JS, To K, Chang KC, Takasu O, Osborne DF, Walton AH, Bricker TL, Jarman SD 2nd, Kreisel D, Krupnick AS, Srivastava A, Swanson PE, Green JM, Hotchkiss RS (2011) Immunosuppression in patients who die of sepsis and multiple organ failure. *JAMA* 306:2594–2605
33. Wang JF, Li JB, Zhao YJ, Yi WJ, Bian JJ, Wan XJ, Zhu KM, Deng XM (2015) Up-regulation of programmed cell death 1 ligand 1 on neutrophils may be involved in sepsis-induced immunosuppression: an animal study and a prospective case-control study. *Anesthesiology* 122:852–863
34. Pene F, Courtine E, Cariou A, Mira JP (2009) Toward theragnostics. *Crit Care Med* 37:S50–S58
35. Shankar-Hari M, Rubenfeld GD (2017) The use of enrichment to reduce statistically indeterminate or negative trials in critical care. *Anaesthesia* 72:560–565
36. Prescott HC, Calfee CS, Thompson BT, Angus DC, Liu VX (2016) Toward smarter lumping and smarter splitting: rethinking strategies for sepsis and acute respiratory distress syndrome clinical trial design. *Am J Respir Crit Care Med* 194:147–155
37. Hotchkiss RS, Sherwood ER (2015) Immunology. Getting sepsis therapy right. *Science* 347:1201–1202
38. Venet F, Monneret G (2018) Advances in the understanding and treatment of sepsis-induced immunosuppression. *Nat Rev Nephrol* 14:121–137
39. Jalava-Karvinen P, Hohenthal U, Laitinen I, Kotilainen P, Rajamaki A, Nikoskelainen J, Lilius EM, Nuutila J (2009) Simultaneous quantitative analysis of Fc gamma RI (CD64) and CR1 (CD35) on neutrophils in distinguishing between bacterial infections, viral infections, and inflammatory diseases. *Clin Immunol* 133:314–323
40. Wang X, Li ZY, Zeng L, Zhang AQ, Pan W, Gu W, Jiang JX (2015) Neutrophil CD64 expression as a diagnostic marker for sepsis in adult patients: a meta-analysis. *Crit Care* 19:245
41. Lewis SM, Treacher DF, Edgeworth J, Mahalingam G, Brown CS, Mare TA, Stacey M, Beale R, Brown KA (2015) Expression of CD11c and EMR2 on neutrophils: potential diagnostic biomarkers for sepsis and systemic inflammation. *Clin Exp Immunol* 182:184–194
42. Netea MG, Balkwill F, Chonchol M, Cominelli F, Donath MY, Giamarellos-Bourboulis EJ, Golenbock D, Gresnigt MS, Heneka MT, Hoffman HM, Hotchkiss R, Joosten LAB, Kastner DL, Korte M, Latz E, Libby P, Mandrup-Poulsen T, Mantovani A, Mills KHG, Nowak KL, O'Neill LA, Pickkers P, van der Poll T, Ridker PM, Schalkwijk J, Schwartz DA, Siegmund B, Steer CJ, Tilg H, van der Meer JWM, van de Veerdonk FL, Dinarello CA (2017) A guiding map for inflammation. *Nat Immunol* 18:826–831
43. Pillay J, Kamp VM, van Hoffen E, Visser T, Tak T, Lammers JW, Ulfman LH, Leenen LP, Pickkers P, Koenderman L (2012) A subset of neutrophils in human systemic inflammation inhibits T cell responses through Mac-1. *J Clin Invest* 122:327–336
44. Pillay J, Tak T, Kamp VM, Koenderman L (2013) Immune suppression by neutrophils and granulocytic myeloid-derived suppressor cells: similarities and differences. *Cell Mol Life Sci* 70:3813–3827
45. Mare TA, Treacher DF, Shankar-Hari M, Beale R, Lewis SM, Chambers DJ, Brown KA (2015) The diagnostic and prognostic significance of monitoring blood levels of immature neutrophils in patients with systemic inflammation. *Crit Care* 19:57
46. Jamsa J, Huotari V, Savolainen ER, Syrjala H, Ala-Kokko T (2015) Kinetics of leukocyte CD11b and CD64 expression in severe sepsis and non-infectious critical care patients. *Acta Anaesthesiol Scand* 59:881–891
47. Brunialti MK, Martins PS, Barbosa de Carvalho H, Machado FR, Barbosa LM, Salomao R (2006) TLR2, TLR4, CD14, CD11B, and CD11C expressions on monocytes surface and cytokine production in patients with sepsis, severe sepsis, and septic shock. *Shock* 25:351–357
48. Wagner C, Kotsougiani D, Pioch M, Prior B, Wentzensen A, Hansch GM (2008) T lymphocytes in acute bacterial infection: increased prevalence of CD11b(+) cells in the peripheral blood and recruitment to the infected site. *Immunology* 125:503–509
49. Demaret J, Venet F, Friggeri A, Cazalis MA, Plassais J, Jallades L, Malcus C, Poitevin-Later F, Textoris J, Lepape A, Monneret G (2015) Marked alterations of neutrophil functions during sepsis-induced immunosuppression. *J Leukoc Biol* 98:1081–1090

UNDERSTANDING THE DISEASE



Non-antiarrhythmic interventions in new onset and paroxysmal sepsis-related atrial fibrillation

Antoine Vieillard-Baron^{1,2,3*} and John Boyd⁴

© 2017 Springer-Verlag GmbH Germany, part of Springer Nature and ESICM

Introduction

Although rarely the primary diagnosis, atrial fibrillation (AF) occurs frequently in critically-ill patients. For instance, in 1341 patients without pre-existing cardiac disease admitted to an intensive care unit, 8.4% exhibited supraventricular arrhythmia, representing AF in 77% of cases [1]. Patients with sepsis appear to be at highest risk; cumulative risk for de novo AF has been reported as 10, 22 and 40% in patients with sepsis, severe sepsis and septic shock, respectively [2]. AF mainly occurs in the first 3 days following admission and is associated with increased ICU-, 30 days-, 90 days-, and 1 year-mortality, even after adjusting for such confounding factors as age, gender, scores of severity, malignancy, and cardiovascular diseases [2]. In 8356 consecutive critically-ill adult patients admitted in medical and surgical ICU, Moss et al. recently reported that de novo AF is associated with increased hospital mortality and length of stay [3].

While it is possible that new onset AF is simply a marker of the overall severity of disease, AF may drive the poor outcome by worsening acute or pre-existing heart failure and by favoring development of thrombo-embolic events such as stroke or myocardial infarction, even though myocardial infarction in this situation is more the consequence of a rate-related oxygen demand–supply imbalance. Side effects of antiarrhythmic drugs (especially beta-blockers and amiodarone) and anticoagulation may also be directly related to morbidity and mortality in medically fragile patients [4], particularly

those with acute or pre-existing heart failure. Due to this unknown benefit/risk balance, there is no consensus on the best treatment for patients who develop new AF but who are not so unstable that they require immediate cardioversion. This is particularly true in patients with sepsis, in which the combination of cardiac failure, microcirculation alterations and changes in vasomotor tone elevates the risks associated with anti-arrhythmic drugs, even compared to other critically-ill patients. Electrical cardioversion is associated with a poor success rate and a very high rate of immediate/early recurrence of AF. Prevention of thrombo-embolic complications of AF with anticoagulation is challenging in the critically ill, as these patients have a high prevalence of disseminated intravascular coagulation and are difficult to maintain in the therapeutic range with unfractionated heparin [5]. In less complex patients with postoperative AF, it is only recommended in hemodynamically unstable patients related to AF to perform a cardioversion, and that rate control is preferred over rhythm control in patients with “acceptable” symptoms [6]. Thus AF is both highly prevalent and associated with death and disability in complex critically ill patients, especially with sepsis. While there is no proven best strategy to treat new onset AF in these patients, we propose in this short piece a simple and systematic approach with a check-list, non-anti-arrhythmic-based, to prevent AF or to reduce AF heart rate. This approach represents the current clinical practice of the authors combined with an analysis of the literature, however it is not based upon high-grade randomized controlled trial evidence.

Pathophysiology of AF

The pathophysiology of AF is complex but may be separated into 2 parts, the cardiac substrate and the trigger.

*Correspondence: antoine.vieillard-baron@aphp.fr

¹ Assistance Publique-Hôpitaux de Paris, Section Thorax-Vascular Disease-Abdomen-Metabolism, Intensive Care Unit, University Hospital Ambroise Paré, 92100 Boulogne-Billancourt, France
Full author information is available at the end of the article

The substrate is for the most part unmodifiable in the ICU during what is typically a very short period of time and is related to left atrium remodeling (dilatation, fatty infiltration, fibrosis, and inflammation), which may favor atrial reentry. Pre-existing risk factors are chronic heart failure, male gender, coronary artery diseases, history of hypertension, left ventricular hypertrophy, diabetes mellitus, aging, obesity (BMI per unit) and excessive alcohol use [7]. Frailty, increased pulse pressure (as a measure of aortic stiffness), chronic kidney disease, valvular heart disease, and sleep apnea have also been reported as risk factors. All these factors are commonly observed in critically-ill patients. The trigger which initiates AF is atrial ectopic discharges, mainly derived from the pulmonary veins. It is this trigger that offers intensivists a therapeutic approach to reduce the incidence and duration of AF. Briefly, atrial ectopic discharges are secondary to diastolic leak of Ca^{2+} from the sarcoplasmic reticulum, leading to early and delayed afterdepolarization. Factors which shorten the refractory period of cardiac myocytes favor AF initiation. It has been reported that metabolic disturbances, as hypokalemia and hyponatremia, may favor de novo AF [8]. Indeed, a low K^+ (< 3.5 mmol/L) or a low Na^+ (< 135 mmol/L) induces delayed afterdepolarization by acting on the $\text{Na}^+/\text{Ca}^{2+}$ exchanger [9]. Another important electrolyte is magnesium (Mg). In more than half of the patients with hypokalemia, hypomagnesemia is associated. Magnesium is known to modulate potassium and calcium channels in the atria and the ventricles and then to have membrane-stabilizing properties. In the Framingham heart study, a low serum Mg level (≤ 1.77 mg/dL) was moderately associated with the development of AF [10] and the infusion of a K-Mg electrolyte solution facilitated electric cardioversion of AF without inducing side effects. Sympathetic activation is another risk factor for atrial ectopic discharges. This may be inherent to the disease, but also due to withdrawal of chronic anti-sympathetic therapies such as beta-blockers and the choice of the vasoactive drug therapy. In the classic report describing stress-induced cardiomyopathy, catecholamine levels were 2–5 fold those of patients with an acute myocardial infarction, equivalent to 3.5 ng/mL norepinephrine, while plasma levels of catecholamines during norepinephrine infusions range from 3 ng/mL up to 170 ng/mL. Clearly a patient with the appropriate cardiac substrate may be triggered into AF with this vasopressor induced chronotropy. Dopamine and epinephrine have the greatest chronotropic effects, both increasing the heart rate by 15% when compared to norepinephrine [11]. Vasopressin in its currently recommended usage, for refractory shock despite norepinephrine, is largely a hormone used as a beta-agonist sparing agent and reduces heart rate by 10%, likely mediated by a 50% reduction in

norepinephrine dose [11]. The synthetic analogues Terlipressin and Selepressin both exert similar 10% reductions in heart rate; Selepressin is currently being studied for use in septic shock (clinical trial identifier NCT02508649). Levosimendan, a calcium sensitizing agent was recently found to exert its main hemodynamic effect via increases in heart rate (more than 10% compared to norepinephrine), with either no change or a decline in stroke volume [12]. Acute hypovolemia has been reported to favor AF [13], probably by inducing sympathetic stimulus. Given that left atrial (LA) diameter > 5 cm measured by echocardiography is the strongest predictor of chronic AF [14], it is likely (though unproven to date) that over-prescription of fluid and fluid overload lead to LA dilation and increases the probability of new AF. Systemic inflammation, through inflammatory mediators (IL-6, IL-1, CRP, TNF- α) is another risk factor for AF. One may assume that fever control could limit AF. However, no data currently support such a hypothesis and in the randomized controlled trial by Schortgen et al. on the effect of fever control in septic patients, no information is given about the incidence of AF [15]. More, such an approach is probably not a benign intervention in septic patients and requires more investigation. Finally, it is suspected that pericardial effusion, as well as mal-positioned central venous catheter into the right atrium, might favor de novo AF. In cases of de novo AF, treating these factors, especially fever and sympathetic activation, may help control heart rate. Gillinov et al. have reported no difference for length of stay, 60 days-mortality and sinus rhythm between rate control targeting 90–110 beats per minute versus rhythm control groups in patients after cardiac surgery [16], although in a different population than medically complex critically ill patients.

Non-antiarrhythmic interventions

Based on the description above of AF pathophysiology and of the main risk factors for atrial ectopic discharges, we can propose for stable patients, i.e. those not requiring urgent electrical cardioversion, a list that has to be checked before discussing any anti-arrhythmic drugs (Table 1). Briefly, electrolytes have to be rigorously normalized, focusing on potassium, sodium and magnesium. Fluid status must be optimized, avoiding hypovolemia but also fluid overload, with a particularly conservative fluid administration if hemodynamic monitoring reveals a LA size enlargement. When selecting vasoactive drugs, avoid highly active chronotropes such as epinephrine and dopamine, and reduce the overall dose of adrenergic agent by choosing the lowest acceptable blood pressure target according to organ perfusion and history of chronic kidney disease. Strategies such as early vasopressin infusion to reduce norepinephrine dose could

Table 1 A standardized approach to prevent new atrial fibrillation in critically ill patients

Modifiable Risk Factor	Diagnosis	Strategy to avoid AF
Hypovolemia ^a	Tachycardia, hypotension, with low central venous pressure or positive dynamic parameters of fluid-responsiveness	Fluid resuscitation targeting parameters of fluid-responsiveness if the patient is still hypotensive
Volume overload ^a	Peripheral and/or pulmonary edema. Echocardiography with LA diameter > 5 cm ^b	Discontinue fluids in favor of non-chronotropic vasopressors. Try limiting central venous pressure below 8 mmHg
Potassium < 3.5 mmol Magnesium < 1.77 mg/dL Sodium < 135 mmol	Electrolyte screen	Concurrent replacement of potassium and magnesium. Avoid hypotonic solutions in initial resuscitation and consider correcting hyponatremia, if any
Excess chronotropy due to vasopressors	Recognition of high chronotropic potency of Epinephrine, dopamine and levosimendan	Avoid Epinephrine, dopamine and levosimendan in high risk patients. Vasopressin or synthetic analogue co-administration with noradrenaline in patients at high risk of AF could be discussed
Pericardial effusion	Echocardiography	Drain if hemodynamically significant
Mal-positioned central venous Catheter	Chest X-ray or ultrasonography	Withdraw catheter to caval-atrial junction
Persistent fever (> 38°) despite adequate antibiotics	Routine vital sign monitoring	Check for any unexpected cause or complication (superinfection). External cooling and anti-pyretics could be discussed

LA left atrium

^a As all the recommendations regarding fluid management, some of these aspects may be subject to controversy^b LA size reflects the cumulative effects of increased LV filling pressures over time. In other words, acute increase in LV filling pressures may exist without any LA dilatation

be discussed in patients at highest risk of AF. Echocardiography may be performed to look for LA diameter, a pericardial effusion that could be drained if significant and a chest X-ray (or an ultrasonography) to control the position of the central venous catheter, if any (a catheter too far into the right atrium must be slightly removed). Finally, reasons for persistent fever have to be investigated and external cooling or antipyretic medication could favor AF reduction, even though no data currently support this assumption.

Conclusion

Atrial fibrillation is frequent in critically-ill patients and its incidence increases in sepsis and septic shock. Despite a well-demonstrated association with length of stay and mortality, the need for a specific treatment, as well as the type of this treatment, cannot be recommended. Based on the pathophysiology of AF and its risk factors, mainly present in septic patients, we propose a systematic and simple non-antiarrhythmic-based approach to reduce AF, focusing on optimization of electrolytes and fluid status, limitation of sympathetic activation, whatever its cause (inflammation, fever, vasoactive drugs), and control of the central venous catheter position.

Author details

¹ Assistance Publique-Hôpitaux de Paris, Section Thorax-Vascular Disease-Abdomen-Metabolism, Intensive Care Unit, University Hospital Ambroise Paré, 92100 Boulogne-Billancourt, France. ² Faculty of Medicine, University of Versailles Saint-Quentin en Yvelines, Paris Ile-de-France Ouest, 78280 Saint-Quentin en Yvelines, France. ³ INSERM U-1018, CESP Team 5 (EpReC Renal and Cardiovascular Epidemiology), UVSQ, 94807 Villejuif, France. ⁴ Centre for Heart Lung Innovation, University of British Columbia, Vancouver, BC, Canada.

Compliance with ethical standards

Conflicts of interest

The authors declare that they have no conflict of interest.

Received: 13 September 2017 Accepted: 31 October 2017

Published online: 7 November 2017

References

1. Annane D, Sébille V, Duboc D, Le Heuzey JY, Sadoul N, Bouvier E, Belissant E (2008) Incidence and prognosis of sustained arrhythmias in critically ill patients. *Am J Respir Crit Care Med* 178:20–25
2. Klein Klouwenberg P, Frencken J, Kuipers S, Ong D, Peelen L, van Vught L, Schultz M, van der Poll T, Bonten M, Cremer O, On behalf of the MARS consortium (2017) Incidence, predictors, and outcomes of new-onset atrial fibrillation in critically ill patients with sepsis. *Am J Respir Crit Care Med* 195:205–211
3. Moss T, Forrest Calland J, Enfield K, Gomez-Manjarres D, Ruminski C, DiMarco J, Lake D, Moorman R (2017) New-onset atrial fibrillation in the critically ill. *Crit Care Med* 45:790–797
4. Walkey A, Hogarth K, Lip G (2015) Optimizing atrial fibrillation management. From ICU and beyond. *Chest* 148:859–864

5. Ghassemi M, Richter S, Eche I, Chen T, Danziger J, Celi L (2014) A data-driven approach to optimized medication dosing: a focus on heparin. *Intensive Care Med* 40:1332–1339
6. The Task Force for the management of atrial fibrillation of the European Society of cardiology (ESC) (2016) 2016 ESC guidelines for the management of atrial fibrillation developed in collaboration with EACTS. *Eur Heart J* 37:2893–2962
7. Lau D, Nattel S, Kalman J, Sanders P (2017) Modifiable risk factors and atrial fibrillation. *Circulation* 136:583–596
8. Krijthe B, Heeringa J, Kors J, Hofman A, Franco O, Witteman J, Stricker B (2013) Serum potassium levels and risk of atrial fibrillation. The Rotterdam study. *Int J Cardiol* 168:5411–5415
9. Lu YY, Cheng CC, Chen YC, Lin YK, Chen SA, Chen YJ (2016) Electrolytes disturbances differentially regulate sinoatrial node and pulmonary vein activity: a contribution to hypokalemia-or hyponatremia-induced atrial fibrillation. *Heart Rhythm* 13:781–788
10. May Khan A, Lubitz S, Sullivan L, Sun J, Levy D, Vasan R, Magnani J, Ellinor P, Benjamin E, Wang T (2013) Low serum magnesium and the development of atrial fibrillation in the community. The Framingham heart study. *Circulation* 127:33–38
11. Rhodes A, Evans L, Alhazzani W, Levy M, Antonelli M, Ferrer F et al (2017) Surviving sepsis campaign: international guidelines for management of sepsis and septic shock: 2016. *Intensive Care Med* 43:304–377
12. Gordon AC, Perkins GD, Singer M, McAuley DF, Orme RM, Santhakumaran S, Mason AJ, Cross M, Al-Beidh F, Best-Lane J, Brealey D, Nutt CL, McNamie JJ, Reschreiter H, Breen A, Liu KD, Ashby D (2016) Levosimendan for the prevention of acute organ dysfunction in sepsis. *N Engl J Med* 375:1638–1648
13. Edwards JD, Wilkins RG (1987) Atrial fibrillation precipitated by acute hypovolemia. *Br Med J* 294:283–284
14. Psaty BM, Manolio TA, Kuller LH, Kronmal RA, Cushman M, Fried LP, White R, Furberg CD, Rautaharju PM (1997) Incidence of and risk factors for atrial fibrillation in older adults. *Circulation* 96:2455–2461
15. Schortgen F, Clabault K, Katsahian S, Devaquet J, Mercat A, Deye N, Dellamonica J, Bouadma L, Cook F, Beji O, Brun-Buisson C, Lemaire F, Brochard L (2012) Fever control using external cooling in septic shock. A randomized controlled trial. *Am J Respir Crit Care Med* 185:1088–1095
16. Gillinov A, Bagiella E, Moskowitz A, Raiten J, Groh M, Bowditch M et al (2016) Rate control versus rhythm control for atrial fibrillation after cardiac surgery. *N Engl J Med* 374:1911–1921

EDITORIAL

Using multiple 'omics strategies for novel therapies in sepsis

James A. Russell^{1*}, Peter Spronk² and Keith R. Walley^{1,2}

© 2018 Springer-Verlag GmbH Germany, part of Springer Nature and ESICM

Sepsis is life-threatening organ dysfunction caused by dysregulated host response to infection [1]. Treatment is complicated because sepsis is heterogeneous, explaining the lack of effective drugs. Sepsis treatment includes broad-spectrum antibiotics, vasopressors, ventilation and dialysis. A limitation of antibiotics is that they do not directly remove the bacterial endotoxins and exotoxins, which may cause organ failure.

Exotoxins are potent immunologic stimulators—very low concentrations stimulate deleterious immunologic responses. Most exotoxins function as if they are superantigens [that bind to T cell receptors, activate T cells (especially T helper cells) and stimulate cytokine release].

Blocking endotoxin effects by blocking the TLR4 receptor with eritoran (a TLR4 blocker) was unsuccessful in severe sepsis [2]. This pivotal trial may have been negative because the timing was inadequate, the patients had severe sepsis rather than just septic shock, patients were not sick enough (placebo mortality 56% in the prior phase II trial but only 27% in the pivotal trial), eritoran may have worsened outcomes in gram-positive sepsis (46% of patients) (mortality rates: eritoran 34% vs. placebo 25%) or other causes.

There are no novel drugs available to treat sepsis. We propose a new drug discovery strategy that focuses on (1) the early infectious stage, (2) multiple 'omics and (3) an inverted drug discovery sequence to increase the chances of success.

Why focus on early sepsis?

Prior drug discoveries in sepsis that focused on the host inflammatory responses failed. Early antibiotics remain

the only effective treatment [3], so we focus on the early infectious phase. “Early” is difficult to define for sepsis because determining ‘time-zero’ in human sepsis is impossible. Herein, we define early as inclusion within the first 24 h after emergency department arrival.

Antibiotics are recommended within 1 h of presentation [4] because each 1-h delay is associated with 4–6% decreased survival [5]. However, antibiotics do not directly remove bacterial endotoxins that stimulate immune, inflammatory, apoptotic and coagulation pathways causing organ failure and death [6].

Why multi-'omics?

Most sepsis drugs were developed by understanding the disease mechanism and targeting a relevant pathway. An 'omics association is typically an unbiased discovery that points to a possible mechanistic pathway. We define multi-'omics as measurement and examination of associations of at least two types of 'omics variables, from genomics, lipidomics, proteomics to metabolomics. Multi-'omics confirmation refines mechanistic understanding so that high probability drug targets can be identified.

Death due to infection is more heritable than death due to cancer or heart disease [7]. More recently, it has been proposed that environmental influences in early life may override genetic influences [8]. However, there is great value in evaluating the associations of genetic variations with impaired endotoxin clearance, organ dysfunction and death to facilitate drug discovery.

As an example, the endotoxin clearance cascade is a strong candidate pathway for study. Variation of endotoxin clearance cascade genes could alter endotoxin clearance, inflammation, bacterial load and survival. Key aspects of endotoxin cascade neutralization include binding to HDL, modulation by proprotein convertase subtilisin/kexin type 9 (PCSK9), transfer to LDL, LDL/

*Correspondence: Jim.Russell@hli.ubc.ca

¹ Division of Critical Care Medicine, Centre for Heart Lung Innovation, St. Paul's Hospital, 1081 Burrard Street, Vancouver, BC V6Z 1Y6, Canada
Full author information is available at the end of the article

endotoxin clearance via the hepatic LDL receptor, VLDL binding of endotoxin and the role of VLDL receptors in adipose tissue and transfer proteins (e.g., cholesterylester transfer protein).

Using a candidate gene approach, we discovered that PCSK9 inhibition acts as a broad-spectrum adjunct to all antibiotics in severe infection. We evaluated *PCSK9* because PCSK9 inhibitors were developed to lower cholesterol [9–12] and because endotoxins are lipid rich. LPS bound to LDL is cleared via hepatic LDL receptors and then excretion in bile. PCSK9 impedes LPS clearance by decreasing LDL receptor density [9]. Septic patients with *PCSK9* loss-of-function (LOF) genotypes have higher survival and lower plasma cytokine concentrations than wild type and patients carrying gain-of-function polymorphisms (GOF) [9].

The most common single-nucleotide polymorphisms of PCSK9 [13] are missense LOF variants rs11591147 (R46L), rs11583680 (A53V) and rs562556 (V474I); the most common missense GOF variant is rs505151 (G670E). The minor allele frequencies in sepsis patients are: rs11591147: 0.6–1.2%, rs11583680: 11–13%, rs562556: 16–17% and rs505151: 4–5% [9] similar to the general population. These PCSK9 mutations are pleiotropic [14]; the degree of cardiovascular protection is greater than expected by the LDL reduction perhaps because of other aspects of lipoprotein metabolism, inflammation, thrombosis, immune function (anti-viral and -malarial properties) and PCSK9 function in non-hepatic tissues.

Why an inverted drug discovery sequence?

Previous sepsis drugs arose from classic drug discovery: researchers identified mechanism(s) of sepsis in animal models and then did trials in humans. This strategy does not account for genetic heterogeneity of microorganisms and the host. We propose inverting (as in our PCSK9 discovery) the standard drug discovery sequence by starting with human 'omics, confirm mechanisms in models and then make go/no-go decisions for potential targets for clinical development.

One could extend our PCSK9 genomics-based approach, by adding multi-'omics to discover other novel targets. First, sequence genes of a relevant pathway (e.g., 32 endotoxin clearance cascade genes) and determine associations with 28-day survival. Then, measure multi-'omics in the same sepsis cohorts to determine associations of variants with multi-'omics in those cohorts. Next, examine associations of gene variants with multi-'omics in human volunteers administered low-dose lipopolysaccharide to select candidate targets meeting three criteria: variants with (1) significantly decreased survival, (2) significantly different level(s) of multi-'omics and (3) significantly different multi-'omics in the human lipopolysaccharide infusion cohort. Selected candidate targets would be evaluated for mechanisms in (1) human hepatocytes (because the liver clears endotoxins) and (2) murine gene knock-out models (e.g., peritonitis). Targets with mechanisms of action are taken to drug synthesis (antibody and small molecules). We did such a feasibility study of multi-'omics in 24 septic shock patients and 99 healthy controls and found significantly lower levels of

Table 1 Associations of PCSK9 genotype (wild type vs. loss of function) with protein, lipid and metabolite concentrations in patients with septic shock (n = 24)

Metabolites and lipids	PCSK9 genotype wild type (n = 13)	PCSK9 genotype LOF (n = 11)	p
Citrulline	18.5 (15.9–21.3)	21.9 (20.0–24.1)	0.026
Glutamic acid	37.3 (24.5–44.7)	38.9 (35.4–54.7)	0.043
Lysophosphatidylcholine C18:2	2.2 (1.5–7.3)	1.1 (0.9–3.2)	0.046
Ornithine	59.1 (44.8–72.7)	104.3 (68.3–116.6)	0.010
Phenylalanine	73.7 (64.9–81.2)	86.9 (77.0–96.8)	0.009
Phosphatidylcholine acyl-alkyl C30:1	0.03 (0.02–0.07)	0.06 (0.04–0.12)	0.042
Phosphatidylcholine diacyl C42:5	0.20 (0.17–0.25)	0.16 (0.12–0.18)	0.042
Trans-OH-proline	6.5 (5.3–9.1)	10.8 (8.4–18.3)	0.016
Proteins			
Apolipoprotein A-IV	41.1 (31.0–54.5)	60.3 (46.1–109.7)	0.046
Apolipoprotein B-100	694 (445–830)	354 (231–523)	0.003
Coagulation factor V	7.8 (6.0–8.5)	4.4 (3.7–7.0)	0.002
Complement component C7	38.0 (27.8–52.9)	62.1 (52.0–79.3)	0.004
IgGfC-binding protein	10.0 (8.3–12.4)	27.7 (11.0–42.7)	0.019
Serotransferrin	784 (618–1196)	1293 (878–1611)	0.025
Thyroxine-binding globulin	3.1 (2.5–4.0)	2.5 (2.0–2.8)	0.026

proteins, lipids and metabolites compared with controls (Genga KR 2018). We evaluated PCSK9 gene variants and found significant differences in proteins, lipids and metabolites between PCSK9 loss-of-function and wild-type patients (Table 1; supplement text).

In summary, focus on early sepsis, harnessing the power of multi-omics and inverting the drug discovery sequence could enhance drug discovery in sepsis.

Electronic supplementary material

The online version of this article (<https://doi.org/10.1007/s00134-018-5122-z>) contains supplementary material, which is available to authorized users.

Author details

¹ Division of Critical Care Medicine, Centre for Heart Lung Innovation, St. Paul's Hospital, 1081 Burrard Street, Vancouver, BC V6Z 1Y6, Canada. ² Department of ICU, Gelre Hospitals Apeldoorn, Apeldoorn, The Netherlands.

Received: 9 February 2018 Accepted: 1 March 2018

Published online: 15 March 2018

References

1. Singer M, Deutschman CS, Seymour CW et al (2016) The third international consensus definitions for sepsis and septic shock (sepsis-3). *JAMA* 315:801–810
2. Opal SM, Laterre PF, Francois B et al (2013) Effect of eritoran, an antagonist of MD2-TLR4, on mortality in patients with severe sepsis: the ACCESS randomized trial. *JAMA* 309:1154–1162
3. Seymour CW, Gesten F, Prescott HC et al (2017) Time to treatment and mortality during mandated emergency care for sepsis. *N Engl J Med* 376:2235–2244
4. Rhodes A, Evans LE, Alhazzani W et al (2017) Surviving sepsis campaign: international guidelines for management of sepsis and septic shock: 2016. *Crit Care Med* 45:486–552
5. Kumar A (2010) Early antimicrobial therapy in severe sepsis and septic shock. *Curr Infect Dis Rep* 12:336–344
6. Angus DC, van der Poll T (2013) Severe sepsis and septic shock. *N Engl J Med* 369:840–851
7. Sorensen TI, Nielsen GG, Andersen PK, Teasdale TW (1988) Genetic and environmental influences on premature death in adult adoptees. *N Engl J Med* 318:727–732
8. Petersen L, Sorensen TI, Andersen PK (2010) A shared frailty model for case-cohort samples: parent and offspring relations in an adoption study. *Stat Med* 29:924–931
9. Walley KR, Thain KR, Russell JA et al (2014) PCSK9 is a critical regulator of the innate immune response and septic shock outcome. *Sci Transl Med* 6:258
10. Roth EM, McKenney JM, Hanotin C, Asset G, Stein EA (2012) Atorvastatin with or without an antibody to PCSK9 in primary hypercholesterolemia. *N Engl J Med* 367:1891–1900
11. Seidah NG, Abifadel M, Prost S, Boileau C, Prat A (2017) The proprotein convertases in hypercholesterolemia and cardiovascular diseases: emphasis on proprotein convertase subtilisin/kexin 9. *Pharmacol Rev* 69:33–52
12. Stein EA, Mellis S, Yancopoulos GD et al (2012) Effect of a monoclonal antibody to PCSK9 on LDL cholesterol. *N Engl J Med* 366:1108–1118
13. Abifadel M, Rabes JP, Devillers M et al (2009) Mutations and polymorphisms in the proprotein convertase subtilisin kexin 9 (PCSK9) gene in cholesterol metabolism and disease. *Hum Mutat* 30:520–529
14. Stower H (2011) Human genetics: pleiotropic mutations. *Nat Rev Genet* 13:5

In Severe Sepsis and Acute Pancreatitis

Only
Rs. 1399/-



Rx

Ulin4fit

Inj. Ulinastatin 1,00,000 IU

Ultimate fit between Expectancy & cost

Reduces Mortality

Prevents new onset of organ dysfunction

Faster recovery

Fewer adverse reactions



Website: www.gufic.com • Email: info@guficbio.com

